



This is the peer reviewed version of the following article: *Inmunología*. 2009, 28(1):32-45, which has been published in final form at [https://doi.org/10.1016/S0213-9626\(09\)70025-3](https://doi.org/10.1016/S0213-9626(09)70025-3)

This article may be used for non-commercial purposes in accordance with Elsevier Terms and Conditions for Use of Self-Archived Versions.

# ACEPTADO

## Th17 lineage: answers to some immunological questions

C. González-García, F.M. Martín-Saavedra, A. Ballester and S. Ballester

Unidad de Regulación Génica, Centro Nacional de Microbiología, Instituto de Salud Carlos III, Majadahonda, Madrid.

### RESUMEN

En los últimos años se han estudiado exhaustivamente las funciones y las rutas de desarrollo del subtipo de células T helper especializado en la producción de IL-17 (Th17). Este linaje celular de células efectoras desempeña un papel decisivo tanto en la respuesta inmune a agentes infecciosos, como en inmunopatologías. Al igual que para los subtipos Th1 y Th2, la definición de Th17 está dirigida por citoquinas y factores de transcripción específicos. La combinación de TGF- $\beta$  e IL-6, y los factores de transcripción ROR $\gamma$ t, ROR $\alpha$  y Stat3 son esenciales para comprometer el subtipo Th17. IL-23 juega un papel clave en la estabilización del fenotipo y de la actividad patogénica de células productoras de IL-17. La citoquina IL-21 producida por células Th17 participa en un mecanismo de retroalimentación para favorecer el desarrollo de células productoras de IL-17; mientras que las citoquinas IL-27, IL-4, IFN $\gamma$ , IL-25 e IL-2 limitan el fenotipo Th17. Células T reguladoras CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> (Treg) siguen una ruta de desarrollo divergente al establecimiento de IL-17, aunque ambas alternativas son gobernadas por TGF- $\beta$ , el cual dirige el destino de células CD4<sup>+</sup> *naïve* hacia uno u otro de estos subtipos celulares mutuamente excluyentes dependiendo de la presencia de IL-6. Además, datos recientes indican que células Treg ya establecidas pueden modificar su programa genético para convertirse en células Th17. En esta revisión se resumen y analizan los datos disponibles actualmente acerca de la biología de células Th17.

### SUMMARY

In recent years the function and developmental pathway for the T helper subset specialized in IL-17 production (Th17) have been exhaustively studied. This lineage of effector cells plays a decisive role in the immune response to infectious agents, as well as in immunopathologies. Similar to Th1 and Th2 subsets, Th17 definition is orchestrated by specific cytokines and transcription factors. A combination of TGF- $\beta$  plus IL-6, and the transcription factors ROR $\gamma$ t, ROR $\alpha$  and Stat3 are essential for Th17 commitment. IL-23 plays a key role in the stabilization of the phenotype and in the promotion of the pathogenic activity of IL-17 producer cells. IL-21 cytokine produced by Th17 cells participates in a feedback mechanism to favour this phenotype; while IL-27, IL-4, IFN $\gamma$ , IL-25 and IL-2 cytokines limit the Th17 response. CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> regulator cells (Treg) follow a development pathway divergent to Th17 establishment, although both alternatives are governed by TGF- $\beta$  that directs the fate of naïve CD4<sup>+</sup> cells to each of these mutually exclusive T cell subsets depending on the presence of IL-6. Furthermore, recent data indicate that pre-established Treg cells can switch its genetic program to become IL-17 producer cells. In this review we summarize and discuss the current available data about the biology of Th17 cells.

**Key words:** Th17, TGF- $\beta$ , IL-6, IL-23, ROR $\gamma$ t.

**Corresponding author:** Sara Ballester, Unidad de Regulación Génica, Centro Nacional de Microbiología, Instituto de Salud Carlos III, Ctra. Majadahonda-Pozuelo Km2, 28220, Madrid. Tfno: 918223922, e-mail: sballes@isciii.es.

# ACEPTADO

## IL-17 FAMILY

Since Mossman *et al.* proposed the model wherein CD4<sup>+</sup> T helper cells were classified in Th1 and Th2 subtypes with different functions in immune responses according to the profile of cytokines produced (1), the Th1/Th2 paradigm provided valuable tools for understand the interplay of innate and adaptive immunity and the CD4<sup>+</sup> T cell function. However, there have arisen some discrepancies related to results that did not fit in the Th1/Th2 hypothesis. During the past years new studies have identified a new subset of CD4<sup>+</sup> T cells that secrete IL-17 and the closely related cytokine IL-17F as well as other inflammatory cytokines such as IL-6 and IL-22 (2, 3).

Murine IL-17 was described as CTLA-8 (4), and a 63% homologous human cytokine was soon found (5). Currently the IL-17 cytokines include a family of six members (IL-17A-F), at least two of which have potent proinflammatory properties: IL-17A or CTLA-8 (the founder member of the family also named IL-17), and IL-17F. Both are produced by the recently described Th17 cell subset, are localized at the same chromosomal locus (1A4), share a 55% of homology at the protein level, and seem to have similar functions. IL-17A and IL-17F work mostly as homodimers, but IL-17A/F heterodimers have been recently described in several independent reports (6-8), suggesting a role in inflammatory response regulation for such IL-17 complexes. IL-17D and IL-17E, whose respective alternative names are IL-27 and IL-25, are the two members of the IL-17 family with lowest homology (16% at protein level) to IL-17A. None of them is produced by Th17 cells, and both of them, as discussed later, exert a negative control on the Th17 subset development.

IL-17 receptors are a family of five members of membrane proteins, IL-17RA-F, reviewed in (9). Except IL-17RA, each of these receptors has alternative splicing variants, that for IL-17RB and IL-17RC result in secreted soluble proteins, which could serve to antagonize their ligands (10). IL-17A and IL-17F bind to IL-17RA, although IL-17A binds with much higher affinity (11), which correlates with its greater potency in functions as induction of chemokine expression (8). In addition to IL-17RA, IL-17F can also bind to IL-17RC. By signaling through IL-17RA, which is ubiquitously expressed, IL-17 can induce the production of different kind of proteins, many of them related to inflammation, as chemokines (CXCL-1, CXCL-2, CXCL-8-10, CCL-2, CCL20), cytokines (IL-6, TNF $\alpha$ , G-CSF, GM-CSF), proteins of acute phase response, tissue remodelling factors (MMP1, MMP3, MMP9, MMP13, TIMP2), and anti-microbial products ( $\beta$ -defensins, mucins, calgranulins) (12). Of main importance in the signal transduction of IL-17 seems to be NF $\kappa$ B and C/EBP

transcription factors, with the involvement of MAP kinase pathways (13-16). Moreover, TRAF6 was shown to be involved in the activation of NF $\kappa$ B by IL-17 (17). On the other hand, IL-17 can increase expression of some of its target genes through mRNA stabilization (18).

IL-17B and IL-17C are members of the family whose cellular sources are unknown yet, and whose biology seems to be no related to IL-17A. In this review, we will refer to IL-17A as IL-17.

## CLUES TO IDENTIFICATION OF A NEW T HELPER CELL LINEAGE PRODUCING IL-17

The first suggestion of a new T helper subset, distinct from Th1 and Th2, was provided by the finding of T CD4<sup>+</sup> cells producing high levels of IL-17 without expression of IFN $\gamma$  or IL-4, the respective prototype cytokines produced by Th1 and Th2 (19). Other important clues were supplied by studies on animal models of autoimmunity. Pathologies as experimental autoimmune encephalomyelitis (EAE) and collagen-induced arthritis (CIA), mouse models for human multiple sclerosis (MS) and rheumatoid arthritis (RA) respectively, have been traditionally considered as Th1-mediated diseases. However, during years, several experimental results showed inconsistencies with the Th1/Th2 hypothesis. For example, depletion of the Th1 cytokine IFN $\gamma$ , its receptor IFN $\gamma$ R, or the receptor of IL-12 (IL-12R $\beta$ 2), the master inducer of Th1 phenotype, increased the susceptibility to EAE (20-26).

With the discovery of IL-23 as an heterodimeric cytokine that shares its p40 subunit with IL-12 (27), could arise the study of the relative contribution of IL-12 and IL-23 to chronic inflammation. IL-12 heterodimer is composed by the p40 and p35 subunits, while IL-23 comprises p40 and a different p19 protein. By using mice lacking IL-23 (*p19*<sup>-/-</sup>), IL-12 (*p35*<sup>-/-</sup>) or both cytokines (*p40*<sup>-/-</sup>), it could be demonstrated that IL-23, and not IL-12, is the critical cytokine for autoimmune inflammation of central nervous system (CNS) during EAE (28). This study was followed by many other evidences about the role of IL-23 in inflammation (29-31). In parallel with these findings, it was reported that IL-17-producer cells can be generated independently of the cytokines and transcription factors required for Th1 or Th2 subtypes differentiation. The development of Th17 from *naïve* cells was potently inhibited by IFN $\gamma$  and IL-4, whereas memory Th17 cells were resistant to suppression by Th1 or Th2 cytokines, indicating that the phenotype of these cells had been permanently committed. All these data together allowed to establish that a T helper lineage distinct from Th1 and Th2 can differentiate from naïve CD4<sup>+</sup> cells to produce IL-17 (2, 3).

In 2006, three independent reports provided the basis for understanding the process of Th17 cell promotion (32-34). All of them found that transforming growth factor  $\beta$  (TGF- $\beta$ ) is critical while IL-23 is not required for Th17 commitment. TGF- $\beta$  is associated with immunosuppressive functions through induction of Foxp3 expression, and Treg cell activity with inhibitory properties on effector cells (35). Nevertheless, it is now clear that TGF- $\beta$  may also facilitate proinflammatory responses through Th17 development, although it needs to act in concert with inflammatory cytokines as IL-6. Thus, the TGF- $\beta$ -induced Treg or Th17 subsets are mutually exclusive depending on the presence of IL-6 (32, 34). The finding of these reciprocal pathways promoted by TGF- $\beta$  could explain the apparent discrepancy that TGF- $\beta$  is involved in both anti- and pro-inflammatory events in the immune system.

Therefore, when the immune system is not activated, in absence of inflammatory cytokines, TGF- $\beta$  favours the generation of adaptative Treg cells, which prevent inflammation and autoimmunity. After infection, cytokines as IL-6 produced as innate response can inhibit such process and collaborate with Treg-produced TGF- $\beta$  to induce proinflammatory Th17 cells (Figure 1).

### TGF- $\beta$ ACTION ON HUMAN Th17 ESTABLISHMENT

Despite the well-known role for TGF- $\beta$  on murine Th17 differentiation, many doubts have arisen about such function on human Th17. Several important reports in 2007 refuted the requirement of TGF- $\beta$  in human IL-17 production (36-39). While some of these studies consider that the central cytokines needed for human Th17 function are IL-23 and IL-1 $\beta$ , other found that IL-1 $\beta$  collaborates with IL-6; however, all of them emphasized that TGF- $\beta$  was dispensable, suggesting important differences in the requirements for the differentiation of Th17 in human and mice.

More recently, three simultaneously published works assure that TGF- $\beta$  is indeed required for the establishment of human Th17 (40-42). Several explanations for these divergent conclusions can be found in different considerations: i) the difficulty to obtain real *naïve* cells from human samples, caution which have been ensured in the works of Manel et al. (41) and Volpe et al. (42); ii) the importance of avoiding any residual contamination with platelets, a main store of TGF- $\beta$ ; iii) or the culture media used in which serum may contain endogenous TGF- $\beta$ . These issues are discussed at length by O'Garra (43). Thus, these new findings support that similar cytokine pathways are involved in Th17 development in mice and humans.

### OTHER INFLUENTIAL CYTOKINES THAT FAVOUR Th17 ACTIVITY

As mentioned above, the early proposal that IL-23 directly drives differentiation to Th17 cells has been discontinued due three main findings: i) IL-23-deficient mice produce Th17 cells which can be expanded *in vitro* by exogenous IL-23 (33); ii) although IL-23 can induces memory T cells to produce IL-17, *naïve* T cells do not express the receptor for IL-23 (44); and iii) it was clearly demonstrated that IL-23 is dispensable for initial IL-17 generation (2, 3, 29). Nevertheless, there is not doubt about the essential role of IL-23 in the maintenance of Th17 cell activity. It has been proposed that IL-23 can have a function of promoting Th17 cell expansion or survival (45). A recent report suggests that IL-23 maintains the Th17 phenotype without affecting proliferation or survival (46). On the other hand, IL-23 has been demonstrated to maintain the pathogenic Th17 functions compared with culture under TGF- $\beta$  and IL-6, depending on IL-10 production by Th17 cells (47).

Currently, IL-6 is the main partner of TGF- $\beta$  in priming *naïve* T cells to IL-17 production (32, 33, 36, 45, 48, 49). It is able to inhibit the expression of Foxp3, which directs the differentiation of CD4<sup>+</sup>CD25<sup>+</sup> Treg cells (50). This may constitute the beginning of the Th17 commitment to detriment of Treg activity during immune response. Moreover, IL-6 orchestrates a series of downstream cytokine-dependent signalling pathways to amplify Th17 cell differentiation. In fact, IL-6 is able to induce the IL-23R expression in T cells making them responsive to the phenotype stabilizer IL-23 (48). On the other hand, after induction of IL-17 expression, Th17 cells start to secrete IL-6 and IL-21, which in turn act as autocrine factors (49, 51).

Although IL-21 does not seem to be essential for the lineage commitment of Th17, it is able to induce IL-17 expression in collaboration with TGF- $\beta$  even in absence of IL-6 (48, 51, 52). Furthermore, generation of Th17 cells is attenuated by blocking IL-21 (53), and loss of its expression, or its receptor, results in defective Th17 differentiation. Similar to IL-6, IL-21 inhibits Foxp3 expression induced by TGF- $\beta$  (51, 52). IL-21 is produced by Th17 cells under IL-6 induction and autocrinally induces its own synthesis and the expression of IL-23R to allow IL-23 responsiveness (48, 51).

Another positive regulator of Th17 development is IL-1. It has been reported that IL-1 $\beta$  can increase the effect of IL-6 and TGF- $\beta$  on Th17 definition (45). Besides, the induction of antigen-specific Th17 cells is abrogated in IL-1R1-deficient mice, without effect on Th1 or Th2 cells (54). However, the certain mechanism through which IL-1 influences Th17 differentiation is not determined

yet. The *IL-18* signaling pathway might be involved in Th17 cell definition as well. Remarkably, in this action would be involved the receptor of IL-18 (IL-18R $\alpha$ ), but not IL-18 itself. Gutcher *et al.* (55) showed that, while IL-18-deficient mice were susceptible to EAE, IL-18R $\alpha$ -deficient animals were resistant to the disease showing a deficient Th17 response, and proposed that IL-18 signaling is involved in Th17-mediated immunopathology through binding of an unknown alternative ligand distinct from IL-18.

#### **NEGATIVE INFLUENTIAL CYTOKINES FOR THE PROMOTION OF Th17 SUBSET**

As Th1 and Th2 cells inhibit the polarization of one another by *IFN $\gamma$*  and *IL4*, respectively (56), there is evidence that these cytokines antagonize initial Th17 development (2, 3). However, after Th17 cells become established effectors, *IFN $\gamma$*  and *IL-4* cannot suppress or revert their phenotype.

One of the main negative regulator of Th17 development is *IL-27* (57, 58), a cytokine structurally related to *IL-6*, but with many different actions. Definitive studies showing such damaging role of *IL-27* on *IL-17* producer cells were reported by Batten *et al.* and by Stumhofer *et al.* (59, 60). Both reports conclude that the absence of *IL-27* signalling exacerbates chronic inflammation in correlation with increased number of Th17 cells. Moreover, *IL-27* is able to promote *IL-10* production (61), another negative player in the network of Th17 activity regulation (62). *IL-10* was first described as a product of Th2 cells, but now is well-known that it is also produced by other cell types as Th1 cells or regulatory Tr1 cells, with important meaning in limiting T-cell mediated immunopathology. The involvement of *IL-10* in regulating the pathogenic function of Th17 cells has been definitively demonstrated by McGeachy *et al.* (47), who have described a non-pathogenic Th17 subtype expressing *IL-10* generated by *IL-6* and *TGF- $\beta$*  in the absence of *IL-23*. Such type of cells not only are non-pathogenic on EAE, but they are also able to prevent the induction of the disease in an *IL-10*-dependent way.

Other negative regulator of Th17 cells is *IL-25* (IL-17E), identified by database searching for genes homologous to *IL-17*. Although *IL-25* is included in the *IL-17* family, it is not produced by Th17, but by Th2 and mast cells. This cytokine is involved in the expression of the Th2 products *IL-5* and *IL-13*, and favour Th2 responses (63). On the other hand, *IL-25* deficiency is involved in pathologic inflammation, associated with increased expression of *IL-17* and *IL-23* (64, 65).

At least in mice, *IL-2* also antagonizes Th17 activity. In spite of the essential function of *IL-2* as growth factor of T effector cells, its deficiency leads to systemic autoimmune disease

(66). This is justified in part to its involvement in differentiation and survival for Treg cells (67). Besides, a recent work has revealed that *IL-2* constrains *IL-17* production since *IL-2* deficiency promotes differentiation of the Th17 cell subset in a *Stat5*-dependent mechanism (68).

#### **ROLE OF Th17 CELLS IN HOST DEFENCE**

The role of *IL-17* in immune response against infections has been widely exemplified (69). *IL-17* is a potent activator of neutrophils, both through cell expansion and through regulation of chemokine expression to cell recruitment. Signaling mediated by *IL-17* induce the target cells to express *CXCL-8* (*IL-8*) and *G-CSF*, which results in generation and accumulation of neutrophils. In addition, *IL-17* induces expression of various anti-microbial genes such as  $\beta$ -defensins. *IL-17* deficiencies are associated with neutrophil defects leading to disease (70). The lamina propria, constituent of mucosa located at respiratory, gastrointestinal and urogenital tracts, is the only tissue where constitutive expression of *IL-17* is detected. Due to the permanent interaction of this tissue with microbial flora, a defensive T cell population prompted to respond to infection may be of great usefulness (71).

Both *IL-17*- and *IL-17RA*-deficient mice show enhanced susceptibility to experimental *Klebsiella pneumoniae* pulmonary infection and reduced *G-CSF* and *CXCL1* in lung in response to this infection (72, 73). In addition to *K. pneumoniae* *IL-17F* has also demonstrated to be involved in pulmonary recruitment of neutrophils (74). Other bacterial infections that induce a preferential Th17 response are infections with *Borrelia burgdorferi*, *Bacteroides fragilis* or *Mycobacterium tuberculosis* (19, 75).

*IL-17* seems to have also a role in protective immunity against fungal infections. Deficiency in *IL-17R* leads to decreased survival and increased damage in kidney after infection with *Candida albicans*, with delayed mobilization of peripheral neutrophils (76). Another major human fungal pathogen which provokes a Th17 response is *Aspergillus fumigata* (77). Other examples of fungal infections which evolve the *IL-23/IL-17* axis are *Cryptococcus neoformans* and *Penicillium carinii* (78, 79).

*IL-17* could be also defensive against some parasites, as infection with the protozoan *Toxoplasma gondii* (60, 80). However, and although an homologue of *IL-17* is encoded by herpes virus Saimiri (81), the role of Th17 in viral infections is unclear. Several studies suggest pathogenic activity of *IL-17* rather than protective function (82, 83). Only against rotavirus infection it has been clearly demonstrated a protective role of Th17 (84).

## ROLE OF Th17 CELLS IN IMMUNOPATHOLOGY

The Th17 subset has a key role in induction and progress of immunopathologies. IL-17 and other cytokines related to the development and function of Th17 cells are closely associated with several immune disorders, not only in animal models, but also in human diseases. As we have referred before, the importance of the Th17 cell activity in autoimmunity was first demonstrated in mice deficient in the p19 chain of IL-23, which showed an important impairment of IL-17 production and were highly resistant to EAE (28) and CIA (31). Using passive transfer studies in EAE, it was demonstrated that IL-17 producing cells are highly pathogenic and essential for the establishment of organ-specific inflammation (29). On the other hand, neutralization of IL-23 can decrease IL-17 expression in central nervous system and prevent EAE (30). In addition, IL-17A deficient mice show reduced symptoms in EAE with delayed onset and early recovery (85), and are resistant to CIA (86). In several independent assays it could be demonstrated that vaccination against IL-17 prevents EAE and CIA (87-89). All these experiments highlighted the importance of the IL-23/IL-17 axis in the pathogenesis of several disorders that was previously thought to be mediated by Th1 subset. Many previous data, inconsistent with the Th1 hypothesis, could be then understood (20-26). The role of IL-17 in autoimmunity has been later underlined by the relationship between therapeutic treatments and IL-17 production impairment. IFN $\beta$ , the currently main therapy used for MS (90), has shown to inhibit IL-17 expression in peripheral and CNS infiltrates T lymphocytes during EAE (91, 92), and to increase the expression of the IL-17 negative regulator IL-27 (93). On the other hand, after transfer of ICOS<sup>+</sup> cells to ICOS-deficient mice, that have enhanced susceptibility to EAE, lymph node cells showed decreased IL-17 production and could reduce the severity of the disease (94).

IL-17 seems to be also associated with the human diseases MS and RA. IL-17 and IL-23 are present in the sera, synovial fluids and synovial biopsies of RA patients (95-97). In addition, it is known that IL-17 mediates induction of IL-6 and IL-8 in RA synovial fibroblasts (98). Before the explosion of the recent advances in the murine Th17 subset knowledge, high IL-17 mRNA expression had been observed in blood and CSF mononuclear cells in human MS (99), and a gene-microarray analysis of MS lesions had suggested that IL-17 and IL-6 could be possible targets for MS therapy (100). Although human Th17 cells are less characterized than the murine subset, some specific features in addition to IL-17 production have been proposed for them, mainly related with specific patterns of chemokine receptor expression which suggest its

involvement in migration of Th17 cells and recruitment of other inflammatory cells (101, 102). More recently, an elevated number of IL-23-expressing dendritic cells has been found in MS patients, concurrent with increased IL-17 production by T cells (103). Moreover, evidence has been provided of that IL-17 and IL-22 induce a breach in the blood-brain barrier and promote recruitment of additional CD4<sup>+</sup> lymphocytes (104). Of particular interest has been the report of Tzartos *et al.*, who found mRNA and protein expression of IL-17 in active areas of MS lesions, where the cell sources of IL-17 were both infiltrating T cells and resident astrocytes and oligodendrocytes (105).

Teunissen *et al.* (106) described the upregulation of IL-17A in psoriasis, an autoimmune disorders affecting skin. IL-17 collaborates with IL-22 in the induction of anti-microbial peptides as a function in host defence against pathogens. However, these defensive peptides can enhance the expression of factors related to psoriatic skin (107). Moreover, expression of different products of Th17 cells has been found in psoriatic skin (39). Another autoimmune disease that has been recently linked to inappropriated Th17 cell response is systemic lupus erythematosus (SLE). IL-17, IL-23 and the number of Th17 cells were higher in plasma from SLE patients than in control individuals, suggesting involvement of the IL-23/IL-17 axis in inflammatory immunity in SLE (108).

There are also data about IL-17 and IL-23 involvement in animal models of induced colitis (109-111). Patients with ulcerative colitis or Crohn's disease display an elevated expression of IL-17 in the intestinal mucosa, which is augmented during active exacerbations of inflammatory bowel disease (112, 113). All these data showing the contribution of IL-17 activity to autoimmune diseases justify clinical trials using a monoclonal antibody anti-IL-17 (AIN457) for Crohn's disease and psoriasis (114).

Allergic asthma is considered to be a Th2-dominant chronic inflammatory disease with a main involvement of eosinophil activity. However, some asthmatic process seem to be mediated by neutrophil infiltration rather than by eosinophil cells, and have been described as "non-eosinophilic asthma" (115-119). Some results in experimental models suggest that Th17 cells may be important for neutrophilic activity in acute airway inflammation (120-122). These data lead some researchers to think that Th17 might be responsible of allergic process mediated by neutrophil cells. However, the role of IL-17 in allergy is still largely unclear. Current results related to this issue has been discussed in length by Oboki *et al.* (123).

## GENETIC PROGRAM GOVERNING TH17 COMMITMENT

In contrast to our wide understanding of the genetic and epigenetic control of Th1 and Th2 regulation, we are at the beginning of the discernment of the genetic mechanisms for the Th17 subset commitment. Th1 and Th2 development are initiated by TCR signalling in conjunction with master transcription factors regulators, T-bet for Th1 and GATA-3 for Th2, which trigger integrated signals with inheritable epigenetic changes that allow tissue-specific gene expression (124). Each of these processes requires the participation of factors that are activated by specific cytokines. IL-12 signalling through Stat4 is associated with Th1 differentiation, and IL-4 through Stat6 governs Th2 cell fate (56). The transcription factors *ROR $\gamma$*  and *ROR $\alpha$*  seems to be hallmark regulators for Th17 development (71, 125). Both factors, belong to the family of retinoic-acid-related orphan nuclear receptors, which in turn is included in the hormone nuclear receptor superfamily. The orphan label is due to that the ligand for these receptors is unknown. While *ROR $\gamma$*  is expressed broadly, the isoform *ROR $\gamma$* t is exclusively found in cells of the immune system (126). *ROR $\gamma$* t is coexpressed with IL-17 in the mucosa constituent lamina propria. Ivanov *et al.* found that *ROR $\gamma$* t is required for IL-17 expression in response to IL-6 and TGF- $\beta$ , and *ROR $\gamma$* t-deficient mice have attenuated EAE and lack tissue-infiltrating Th17 cells (71). Nevertheless, residual Th17 cells are still present in conditions of *ROR $\gamma$*  deficiency and EAE was not completely abolished. This can be justified by the results of Yang *et al.*, showing that another related nuclear receptor, *ROR $\alpha$* , also induced by TGF- $\beta$  and IL-6, can collaborate in Th17 promotion (125). In fact, double deficiencies in *ROR $\alpha$*  and *ROR $\gamma$*  globally impairs Th17 generation and confer more resistance to inflammatory disease than *ROR $\gamma$* -deficient mice. However, single *ROR $\alpha$*  deficiency in T cells only resulted in modest decrease of IL-17 and IL-23R expression, and had a very moderate inhibition of EAE. Thus, these two ROR factors seem to have redundant functions acting in a synergistic way in Th17 cell promoting.

Upregulation of *ROR $\gamma$* t is *Stat3*-dependent (127). This is not the only level at which Stat3 regulates Th17 development since it can also induce the expression of IL-23 R (128). Moreover, the positive effect of IL-6, IL-21 and IL-23 on Th17 subset is dependent of Stat3 (48, 51, 129). On the other hand, negative regulation of Th17 generation by *Socs3* was found to act mainly on IL-23-mediated Stat3 phosphorylation (130). A definitive result to ascribe a role for Stat3 in Th17 development was the finding that Stat3 deficiency resulted in defective Th17-cell differentiation in vivo and protection against EAE (128). Collecting these data together, Stat3 seems to act at several levels during the process of Th17 phenotype

definition, taking part in sequential and feedback loops of cytokine functions: first, IL-6 induce IL-21 production in a Stat-3 dependent mechanism. Once produced, IL-21 autocrinally stimulates its own synthesis and the IL-23R expression through Stat3 activity. This allows IL-23 signalling leading to *ROR $\gamma$* t induction, which in turn up-regulates IL-23R expression in collaboration with Stat3. In addition, IL-21 cooperates with TGF- $\beta$  to promote IL-17 expression with the involvement of Stat3 and *ROR $\gamma$* t (and/or *ROR $\alpha$* ).

Interferon-regulatory factor 4 (*IRF4*), essential for the development of Th2 cells (131-133), is also critical for the generation of Th17 cells and for EAE induction (134). *IRF4*-deficient mice did not develop EAE and showed a fault of IL-17 expression by T helper cells, while transfer of wild-type T helper cells allowed EAE susceptibility. Other feature of *Irf4*<sup>-/-</sup> T cells is the reduced expression of *ROR $\gamma$* t. However, overexpression of this factor in *Irf4*<sup>-/-</sup> T cells only partially restored Th17-cell differentiation. Thus, if *IRF4* acts upstream or downstream of *ROR $\gamma$* t was not clarified.

Recently, it has been reported that *Ets-1* deficiency is associated with increased expression of IL-17, IL-22 and IL-23R in response to IL-6 and TGF- $\beta$ , indicating an enhanced efficiency to Th17 cell differentiation (134). However, *Ets-1* apparently does not affect directly the expression of these genes. Rather, *Ets-1*-deficient T cells produce less IL-2 and have impaired responsiveness in terms of IL-2-mediated inhibition of Th17 differentiation. The resistance to IL-2 suppression was caused by a defect downstream of Stat5 phosphorylation, but was not caused by a difference in the level of *ROR $\gamma$* t.

Regarding direct regulation of IL-17 promoter, chromatin immunoprecipitation (Chip) assays determined that Stat3 directly binds to the murine IL-17A and IL-17F promoters (130). Ichiyama *et al.* reported that *ROR $\gamma$* t directly binds the IL-17 promoter, and found two potential ROR binding sites (135). Besides, *ROR $\gamma$* t was sufficient for activation of IL-17 promoter. Another recent work describes that the transcription factor Runx1 collaborates with *ROR $\gamma$* t to activate this promoter, although in the absence of *ROR $\gamma$* t, Runx1 was not able to induce IL-17 transcription (136). Fewer information is available about the activity of the human IL-17 gene promoter, and it is unknown if *ROR $\gamma$* t is able to bind it. Nevertheless, the minimal promoter has been defined in a region between 232 and 159 nucleotides upstream to the start transcription point. This region contains two NFAT recognition sites which seem to be functional after TCR stimulation at least in Jurkat cells (137). However, no more data about

NFAT role in IL-17 expression control have been reported up to now.

In spite of our poor understanding about the genetic programs directing Th17 cells development and activity, the current thought is that, as for Th1 and Th2 subsets, Th17 polarization obey to an epigenetic control, most probably controlled by ROR $\gamma$ t and ROR $\alpha$  transcription factors, with contribution of other players as Stat3, IRF4, or other still unidentified factors. In mice, the genes for IL-17A and IL-17F are linked on chromosome 1, and their expression also appears to be linked, similarly to the IL-4, IL-5 and IL-13 locus in Th2 cells. The possibility of chromatin remodelling events allowing epigenetic control of Th17-cell development is supported by the results obtained by Akimzhamov *et al.* (138), who showed that *Il17a* and *Il17f* genes undergo H3 acetylation in response to TGF- $\beta$  and IL-6, implying increased accessibility of the locus.

#### REGULATION OF Th17/Treg BALANCE

Current data indicate that the reciprocal developmental pathways for the generation of effector Th17 and Treg cells are controlled by the cytokine environment through an strict transcription program. It is established that CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> Treg cells are generated in the thymus (natural Treg) or may be induced by TGF- $\beta$  in the periphery from naïve CD4<sup>+</sup>CD25<sup>-</sup> cells (induced or adaptive Treg) (67, 139-142). Dichotomy in the generation of Th17 and regulatory Foxp3<sup>+</sup> T cells was first demonstrated by Betelli *et al.* by using of IL-6, which completely inhibited the generation of Foxp3<sup>+</sup> Treg cells induced by TGF- $\beta$ , directing the phenotype fate toward IL-17-producer cells  $\square$ (32).

TGF- $\beta$  alone is able to induce the expression of both master transcription factors involved in each genetic program: ROR $\gamma$ t for Th17 cells and Foxp3 for Treg cells (Figure 2). In spite of this, TGF- $\beta$  does not initiate Th17 differentiation unless pro-inflammatory factors, such as IL-6 or IL-21, are also present. In the absence of these kind of cytokines, Foxp3 interacts with ROR $\gamma$ t suppressing IL-17 transcription (135). IL-6, IL-21, and IL-23 are able to relieve the Foxp3 inhibition of ROR $\gamma$ t, and to active Stat3, allowing Th17 cell differentiation. Zhou *et al.* have proposed that these subtle decision can be influenced by the environmental amount of TGF- $\beta$  (143). According to these authors, at low concentrations, TGF- $\beta$  would synergize with interleukin IL-6 and IL-21, favouring Th17 cell differentiation, while high concentrations of TGF- $\beta$  would repress IL-23R expression and favour Foxp3<sup>+</sup> Treg cells. Runx1 has just been

proposed as a new player in this network of transcription factors regulating Th17/Treg balance (136). Results in this report show that Runx1, in addition to interact with ROR $\gamma$ t on the IL-17 promoter, can be bound by Foxp3 and probably be neutralized in its ability to activate IL-17 transcription.

The function of IL-2, needed to Treg cell survival and negative regulator of Th17 cells, could constitute an additional control in the homeostasis of Treg and Th17 subsets. On the other hand, *retinoids* appear to be another physiologic regulator Th17/Treg differentiation (144-146). Vitamin A derivatives are protective in animal models of autoimmune disease and current data suggest that retinoic acid produced by dendritic cells reciprocally regulates Th17 and Treg differentiation.

It is well-known that cAMP inhibits T cell proliferation (147, 148). This second messenger is involved in suppressor activity by Treg cells (149). In effector cells upon coactivation with Treg cells cAMP is strongly increased, and it was proposed that cAMP can be transferred from Treg to effector targets via cell contact-dependent gap junctions. Although the role of cAMP concerning to Th17 cell function is largely unclear, there are some indications of increased IL-17 production mediated by this second messenger. Chizzolini *et al.* (150), have demonstrated that prostaglandin E2, whose signalling leads to elevated cAMP, synergizes with IL-23 to favor Th17 expansion. Nevertheless, the authors did not analyze if this effect is or not cAMP-dependent. Besides, Yadav *et al.* (151) found that the vasoactive intestinal peptide (VIP) induces Th17 differentiation, in a mechanism suppressed by inhibition of protein kinase A, a main target of cAMP signalling. On the other hand, our unpublished results suggest that some agents increasing cAMP favour *in vitro* Th17 development. Although more clarifying data should be needed about cAMP involvement in Th17 induction, cAMP might be another mediator involved in the Th17/Treg balance, that, like intracellular signalling of TGF- $\beta$ , could have a double function collaborating in Treg or Th17 differentiation depending on environmental signals.

A remarkable novel suggestion about the mechanisms of Th17/Treg balance has been made by Yang *et al.* (152), who suggest that proinflammatory cytokines produced in the inflamed tissue not only affect the generation of induced Treg cells, but also might inhibit the function of already existing Treg cells. In addition, the absence of Foxp3 expression in T cells leads to increased Th1 cell differentiation without enhancing Th17 development. These

data could be interpreted as that the real opposite pathways are those followed by Th1 and Treg cells, while Th17 and Treg generation share intrinsic common programs which allow some plasticity to reprogramming the phenotype of Treg cells towards Th17 effectors. If confirmed, these data could have important implications in therapies against autoimmune diseases with Treg cells, and it should be required to complement them with suppression of IL-6 to avoid redifferentiation into pathogenic Th17 cells.

#### ACKNOWLEDGEMENTS

Authors thank M.J.Jerez for the carefully grammar correction of this manuscript. This work was supported by grants from Instituto de Salud Carlos III (FIS-PI061012) and MM Foundation (MPY-1156/07).

#### REFERENCES

- Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J Immunol* 1986; 136: 2348-2357.
- Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17-producing CD4<sup>+</sup> effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 2005; 6: 1123-1132.
- Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol* 2005; 6: 1133-1141.
- Rouvier E, Luciani MF, Mattei MG, Denizot F, Golstein P CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. *J Immunol* 1993; 150: 5445-5456.
- Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, et al. Human IL-17: a novel cytokine derived from T cells. *J Immunol* 1995; 155: 5483-5486.
- Chang SH, Dong C A novel heterodimeric cytokine consisting of IL-17 and IL-17F regulates inflammatory responses. *Cell Res* 2007; 17: 435-440.
- Wright JF, Guo Y, Quazi A, Luxenberg DP, Bennett F, Ross JF, et al. Identification of an interleukin 17F/17A heterodimer in activated human CD4<sup>+</sup> T cells. *J Biol Chem* 2007; 282: 13447-13455.
- Liang SC, Long AJ, Bennett F, Whitters MJ, Karim R, Collins M, et al. An IL-17F/A heterodimer protein is produced by mouse Th17 cells and induces airway neutrophil recruitment. *J Immunol* 2007; 179: 7791-7799.
- Moseley TA, Haudenschild DR, Rose L, Reddi AH Interleukin-17 family and IL-17 receptors. *Cytokine Growth Factor Rev* 2003; 14: 155-174.
- Haudenschild D, Moseley T, Rose L, Reddi AH Soluble and transmembrane isoforms of novel interleukin-17 receptor-like protein by RNA splicing and expression in prostate cancer. *J Biol Chem* 2002; 277: 4309-4316.
- Hymowitz SG, Filvaroff EH, Yin JP, Lee J, Cai L, Risser P, et al. IL-17s adopt a cystine knot fold: structure and activity of a novel cytokine, IL-17F, and implications for receptor binding. *Embo J* 2001; 20: 5332-5341.
- Gaffen SL An overview of IL-17 function and signaling. *Cytokine* 2008; 43: 402-407.
- Shalom-Barak T, Quach J, Lotz M Interleukin-17-induced gene expression in articular chondrocytes is associated with activation of mitogen-activated protein kinases and NF-kappaB. *J Biol Chem* 1998; 273: 27467-27473.
- Hata K, Andoh A, Shimada M, Fujino S, Bamba S, Araki Y, et al. IL-17 stimulates inflammatory responses via NF-kappaB and MAP kinase pathways in human colonic myofibroblasts. *Am J Physiol Gastrointest Liver Physiol* 2002; 282: G1035-1044.
- Shen F, Hu Z, Goswami J, Gaffen SL Identification of common transcriptional regulatory elements in interleukin-17 target genes. *J Biol Chem* 2006; 281: 24138-24148.
- Patel DN, King CA, Bailey SR, Holt JW, Venkatachalam K, Agrawal A, et al. Interleukin-17 stimulates C-reactive protein expression in hepatocytes and smooth muscle cells via p38 MAPK and ERK1/2-dependent NF-kappaB and C/EBPbeta activation. *J Biol Chem* 2007; 282: 27229-27238.
- Schwandner R, Yamaguchi K, Cao Z Requirement of tumor necrosis factor receptor-associated factor (TRAF)6 in interleukin 17 signal transduction. *J Exp Med* 2000; 191: 1233-1240.
- Hartupée J, Liu C, Novotny M, Li X, Hamilton T IL-17 enhances chemokine gene expression through mRNA stabilization. *J Immunol* 2007; 179: 4135-4141.
- Infante-Duarte C, Horton HF, Byrne MC, Kamradt T Microbial lipopeptides induce the production of IL-17 in Th cells. *J Immunol* 2000; 165: 6107-6115.
- Betelli E, Sullivan B, Szabo SJ, Sobel RA, Glimcher LH, Kuchroo VK Loss of T-bet, but not STAT1, prevents the development of experimental autoimmune encephalomyelitis. *J Exp Med* 2004; 200: 79-87.
- Zhang GX, Gran B, Yu S, Li J, Siglienti I, Chen X, et al. Induction of experimental autoimmune encephalomyelitis in IL-12

- receptor-beta 2-deficient mice: IL-12 responsiveness is not required in the pathogenesis of inflammatory demyelination in the central nervous system. *J Immunol* 2003; 170: 2153-2160.
22. Billiau A, Heremans H, Vandekerckhove F, Dijkmans R, Sobis H, Meulepas E, et al. Enhancement of experimental allergic encephalomyelitis in mice by antibodies against IFN-gamma. *J Immunol* 1988; 140: 1506-1510.
  23. Gran B, Chu N, Zhang GX, Yu S, Li Y, Chen XH, et al. Early administration of IL-12 suppresses EAE through induction of interferon-gamma. *J Neuroimmunol* 2004; 156: 123-131.
  24. Gran B, Zhang GX, Yu S, Li J, Chen XH, Ventura ES, et al. IL-12p35-deficient mice are susceptible to experimental autoimmune encephalomyelitis: evidence for redundancy in the IL-12 system in the induction of central nervous system autoimmune demyelination. *J Immunol* 2002; 169: 7104-7110.
  25. Voorthuis JA, Uitdehaag BM, De Groot CJ, Goede PH, van der Meide PH, Dijkstra CD. Suppression of experimental allergic encephalomyelitis by intraventricular administration of interferon-gamma in Lewis rats. *Clin Exp Immunol* 1990; 81: 183-188.
  26. Willenborg DO, Fordham SA, Staykova MA, Ramshaw IA, Cowden WB. IFN-gamma is critical to the control of murine autoimmune encephalomyelitis and regulates both in the periphery and in the target tissue: a possible role for nitric oxide. *J Immunol* 1999; 163: 5278-5286.
  27. Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity* 2000; 13: 715-725.
  28. Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. *Nature* 2003; 421: 744-748.
  29. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med* 2005; 201: 233-240.
  30. Chen Y, Langrish CL, McKenzie B, Joyce-Shaikh B, Stumhofer JS, McClanahan T, et al. Anti-IL-23 therapy inhibits multiple inflammatory pathways and ameliorates autoimmune encephalomyelitis. *J Clin Invest* 2006; 116: 1317-1326.
  31. Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, et al. Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. *J Exp Med* 2003; 198: 1951-1957.
  32. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* 2006; 441: 235-238.
  33. Mangan PR, Harrington LE, O'Quinn DB, Helms WS, Bullard DC, Elson CO, et al. Transforming growth factor-beta induces development of the T(H)17 lineage. *Nature* 2006; 441: 231-234.
  34. Veldhoen M, Hocking RJ, Flavell RA, Stockinger B. Signals mediated by transforming growth factor-beta initiate autoimmune encephalomyelitis, but chronic inflammation is needed to sustain disease. *Nat Immunol* 2006; 7: 1151-1156.
  35. Lohr J, Knoechel B, Abbas AK. Regulatory T cells in the periphery. *Immunol Rev* 2006; 212: 149-162.
  36. Acosta-Rodriguez EV, Napolitani G, Lanzavecchia A, Sallusto F. Interleukin 1beta and 6 but not transforming growth factor-beta are essential for the differentiation of interleukin 17-producing human T helper cells. *Nat Immunol* 2007; 8: 942-949.
  37. Chen Z, Tato CM, Muul L, Laurence A, O'Shea JJ. Distinct regulation of interleukin-17 in human T helper lymphocytes. *Arthritis Rheum* 2007; 56: 2936-2946.
  38. van Beelen AJ, Zelinkova Z, Taanman-Kueter EW, Muller FJ, Hommes DW, Zaat SA, et al. Stimulation of the intracellular bacterial sensor NOD2 programs dendritic cells to promote interleukin-17 production in human memory T cells. *Immunity* 2007; 27: 660-669.
  39. Wilson NJ, Boniface K, Chan JR, McKenzie BS, Blumenschein WM, Mattson JD, et al. Development, cytokine profile and function of human interleukin 17-producing helper T cells. *Nat Immunol* 2007; 8: 950-957.
  40. Yang L, Anderson DE, Baecher-Allan C, Hastings WD, Bettelli E, Oukka M, et al. IL-21 and TGF-beta are required for differentiation of human T(H)17 cells. *Nature* 2008; 454: 350-352.
  41. Manel N, Unutmaz D, Littman DR. The differentiation of human T(H)-17 cells requires transforming growth factor-beta and induction of the nuclear receptor RORgamma. *Nat Immunol* 2008; 9: 641-649.
  42. Volpe E, Servant N, Zollinger R, Bogiatzi SI, Hupe P, Barillot E, et al. A critical function for transforming growth factor-beta, interleukin 23 and proinflammatory cytokines in driving and modulating human T(H)-17 responses. *Nat Immunol* 2008; 9: 650-657.
  43. O'Garra A, Stockinger B, Veldhoen M. Differentiation of human T(H)-17 cells does require TGF-beta! *Nat Immunol* 2008; 9: 588-590.

44. Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. *J Biol Chem* 2003; 278: 1910-1914.
45. Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. *Immunity* 2006; 24: 179-189.
46. Stritesky GL, Yeh N, Kaplan MH IL-23 promotes maintenance but not commitment to the Th17 lineage. *J Immunol* 2008; 181: 5948-5955.
47. McGeachy MJ, Bak-Jensen KS, Chen Y, Tato CM, Blumenschein W, McClanahan T, et al. TGF-beta and IL-6 drive the production of IL-17 and IL-10 by T cells and restrain T(H)-17 cell-mediated pathology. *Nat Immunol* 2007; 8: 1390-1397.
48. Zhou L, Ivanov, II, Spolski R, Min R, Shenderov K, Egawa T, et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. *Nat Immunol* 2007; 8: 967-974.
49. Ogura H, Murakami M, Okuyama Y, Tsuruoka M, Kitabayashi C, Kanamoto M, et al. Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction. *Immunity* 2008; 29: 628-636.
50. Kimura A, Naka T, Kishimoto T IL-6-dependent and -independent pathways in the development of interleukin 17-producing T helper cells. *Proc Natl Acad Sci U S A* 2007; 104: 12099-12104.
51. Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, et al. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. *Nature* 2007; 448: 480-483.
52. Korn T, Bettelli E, Gao W, Awasthi A, Jager A, Strom TB, et al. IL-21 initiates an alternative pathway to induce proinflammatory T(H)17 cells. *Nature* 2007; 448: 484-487.
53. Wei L, Laurence A, Elias KM, O'Shea JJ IL-21 is produced by Th17 cells and drives IL-17 production in a STAT3-dependent manner. *J Biol Chem* 2007; 282: 34605-34610.
54. Sutton C, Brereton C, Keogh B, Mills KH, Lavelle EC A crucial role for interleukin (IL)-1 in the induction of IL-17-producing T cells that mediate autoimmune encephalomyelitis. *J Exp Med* 2006; 203: 1685-1691.
55. Gutcher I, Urich E, Wolter K, Prinz M, Becher B Interleukin 18-independent engagement of interleukin 18 receptor-alpha is required for autoimmune inflammation. *Nat Immunol* 2006; 7: 946-953.
56. Murphy KM, Reiner SL The lineage decisions of helper T cells. *Nat Rev Immunol* 2002; 2: 933-944.
57. Stumhofer JS, Silver J, Hunter CA Negative regulation of Th17 responses. *Semin Immunol* 2007; 19: 394-399.
58. Colgan J, Rothman P All in the family: IL-27 suppression of T(H)-17 cells. *Nat Immunol* 2006; 7: 899-901.
59. Batten M, Li J, Yi S, Kljavin NM, Danilenko DM, Lucas S, et al. Interleukin 27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. *Nat Immunol* 2006; 7: 929-936.
60. Stumhofer JS, Laurence A, Wilson EH, Huang E, Tato CM, Johnson LM, et al. Interleukin 27 negatively regulates the development of interleukin 17-producing T helper cells during chronic inflammation of the central nervous system. *Nat Immunol* 2006; 7: 937-945.
61. Stumhofer JS, Silver JS, Laurence A, Porrett PM, Harris TH, Turka LA, et al. Interleukins 27 and 6 induce STAT3-mediated T cell production of interleukin 10. *Nat Immunol* 2007; 8: 1363-1371.
62. Jankovic D, Trinchieri G IL-10 or not IL-10: that is the question. *Nat Immunol* 2007; 8: 1281-1283.
63. Fort MM, Cheung J, Yen D, Li J, Zurawski SM, Lo S, et al. IL-25 induces IL-4, IL-5, and IL-13 and Th2-associated pathologies in vivo. *Immunity* 2001; 15: 985-995.
64. Owyang AM, Zaph C, Wilson EH, Guild KJ, McClanahan T, Miller HR, et al. Interleukin 25 regulates type 2 cytokine-dependent immunity and limits chronic inflammation in the gastrointestinal tract. *J Exp Med* 2006; 203: 843-849.
65. Kleinschek MA, Owyang AM, Joyce-Shaikh B, Langrish CL, Chen Y, Gorman DM, et al. IL-25 regulates Th17 function in autoimmune inflammation. *J Exp Med* 2007; 204: 161-170.
66. Abbas AK The control of T cell activation vs. tolerance. *Autoimmun Rev* 2003; 2: 115-118.
67. Shevach EM, DiPaolo RA, Andersson J, Zhao DM, Stephens GL, Thornton AM The lifestyle of naturally occurring CD4+ CD25+ Foxp3+ regulatory T cells. *Immunol Rev* 2006; 212: 60-73.
68. Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, et al. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. *Immunity* 2007; 26: 371-381.
69. Matsuzaki G, Umemura M Interleukin-17 as an effector molecule of innate and acquired immunity against infections. *Microbiol Immunol* 2007; 51: 1139-1147.
70. Linden A, Laan M, Anderson GP Neutrophils, interleukin-17A and lung disease. *Eur Respir J* 2005; 25: 159-172.
71. Ivanov, II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al.

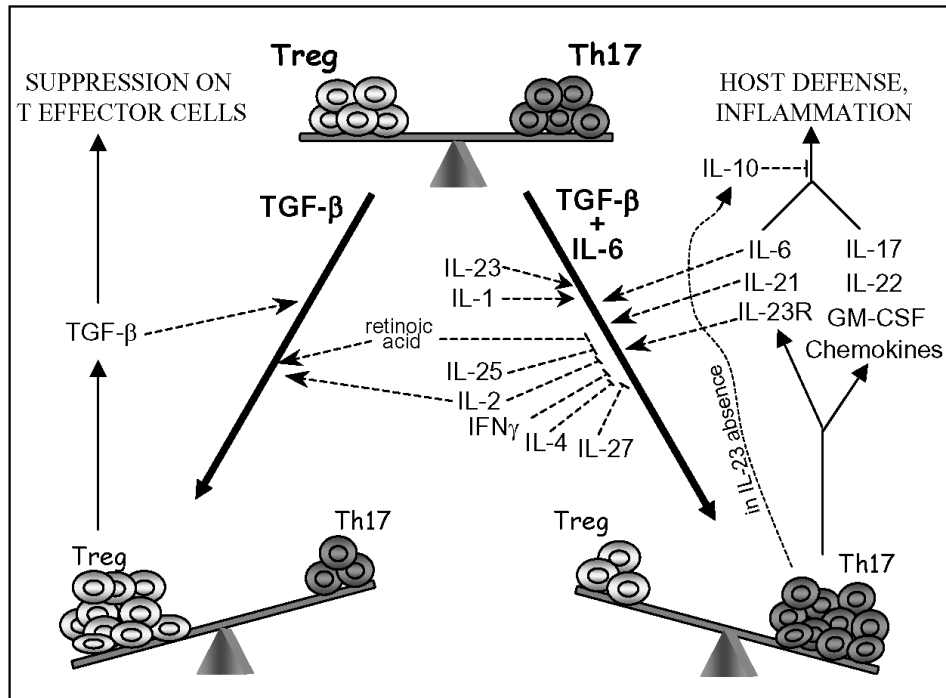
- The orphan nuclear receptor ROR $\gamma$  directs the differentiation program of proinflammatory IL-17<sup>+</sup> T helper cells. *Cell* 2006; 126: 1121-1133.
72. Aujla SJ, Chan YR, Zheng M, Fei M, Askew DJ, Pociask DA, et al. IL-22 mediates mucosal host defense against Gram-negative bacterial pneumonia. *Nat Med* 2008; 14: 275-281.
  73. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, et al. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. *J Exp Med* 2001; 194: 519-527.
  74. Yang XO, Chang SH, Park H, Nurieva R, Shah B, Acero L, et al. Regulation of inflammatory responses by IL-17F. *J Exp Med* 2008; 205: 1063-1075.
  75. Chung DR, Kasper DL, Panzo RJ, Chitnis T, Grusby MJ, Sayegh MH, et al. CD4<sup>+</sup> T cells mediate abscess formation in intra-abdominal sepsis by an IL-17-dependent mechanism. *J Immunol* 2003; 170: 1958-1963.
  76. Huang W, Na L, Fidel PL, Schwarzenberger P Requirement of interleukin-17A for systemic anti-*Candida albicans* host defense in mice. *J Infect Dis* 2004; 190: 624-631.
  77. Zelante T, De Luca A, Bonifazi P, Montagnoli C, Bozza S, Moretti S, et al. IL-23 and the Th17 pathway promote inflammation and impair antifungal immune resistance. *Eur J Immunol* 2007; 37: 2695-2706.
  78. Kleinschek MA, Muller U, Brodie SJ, Stenzel W, Kohler G, Blumenschein WM, et al. IL-23 enhances the inflammatory cell response in *Cryptococcus neoformans* infection and induces a cytokine pattern distinct from IL-12. *J Immunol* 2006; 176: 1098-1106.
  79. Rudner XL, Happel KI, Young EA, Shellito JE Interleukin-23 (IL-23)-IL-17 cytokine axis in murine *Pneumocystis carinii* infection. *Infect Immun* 2007; 75: 3055-3061.
  80. Kelly MN, Kolls JK, Happel K, Schwartzman JD, Schwarzenberger P, Combe C, et al. Interleukin-17/interleukin-17 receptor-mediated signaling is important for generation of an optimal polymorphonuclear response against *Toxoplasma gondii* infection. *Infect Immun* 2005; 73: 617-621.
  81. Yao Z, Fanslow WC, Seldin MF, Rousseau AM, Painter SL, Comeau MR, et al. Herpesvirus Saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. *Immunity* 1995; 3: 811-821.
  82. Maertzdorf J, Osterhaus AD, Verjans GM IL-17 expression in human herpetic stromal keratitis: modulatory effects on chemokine production by corneal fibroblasts. *J Immunol* 2002; 169: 5897-5903.
  83. Patera AC, Pesnicak L, Bertin J, Cohen JI Interleukin 17 modulates the immune response to vaccinia virus infection. *Virology* 2002; 299: 56-63.
  84. Smiley KL, McNeal MM, Basu M, Choi AH, Clements JD, Ward RL Association of gamma interferon and interleukin-17 production in intestinal CD4<sup>+</sup> T cells with protection against rotavirus shedding in mice intranasally immunized with VP6 and the adjuvant LT(R192G). *J Virol* 2007; 81: 3740-3748.
  85. Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *J Immunol* 2006; 177: 566-573.
  86. Nakae S, Nambu A, Sudo K, Iwakura Y Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J Immunol* 2003; 171: 6173-6177.
  87. Lubberts E, Koenders MI, Oppers-Walgreen B, van den Bersselaar L, Coenend Roo CJ, Joosten LA, et al. Treatment with a neutralizing anti-murine interleukin-17 antibody after the onset of collagen-induced arthritis reduces joint inflammation, cartilage destruction, and bone erosion. *Arthritis Rheum* 2004; 50: 650-659.
  88. Rohn TA, Jennings GT, Hernandez M, Grest P, Beck M, Zou Y, et al. Vaccination against IL-17 suppresses autoimmune arthritis and encephalomyelitis. *Eur J Immunol* 2006; 36: 2857-2867.
  89. Uyttenhove C, Van Snick J Development of an anti-IL-17A auto-vaccine that prevents experimental autoimmune encephalomyelitis. *Eur J Immunol* 2006; 36: 2868-2874.
  90. Weinstock-Guttman B, Ramanathan M, Zivadinov R Interferon-beta treatment for relapsing multiple sclerosis. *Expert Opin Biol Ther* 2008; 8: 1435-1447.
  91. Martin-Saavedra FM, Flores N, Dorado B, Eguiluz C, Bravo B, Garcia-Merino A, et al. Beta-interferon unbalances the peripheral T cell proinflammatory response in experimental autoimmune encephalomyelitis. *Mol Immunol* 2007; 44: 3597-3607.
  92. Martin-Saavedra FM, Gonzalez-Garcia C, Bravo B, Ballester S Beta interferon restricts the inflammatory potential of CD4<sup>+</sup> cells through the boost of the Th2 phenotype, the inhibition of Th17 response and the prevalence of naturally occurring T regulatory cells. *Mol Immunol* 2008; 45: 4008-4019.
  93. Guo B, Chang EY, Cheng G The type I IFN induction pathway constrains Th17-mediated autoimmune inflammation in mice. *J Clin Invest* 2008; 118: 1680-1690.

94. Rojo JM, Pini E, Ojeda G, Bello R, Dong C, Flavell RA, et al. CD4+ICOS+ T lymphocytes inhibit T cell activation 'in vitro' and attenuate autoimmune encephalitis 'in vivo'. *Int Immunol* 2008; 20: 577-589.
95. Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest* 1999; 103: 1345-1352.
96. Kim HR, Cho ML, Kim KW, Juhn JY, Hwang SY, Yoon CH, et al. Up-regulation of IL-23p19 expression in rheumatoid arthritis synovial fibroblasts by IL-17 through PI3-kinase-, NF-kappaB- and p38 MAPK-dependent signalling pathways. *Rheumatology (Oxford)* 2007; 46: 57-64.
97. Cho ML, Yoon CH, Hwang SY, Park MK, Min SY, Lee SH, et al. Effector function of type II collagen-stimulated T cells from rheumatoid arthritis patients: cross-talk between T cells and synovial fibroblasts. *Arthritis Rheum* 2004; 50: 776-784.
98. Hwang SY, Kim JY, Kim KW, Park MK, Moon Y, Kim WU, et al. IL-17 induces production of IL-6 and IL-8 in rheumatoid arthritis synovial fibroblasts via NF-kappaB- and PI3-kinase/Akt-dependent pathways. *Arthritis Res Ther* 2004; 6: R120-128.
99. Matusevicius D, Kivisakk P, He B, Kostulas N, Ozenci V, Fredrikson S, et al. Interleukin-17 mRNA expression in blood and CSF mononuclear cells is augmented in multiple sclerosis. *Mult Scler* 1999; 5: 101-104.
100. Lock C, Hermans G, Pedotti R, Brendolan A, Schadt E, Garren H, et al. Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis. *Nat Med* 2002; 8: 500-508.
101. Acosta-Rodriguez EV, Rivino L, Geginat J, Jarrossay D, Gattorno M, Lanzavecchia A, et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. *Nat Immunol* 2007; 8: 639-646.
102. Sato W, Aranami T, Yamamura T Cutting edge: Human Th17 cells are identified as bearing CCR2+CCR5- phenotype. *J Immunol* 2007; 178: 7525-7529.
103. Vaknin-Dembinsky A, Balashov K, Weiner HL IL-23 is increased in dendritic cells in multiple sclerosis and down-regulation of IL-23 by antisense oligos increases dendritic cell IL-10 production. *J Immunol* 2006; 176: 7768-7774.
104. Kebir H, Kreymborg K, Ifergan I, Dodelet-Devillers A, Cayrol R, Bernard M, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med* 2007; 13: 1173-1175.
105. Tzartos JS, Friese MA, Craner MJ, Palace J, Newcombe J, Esiri MM, et al. Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol* 2008; 172: 146-155.
106. Teunissen MB, Kooimen CW, de Waal Malefyt R, Wierenga EA, Bos JD Interleukin-17 and interferon-gamma synergize in the enhancement of proinflammatory cytokine production by human keratinocytes. *J Invest Dermatol* 1998; 111: 645-649.
107. Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med* 2006; 203: 2271-2279.
108. Wong CK, Lit LC, Tam LS, Li EK, Wong PT, Lam CW Hyperproduction of IL-23 and IL-17 in patients with systemic lupus erythematosus: implications for Th17-mediated inflammation in auto-immunity. *Clin Immunol* 2008; 127: 385-393.
109. Uhlig HH, McKenzie BS, Hue S, Thompson C, Joyce-Shaikh B, Stepankova R, et al. Differential activity of IL-12 and IL-23 in mucosal and systemic innate immune pathology. *Immunity* 2006; 25: 309-318.
110. Yen D, Cheung J, Scheerens H, Poulet F, McClanahan T, McKenzie B, et al. IL-23 is essential for T cell-mediated colitis and promotes inflammation via IL-17 and IL-6. *J Clin Invest* 2006; 116: 1310-1316.
111. Zhang Z, Zheng M, Bindas J, Schwarzenberger P, Kolls JK Critical role of IL-17 receptor signaling in acute TNBS-induced colitis. *Inflamm Bowel Dis* 2006; 12: 382-388.
112. Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, et al. Increased expression of interleukin 17 in inflammatory bowel disease. *Gut* 2003; 52: 65-70.
113. Annunziato F, Cosmi L, Santarlasci V, Maggi L, Liotta F, Mazzinghi B, et al. Phenotypic and functional features of human Th17 cells. *J Exp Med* 2007; 204: 1849-1861.
114. Ouyang W, Kolls JK, Zheng Y The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity* 2008; 28: 454-467.
115. Fahy JV, Kim KW, Liu J, Boushey HA Prominent neutrophilic inflammation in sputum from subjects with asthma exacerbation. *J Allergy Clin Immunol* 1995; 95: 843-852.
116. Wenzel SE, Schwartz LB, Langmack EL, Halliday JL, Trudeau JB, Gibbs RL, et al. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and

- clinical characteristics. *Am J Respir Crit Care Med* 1999; 160: 1001-1008.
117. Jatakanon A, Uasuf C, Maziak W, Lim S, Chung KF, Barnes PJ Neutrophilic inflammation in severe persistent asthma. *Am J Respir Crit Care Med* 1999; 160: 1532-1539.
  118. Ordonez CL, Shaughnessy TE, Matthay MA, Fahy JV Increased neutrophil numbers and IL-8 levels in airway secretions in acute severe asthma: Clinical and biologic significance. *Am J Respir Crit Care Med* 2000; 161: 1185-1190.
  119. Gibson PG, Simpson JL, Saltos N Heterogeneity of airway inflammation in persistent asthma : evidence of neutrophilic inflammation and increased sputum interleukin-8. *Chest* 2001; 119: 1329-1336.
  120. Hellings PW, Kasran A, Liu Z, Vandekerckhove P, Wuyts A, Overbergh L, et al. Interleukin-17 orchestrates the granulocyte influx into airways after allergen inhalation in a mouse model of allergic asthma. *Am J Respir Cell Mol Biol* 2003; 28: 42-50.
  121. Prause O, Bozinovski S, Anderson GP, Linden A Increased matrix metalloproteinase-9 concentration and activity after stimulation with interleukin-17 in mouse airways. *Thorax* 2004; 59: 313-317.
  122. Hoshino H, Laan M, Sjostrand M, Lotvall J, Skoogh BE, Linden A Increased elastase and myeloperoxidase activity associated with neutrophil recruitment by IL-17 in airways in vivo. *J Allergy Clin Immunol* 2000; 105: 143-149.
  123. Oboki K, Ohno T, Saito H, Nakae S Th17 and allergy. *Allergol Int* 2008; 57: 121-134.
  124. Lee GR, Kim ST, Spilianakis CG, Fields PE, Flavell RA T helper cell differentiation: regulation by cis elements and epigenetics. *Immunity* 2006; 24: 369-379.
  125. Yang XO, Pappu BP, Nurieva R, Akimzhanov A, Kang HS, Chung Y, et al. T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR alpha and ROR gamma. *Immunity* 2008; 28: 29-39.
  126. Eberl G, Littman DR The role of the nuclear hormone receptor RORgamma in the development of lymph nodes and Peyer's patches. *Immunol Rev* 2003; 195: 81-90.
  127. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *J Biol Chem* 2007; 282: 9358-9363.
  128. Harris TJ, Grosso JF, Yen HR, Xin H, Kortylewski M, Albesiano E, et al. Cutting edge: An in vivo requirement for STAT3 signaling in TH17 development and TH17-dependent autoimmunity. *J Immunol* 2007; 179: 4313-4317.
  129. Chen Z, Laurence A, O'Shea JJ Signal transduction pathways and transcriptional regulation in the control of Th17 differentiation. *Semin Immunol* 2007; 19: 400-408.
  130. Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, et al. Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. *Proc Natl Acad Sci U S A* 2006; 103: 8137-8142.
  131. Hu CM, Jang SY, Fanzo JC, Pernis AB Modulation of T cell cytokine production by interferon regulatory factor-4. *J Biol Chem* 2002; 277: 49238-49246.
  132. Lohoff M, Mittrucker HW, Prechtel S, Bischof S, Sommer F, Kock S, et al. Dysregulated T helper cell differentiation in the absence of interferon regulatory factor 4. *Proc Natl Acad Sci U S A* 2002; 99: 11808-11812.
  133. Rengarajan J, Mowen KA, McBride KD, Smith ED, Singh H, Glimcher LH Interferon regulatory factor 4 (IRF4) interacts with NFATc2 to modulate interleukin 4 gene expression. *J Exp Med* 2002; 195: 1003-1012.
  134. Brustle A, Heink S, Huber M, Rosenplanter C, Stadelmann C, Yu P, et al. The development of inflammatory T(H)-17 cells requires interferon-regulatory factor 4. *Nat Immunol* 2007; 8: 958-966.
  135. Ichiyama K, Yoshida H, Wakabayashi Y, Chinen T, Saeki K, Nakaya M, et al. Foxp3 inhibits RORgamma-mediated IL-17A mRNA transcription through direct interaction with RORgamma. *J Biol Chem* 2008; 283: 17003-17008.
  136. Zhang F, Meng G, Strober W Interactions among the transcription factors Runx1, RORgamma and Foxp3 regulate the differentiation of interleukin 17-producing T cells. *Nat Immunol* 2008; 9: 1297-1306.
  137. Liu XK, Lin X, Gaffen SL Crucial role for nuclear factor of activated T cells in T cell receptor-mediated regulation of human interleukin-17. *J Biol Chem* 2004; 279: 52762-52771.
  138. Akimzhanov AM, Yang XO, Dong C Chromatin remodeling of interleukin-17 (IL-17)-IL-17F cytokine gene locus during inflammatory helper T cell differentiation. *J Biol Chem* 2007; 282: 5969-5972.
  139. Sakaguchi S Regulatory T cells: key controllers of immunologic self-tolerance. *Cell* 2000; 101: 455-458.
  140. Chen W, Jin W, Hardegen N, Lei KJ, Li L, Marinos N, et al. Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. *J Exp Med* 2003; 198: 1875-1886.
  141. Faria AM, Weiner HL Oral tolerance and TGF-beta-producing cells. *Inflamm Allergy Drug Targets* 2006; 5: 179-190.
  142. Pini E, Ojeda G, Portolés P The renaissance of T regulatory cells: Looking for markers in a haystack. *Inmunología* 2007; 26: 100-107.

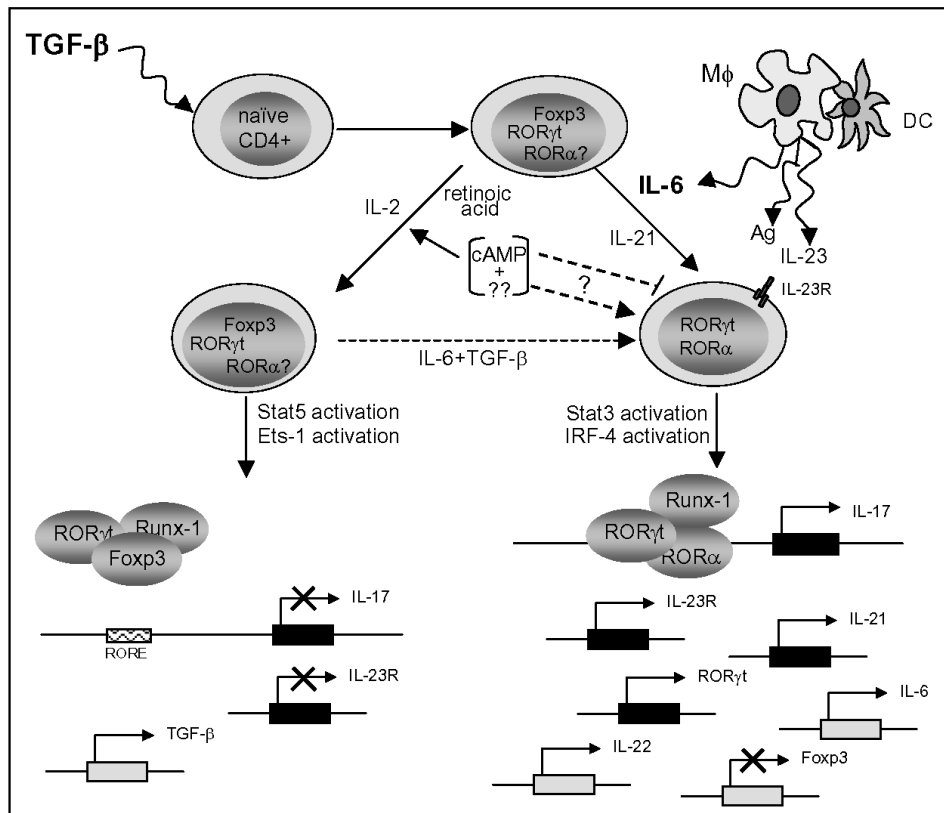
143. Zhou L, Lopes JE, Chong MM, Ivanov, II, Min R, Victora GD, et al. TGF-beta-induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing RORgammat function. *Nature* 2008; 453: 236-240.
144. Schambach F, Schupp M, Lazar MA, Reiner SL. Activation of retinoic acid receptor-alpha favours regulatory T cell induction at the expense of IL-17-secreting T helper cell differentiation. *Eur J Immunol* 2007; 37: 2396-2399.
145. Elias KM, Laurence A, Davidson TS, Stephens G, Kanno Y, Shevach EM, et al. Retinoic acid inhibits Th17 polarization and enhances FoxP3 expression through a Stat-3/Stat-5 independent signaling pathway. *Blood* 2008; 111: 1013-1020.
146. Mucida D, Park Y, Kim G, Turovskaya O, Scott I, Kronenberg M, et al. Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. *Science* 2007; 317: 256-260.
147. Molina CA, Foulkes NS, Lalli E, Sassone-Corsi P. Inducibility and negative autoregulation of CREM: an alternative promoter directs the expression of ICER, an early response repressor. *Cell* 1993; 75: 875-886.
148. Lalli E, Lee JS, Masquillier D, Schlotter F, Foulkes NS, Molina CA, et al. Nuclear response to cyclic AMP: central role of transcription factor CREM (cyclic-AMP-responsive-element modulator). *Biochem Soc Trans* 1993; 21: 912-917.
149. Bopp T, Becker C, Klein M, Klein-Hessling S, Palmetshofer A, Serfling E, et al. Cyclic adenosine monophosphate is a key component of regulatory T cell-mediated suppression. *J Exp Med* 2007; 204: 1303-1310.
150. Chizzolini C, Chicheportiche R, Alvarez M, de Rham C, Roux-Lombard P, Ferrarilacraz S, et al. Prostaglandin E2 synergistically with interleukin-23 favors human Th17 expansion. *Blood* 2008; 112: 3696-3703.
151. Yadav M, Rosenbaum J, Goetzl EJ. Cutting edge: vasoactive intestinal peptide (VIP) induces differentiation of Th17 cells with a distinctive cytokine profile. *J Immunol* 2008; 180: 2772-2776.
152. Yang XO, Nurieva R, Martinez GJ, Kang HS, Chung Y, Pappu BP, et al. Molecular antagonism and plasticity of regulatory and inflammatory T cell programs. *Immunity* 2008; 29: 44-56.

# ACEPTADO



**Figure 1.- The homeostasis of Treg and Th17 cells can be balanced to each phenotype by the cytokine environment.** TGF- $\beta$  is involved in both pathway and IL-6 is the decisive cytokine directing the cell subset fate. In the absence of IL-6, TGF- $\beta$  induces the establishment of Treg cells that in turn produce TGF- $\beta$  and control inflammation and autoimmunity by suppression of T effector cells. This process is favoured by IL-2 and retinoids. The Th17 pathway is triggered by inflammatory cytokines as IL-6. Once Th17 cells are committed, the IL-23R is expressed, allowing responsiveness to IL-23 which stabilizes the phenotype of IL-17 producer cells. IL-6 and IL-21 are also produced, and both of them take part in an autocrine loop that collaborates in Th17 development. Negative regulators of Th17 differentiation are IL-27, IL-4, IFN- $\gamma$ , IL-25, IL-2 and retinoids. In addition to IL-17, IL-21 and IL-6, Th17 cells secrete other products, as chemokines, GM-CSF and IL-22, all of them involved in host defence and inflammation. In the absence of IL-23, Th17 cells can also produce IL-10 which may control the pathogenic phenotype of IL-17 producer cells.

# ACEPTADO



**Figure 2.- Antagonic Treg and Th17 generations share intrinsic genetic program.** TGF- $\beta$  activity on naïve CD4<sup>+</sup> cells induces the expression of both Foxp3 and ROR $\gamma$ t transcription factors. In absence of IL-6, ROR $\gamma$ t and Runx-1 are sequestered by Foxp3 binding; this interaction avoids contact with ROR elements (RORE) in the IL-17 promoter. Cascade signalling triggered by IL-6 leads in naïve, and probably in Treg defined cells, to Stat3 activation, ROR $\gamma$ t expression, down-regulation of Foxp3 expression, and suppress the binding of pre-existing Foxp3 to ROR $\gamma$ t. In a Stat3-dependent mechanism, the activity of ROR $\gamma$ t, ROR $\alpha$  and Runx-1 allows IL-17 gene expression. The subsequent expression of IL-23R, IL-6 and IL-21 guide a feedback of signals to amplify and stabilize the Th17 phenotype. IRF-4 activation by antigen (Ag) presentation also collaborates to Th17 cell differentiation, remaining to be determined if upstream or downstream of ROR $\gamma$ t. Dark boxes correspond to Stat3 regulated genes.