

# SUPPLEMENTARY MATERIALS AND METHODS

## **PanDrugs: a novel method to prioritize anticancer drug treatments according to individual genomic data**

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## SUPPLEMENTARY TEXT

### MATERIAL AND METHODS

#### Drug-Gene data sources

In order to construct PanDrugs database (PanDrugsdb), we have collected and integrated relationships between drugs and genes from 18 sources with different origins and levels of information including data from experimental studies in cancer cell lines (Supplementary Table 1):

**DGIdb:** DGIdb [1] constitutes a comprehensive catalogue of information about gene druggability. They classify this information into two main categories: i) known drug-gene associations from databases and literature, and ii) potentially druggable genes based on their belonging to a particular druggable gene category. To build PanDrugsdb, we focused in the first kind of information that DGIdb mine from several resources (Cancer Commons, Cancer Genome Interpreter (CGI), ChEMBLInteractions, CIViC, CKB, Clarity Foundation, DoCM, DrugBank, FDA, Guide to PHARMACOLOGY, My Cancer Genome, NCI, OncoKB, PharmGKB, TTD and the information in the TALC, TDG and TEND studies). We accessed DGIdb using its Application Programming Interface (API) to retrieve the drugs associated with all the human genes.

**Monoclonal antibodies:** Targeted therapy with monoclonal antibodies is highly selective and has been established as a successful treatment in several diseases and specially in cancer. For this reason we decided to incorporate this information in PanDrugsdb. Most of this information derives from the list of therapeutic monoclonal antibodies (moAb) from Carter and Lazar [2].

**TARGET:** Another source of information is the tumor alterations relevant for genomics-driven therapy (TARGET) database [3] supported by the Cancer Genome Analysis of the Broad Institute, which includes genes somatically altered in cancer, associated with clinical actions in a standard spreadsheet file. We downloaded the last version available at the moment of the query (TARGET\_db\_v3\_02142015.xlsx). We selected among the records those with a specific drug name, extracting also the additional information they provide about the sensitivity or resistance response, the type of drug-gene relation understood as a drug target or biomarker and the type of genomic alteration associated to the drug response.

**Cancer Therapeutics Response Portal (CTRP):** In this study [4,5], corresponding to the CTRP V1, the authors measure the sensitivity of 242 genomically characterized cancer cell lines to a set of 354 small molecules in different approval status (approved, clinical candidates and probes). The information about interactions was extracted from the supplementary file 2 of the article (mmc2.xlsx). We selected the significant drug-gene interactions using as threshold q-value = 0.05. Both sensitivity and unresponsive were maintained. We filtered out from the list that ambiguous information where both types of response (sensitivity or unresponsive) appear related to the alteration of a particular gene.

**Genomics of Drug Sensitivity in Cancer (GDSC):** In this study [6], the authors map the genomic alterations detected in a large-scale cancer study with the annotations from different cancer cell lines, where they have measured the sensitivity response to several anticancer drugs and linked this response to the genomic information. We downloaded the file TableS4C.xlsx from Iorio and collaborators [6] which contains the significant results of the multivariate ANOVA test used in the experiments. The information in this file is stratified by the different tumor types but also for the global PanCancer set. We extracted the ANOVA results for the PanCancer data set.

## Drug name standardization

We used the PubChem Identifier Exchange Service from PubChem resource [7] (accessed on 15th February 2018) where a list of synonyms is provided for a particular compound. For each of the returned alternative names we selected the first one, that is then used as the standard name. Next step consisted on a manual revision to correct possible inconsistencies in the standardization and to assign a standard name to those cases in which there were no entry in PubChem. To obtain the show name we retrieved the file ligands.csv from the Guide to PHARMACOLOGY [8] version 2017.6 where the International Nonproprietary Name (INN) was extracted. This process was followed by manual curation to revert possible inconsistencies.

Since different databases can use alternative names to mention the same compound, drug names were standardized in order to be consistently integrated in PanDrugsdb. There is no clear consensus for the huge range of available synonyms and, in some cases, they are ambiguous and employed in a wrong way. To avoid the complexity in some of the standardized names we provide an alternative name when showing the final results. That is the INN or the source name when this is not available or is the standardized name is too long (which usually happens with chemical nomenclatures of compounds).

## Gene and drug annotations

**KEGG pathways:** We obtained a list of pathway's codes linked to the involved genes from Kyoto Encyclopaedia of Genes and Genomes (KEGG) pathways database [9]. For this purpose we used the REST API available in the release 85.1 of this database. This list was used to map the genes in PanDrugsdb with the pathways they are involved in.

**Drug Family:** To define the family for the drugs we used two sources. One is the Target-based Classification of Drugs from KEGG (release 85.1). This resource provides a hierarchical classification for each drug with several terms that go from the most general to the most specific. To assign a unique category, we chose the second most general term. The other resource is the classification of drugs established in the Connectivity Map (CMap) [10]. We obtained a file with a list of chemicals considered in the CMap resource and the linked Mechanism of Action from the Repurposing section of Clue.io portal for CMap project (accessed on February 2018).

**Drug status:** We used the drug information from FDA [11] and information about clinical trials [12] (February of 2018). The drug status was manually curated in order to identify the current cancer therapies and the therapies that could be used in a repositioning way. Following categories and subcategories were distinguished (Supplementary Figure S2A):

1. Approved:
  - a. In cancer: When the drug appears approved by the FDA and it is indicated in a cancer treatment. The cancer type and the therapy type were also incorporated into the database.
  - b. In cancer clinical trials: When the drug appears approved by the FDA and it is used in other conditions different from cancer, but it is under study in a cancer clinical trial as a potential treatment.
  - c. Other: When the drug appears approved by the FDA and it is used in other conditions different from cancer.
2. Clinical trials:
  - a. In cancer clinical trials: When the drug does not appear approved by the FDA but it is under study in a cancer clinical trial as a potential treatment.
  - b. Other: When the drug does not appear approved by the FDA but it is under study in a cancer clinical trial for a different pathology from cancer.
3. Experimental: When the drug is in a pre-clinical stage.
4. Withdrawn: When the compound appears as withdrawn in the FDA.
5. Undefined: When the drug name refers to a set of compounds but not to a specific one.

**Pathological area:** This information was obtained from FDA labels, classifying the different indications into at least one of the defined categories in Supplementary Figure S2B. For approved drugs in cancer, compounds are encompassed in groups depending on the anatomic location of the tumor type they are prescribed for (Supplementary Figure S2C).

**Definition of direct target or biomarker gene in the drug response:** In each drug-gene association, the gene can have a different role in the drug response. We call direct target to a gene that contributes to a disease phenotype and can be directly targeted by a drug (small molecule, monoclonal antibody...). For example, EGFR in the use of an EGFR tyrosine kinase inhibitor (TKi). In the other hand, we call biomarker to a gene which genetic status is associated with a drug response by clinical or pre-clinical evidences but its protein product is not the direct target of the drug. For example, MET in the use of EGFR TKi, where amplifications in this gene cause a resistance response to these compounds. To assign the target/biomarker label to each relation we have taken into account the type of information stored in the different databases (Supplementary Table 1). Cancer Commons, CGI, Clarity Foundation Clinical Biomarkers, DrugBank, FDA, Guide To PHARMACOLOGY, My Cancer Genome, NCI, TALC, Tdg Clinical Trial, TEND, TTD from DGIdb or additional sources of monoclonal antibodies, discarding some controlled exception, store associations where the gene is the target of the drug. We added to the target set, those records in TARGET database where the gene is stated as target and also well-known associations described as target gene-drug. The remaining drug-gene associations were labelled as biomarker. In its current version PanDrugsdb supports biomarkers such as gene mutations, amplifications, deletions, gene fusions, alterations in gene expression and promoter methylation modifications reported to drive resistance or sensitivity in response to drugs.

**Resistance/Sensitivity drug response:** This information was obtained from CGI, CIViC, CKB, My Cancer Genome, My Cancer Genome Clinical Trial, Clarity Foundation Biomarkers, DoCM, FDA, NCI, TALC, TARGET, GDSC and CTRP that store drug response information. Relations for which information was not available were indicated as sensitivity (Supplementary Table 1).

**Molecular alteration type:** The definition of the molecular alteration type of the drug-gene associations was performed by a combination of the existing information in CGI, CIViC, CKB, Clarity Foundation Biomarkers, DoCM, FDA, NCI, OncoKB and TARGET (Supplementary Table 1) along with a manual curation based on the existing knowledge in the literature. The types of driven molecular alterations included were missense mutation, amplification, deletion, gene fusion, gene expression dysregulation and promoter methylation.

### **Data source integration**

Drug-gene association data was manually downloaded from the different resources. This data was automatically parsed using a combination of custom python and perl scripts when possible, or mixed with a manual intervention in those cases that require a more exhaustive edition. This step created a tabular plain text for each of them. Then, using a perl module, these files were combined, and for each drug-gene interaction, all the corresponding annotations were incorporated and a pre-computed drug score was calculated for each of them.

All records in this database were joined (Supplementary Figure S2D) keeping only one record for the quartet gene-source-original name-standard name, because of the duplications that can appear in some files. In most of the sources, there are cases in which there are several records with different original names that converge in a unique standard name. These records were kept and they were not unified to control the presence of inconsistencies in the drug name standardization process. In any case, they are shown as a unique assignment when suggesting drugs. We finally obtained the PanDrugsdb with 9092 unique drugs, 4804 unique genes and 43909 unique drug-gene interactions. Distribution of this final drug-gene associations is represented in (Supplementary Figure S2E).

### **Gene Score (GScore) calculation**

The Gene Score value (GScore) ranges between 0 and 1 and it allows the prioritization at gene level taking into account the biological relevance of the gene in carcinogenesis and the therapeutic actionability. GScore calculation depends on the provided input type (gene list or VCF file) and is weighted depending on the level of gene association with cancer. If PanDrugs input is a VCF file the GScore will be computed taking into account the information provided by the variants located in each gene resulting in a gene prioritization based on variant information.

1. *List of gene symbols:* The GScore is calculated as shown in the global formula expressed in Supplementary Figure S3A and Supplementary Table 2. To perform

this calculation we consider four evidences: i) the frequency at which the gene appears in different tumors, ii) the probability of being a cancer driver, iii) the gene essentiality based on RNA silencing studies in cancer cell lines, iv) and the oncogenic score of genes based on the integrative analysis of OncoScape [13]. Each of these evidences has an associated weight.

- a. To evaluate the frequency of appearance in carcinogenic processes we used two information sources. On one hand, we use the list of genes in the Cancer Gene Census (CGC) of COSMIC v84 [14] and also the TumorPortal resource [15], assigning a different weight according to the frequency at which the gene is altered in any tumor type. Within TumorPortal, they establish three categories: *Highly significantly mutated*, *Significantly mutated* and *Near significance* ordered from high to low mutation frequency. A decreasing weight is provided for each of them.
  - b. To score the probability of being a tumor driver, we use the information obtained from Tamborero et al. [16] that identify a set of potential tumor drivers using mutational information from TCGA. The weight given by this component will depend on the assigned probability in the study (*High Confidence Driver* or *Candidate Driver*).
  - c. To calculate the essentiality score genes were ranked by the negative Pearson's correlation between the phenotype value calculated by ATARiS algorithm [17] and the gene expression value from all the cancer cell lines in common from the two datasets (n=216) (the Project Achilles [18] and the CCLE data [19]). Thus, the higher is the gene expressions and the lower is the phenotype values in cancer cell lines for a given gene, the more essential is the gene. A similar approach was used in the original study by Shao et al. with 83 cancer cell lines. The resulting ranking of correlations were transformed using a min-max normalization into a continuous 0-1 range.
  - d. OncoScape is a method to identify cancer candidate genes by the integration of different molecular data from 11 cancer types. It integrates information about gene expression data, somatic mutations, DNA copy-number variation, methylation and data from shRNA knock-down screens, this last particularly interesting for our approach. The scores provided in OncoScape were used as a base for our GScore weighting.
2. *VCF file*: We first execute the variant effect predictor of ensembl (VEP) [20]. This tool provides annotations and predictions for the variants that we after enrich with additional information. We keep the variants with an important impact in the transcriptional process (VEP consequence equal to transcript\_ablation, splice\_donor\_variant, splice\_acceptor\_variant, stop\_gained, frameshift\_variant, stop\_lost, start\_lost, transcript\_amplification, inframe\_insertion, inframe\_deletion, missense\_variant, protein\_altering\_variant, splice\_region\_variant, incomplete\_terminal\_codon\_variant and stop\_retained\_variant). The score for each variant is called VScore and is computed as shown in the Supplementary Table 3. Each contribution is conditioned by the role assigned to the gene. To decide the role we use the consensual information of CGC and the prediction made by oncodriveROLE [21]. The genes not present in any of them or that have opposite role labels in each resource are evaluated to see the consequence of

the variant. If the consequence is a stop gain, stop lost, a frameshift or a splice alteration, they are labeled as tumor suppressors. The remaining variants for which we cannot assign a label are graded in the same way as oncogenes. Among all the calculated VScores of a particular gene, we select the highest and establish it as the GScore. Only the most relevant transcripts according to the criteria established by APPRIS [22] are taken into account in the selection.

3. *Ranked list of genes*: The input can be a ranked list of genes based on some experimental results, as for example, data originating from differential expression studies. The provided values are normalized to the 0-1 scale using min-max scaling.

### **Drug Score (DScore) calculation**

The Drug Score (DScore) allows to prioritize the suggested therapies and reflects the suitability of a treatment according to the genomic profile. It goes from -1 to 1 with the negative values corresponding to resistance and positive values corresponding to sensitivity (Supplementary Figure S3B). We have a precomputed DScore in the database which is based on each single drug-gene relation. In its calculation we first take into account the use of the drug in cancer, then the approval status of the drug and finally the definition of the gene as a target or marker in the relation with the drug. Experimental compounds have a different score assignation, but they rank below drugs in another status, giving more relevance to target than to biomarker genes.

### **PanDrugs assignation process**

PanDrugs gene-drug assignation process consists on the generation of a ranked list of drugs associated to the input gene list. In this assignation process PanDrugs suggests drugs in two ways: 1) directly and 2) following pathway-member paradigm. In the direct way PanDrugs search for drugs against direct targets or biomarkers. The pathway member approach allows to expand the therapeutic options and consists on gene-drug assignations where the drug target is a gene located downstream to the altered one.

For pathway member search we built a catalogue of biological pathways involved in a variety of processes related to cancer. To do so, we extracted from the modelled pathways available in hiPathia [23] all the possible subpathways comprising four nodes at maximum. Only those nodes categorized as 'gene' were considered, while other nodes (i.e. glycans) were discarded. Treatment options suggested for these nodes(genes) depends on: i) the functional role of the nodes in cancer (oncogene, tumor suppressor gene or dual role of oncogene and tumor suppressor gene) and ii) the type of interaction (activation or inhibition) with the child node.

During the assignation process the precomputed DScore is adjusted to take into account the information provided by the input data collectively. We define the 'collective gene impact' by assigning a higher DScore to that drug capable to target the highest number of genes found in the input list. We also consider the number of expert curated databases supporting a particular drug-gene relationship. Indirect drug-gene relationships are penalized with respect

to direct ones unless a ‘biomarker’ evidence supports the association. Experimental drugs have no DScore readjustment maintaining the precomputed DScore and only penalizing the indirect cases (Supplementary Figure S3C). If one particular drug has a sensitivity response due to one gene, but a resistance response due to another, the drug response assignment will be “Both”. The drug reference assignment employs the sign of the highest DScore in absolute value (max |DScore|). This means that the drug response with the highest evidence is the one that is going to be selected to allocate the drug in the sensitivity or resistance area by default. Full details regarding sensitivity and resistance for each particular case are downloadable and accessible in PanDrugs through pull-down menus.

## **PanDrugs software implementation**

The back-end application is in charge of (i) storing gene and drugs data, (ii) perform and manage genomic variants analyses and (iii) allow external applications to access the data and services through a public REST API. The database is stored in the MySQL RDBMS. Variant analyses are performed with a Perl script which computes the scores for the mutated genes from user-provided VCF file. A previous annotation step with the VEP release 90 is performed. In order to manage multiple variant analysis simultaneously a Java scheduling program using a thread pool was implemented. The REST API allows external programs to query the PanDrugsdb over HTTP. It was implemented in Java with the JAX-RS API. Both the scheduling program and the REST API service runs in a single Java EE application in Apache Tomcat 8.

The front-end application is in charge of get user queries, communicate with the backend REST API and display results in an user-friendly interface. This application is implemented with AngularJS 1.4. The Highcharts library is used for the Gene and Drugs score chart and the D3 library for the visualization of PanDrugs across TCGA tumoral landscape.

## **RESULTS AND DISCUSSION**

### **Analysis in TCGA data**

Mutations and CNVs data employed this study was obtained from 20 different tumor types available in TCGA project. The workflow followed for this analysis is represented in Supplementary Figure S5.

For mutation data, we took the MAF files from synapse syn1729383 (Supplementary Table 4). In particular, we selected the cleaned\_filtered.maf file with the whole filtered alterations for 19 tumor types (COAD and READ are concatenated) and to speed up the process we removed those genes that do not appear in PanDrugsdb. Then, we calculated the GScore for each gene based on variant information. To do this, we ran the VEP release 90 using the corresponding cache files of the human genome (hg19 version) over an ensembl-format converted file version of the mutations of the MAF files. From the resulting files, we kept those variants with high impact consequence (transcript\_ablation, splice\_donor\_variant, splice\_acceptor\_variant, stop\_gained, frameshift\_variant, stop\_lost, start\_lost, transcript\_amplification, inframe\_insertion, inframe\_deletion, missense\_variant,

protein\_altering\_variant, splice\_region\_variant, incomplete\_terminal\_codon\_variant and stop\_retained\_variant) and we added the annotations used to compute our VScore. Then, from these VScores, the GScore for each gene was established as described for VCF input files. Databases and the corresponding versions used in this process are indicated in Supplementary Table 5.

For CNVs, we took the file `gistic.all_thresholded.by_genes` from synapse (Supplementary Table 4) for each of the tumor types. The GScore was calculated in this case according to a gene level criteria as described above for a list of genes. We kept those records with CNVs defined as -2 (homozygous deletion), 2 (high level amplification), -1 (heterozygous deletion).

In order to establish a suitable threshold to filter genomic events with an unclear functional effect, we performed several filtering steps using increasing values of GScores for both mutations as CNVs. As can be observed in the Supplementary Figure S6A, above a GScore of 0.4 the number of genes with a mutation event drops. Thus, establishing a gene score threshold above 0.4 would remove the bulk of alterations that in principle would not contribute to the pathogenic process. For CNVs the decrease was observed at low threshold values of GScore and it presents a more continuous pattern in the reduction of the number of genes.

Looking at the same distribution and considering the number of patients instead of the number of genes (Supplementary Figure 6B), the selection of this threshold would still cover a great number of patients. The selected CNV events would be present in a large number of patients in comparison with SNVs despite of being significantly reduced.

Taking into account the complementarity of both events (Supplementary Table 6 and Supplementary Figure S6C), from an initial number of 7096 cases, more than 6000 would still maintain some mutational and/or CNV events that could be used to identify potential therapies. Distribution of these events affecting genes EGFR and KRAS across the 20 tumor types of these study can be seen in the (Supplementary Figure S7)

Mutations and CNV data was integrated and affected genes were queried against PanDrugs for each TCGA patient. In this query oncogenes and tumor suppressor genes were managed in a different way. Treatments for oncogenes were provided directly (direct targets) and through pathway members (for KRAS mutant patients MEK inhibitors proposed as pathway members were excluded). However, for tumor suppressor genes, only indirect treatments through pathways members were considered. To determine the behaviour of the gene as oncogene or tumor suppressor gene in this context, where mutational and CNV data are integrated, we considered additional guidelines. If there was a mutation with a role information different from unclassified (when no assumption could be made about the role) in an affected gene, this role was assumed as the role of the gene. Otherwise, and when there were CNVs affecting the gene, it was considered as oncogene if it was affected by an amplification or tumor suppressor gene if it was affected by a deletion. If after this process the role of the gene remained unclassified, the label established in most of the cases for that gene in the patients for that particular tumor type was assigned. Those genes for which it was not possible to determine a role of oncogene or tumor suppressor were queried in both direct and pathway member. Most frequently altered genes suggested for treatment by PanDrugs in TCGA patients are indicated in Supplementary Table 7.

We compared PanDrugs performance applying our methodology to the TCGA cohort previously used in Rubio-Perez C. et al. study [24]. To analyze this data, we focused on the common patients in our TCGA analysis and the core cohort exclusively integrated by the TCGA cases of the Rubio-Perez C. et al. analysis. To that end, we used the information provided in the supplementary material S4D from Rubio-Perez et al manuscript to make the comparison and extracted those cases present in our TCGA study. To make both sets comparable we reduced the alterations detected with our processing to those considered drivers in each tumor type. Drug assignments were then recalculated and represented without filtering by GScore. Only those drugs without evidences of resistance and supported by at least two sources are contemplated.

### ***Application in a cancer case study***

**WES analysis:** Tumoral and normal samples of this patient were sequenced to identify tumor-specific sequence alterations. Exome variant analysis and biological impact predictions were performed by RUBioSeq software [25] using default parameters for somatic variation analysis. In detail, sequencing data were first analysed by FastQC for quality control inspection and then aligned to the human reference genome (GRCh37) using Burrows-Wheeler alignment (BWA-MEM) [26]. Somatic variants were identified using the HaplotypeCaller available at the GATK [27]. For variant calling we used GATK HaplotypeCaller with default parameters for filtering. Biological impact predictions for detected variants were obtained from VEP. In order to estimate the biological impact of the missense mutations, we used different algorithms (i.e. SIFT, PolyPhen-2, CONDEL, Pfam, InterPro, etc.). Then, other GScore annotation sources (cancer essentiality, relevance in cancer, frequency and clinical implication) are incorporated and GScore itself is calculated to rank selected somatic variants.

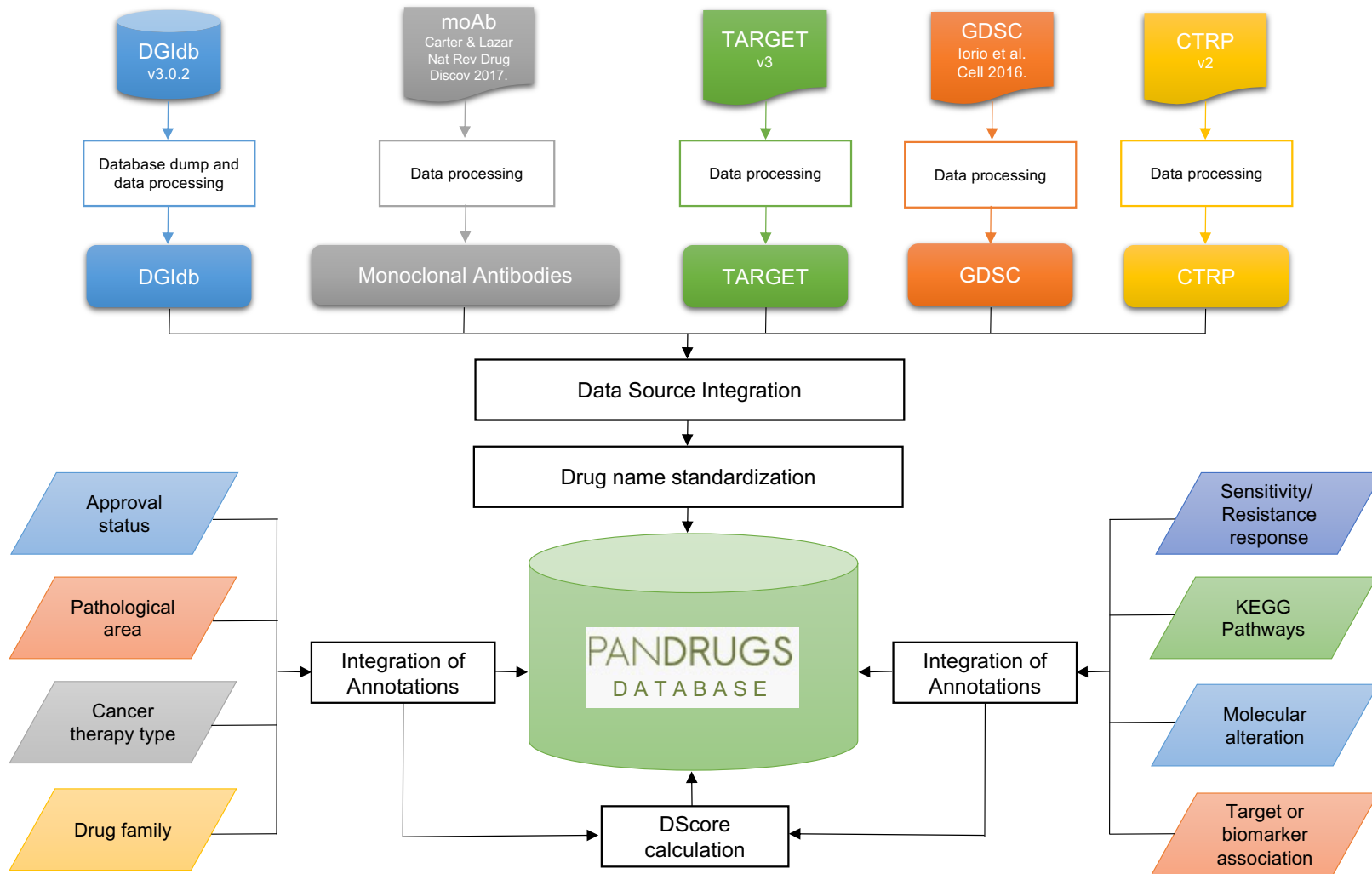
**Drug efficacy testing in PDX:** We evaluated the antitumor activity of a MEK inhibitor (MEKi), a PI3K inhibitor (PI3Ki), an mTOR inhibitor (rapamycin), a multi- BCR/ABL and Src family tyrosine kinase inhibitor (dasatinib) and a HER2 inhibitor (lapatinib), in a low passage lung cancer (squamous cell carcinoma) patient-derived xenograft (PDX) mouse model. Data collected from this study includes animal weights, tumor dimensions and daily observations; this information was used to determine anticancer activity based on tumor growth inhibition or regression. The designated endpoint for this study was a mean control tumor volume of approximately 1.5 cm<sup>3</sup>. Animals were implanted with tumor fragments harvested from host animals and the study initiated up to 50 days later at an average tumor volume of approximately 200 mm<sup>3</sup>. No tumor burden was associated with this model based on lack of weight loss or animal morbidity in the control group.

In this study, the doses used with all drugs were well tolerated with no significant ( $\geq 20\%$ ) weight loss reported in any group. Statistically significant ( $p < 0.05$ ) tumor growth inhibition was reported for MEKi and PI3Ki treatments compared with control group at the time point considered.

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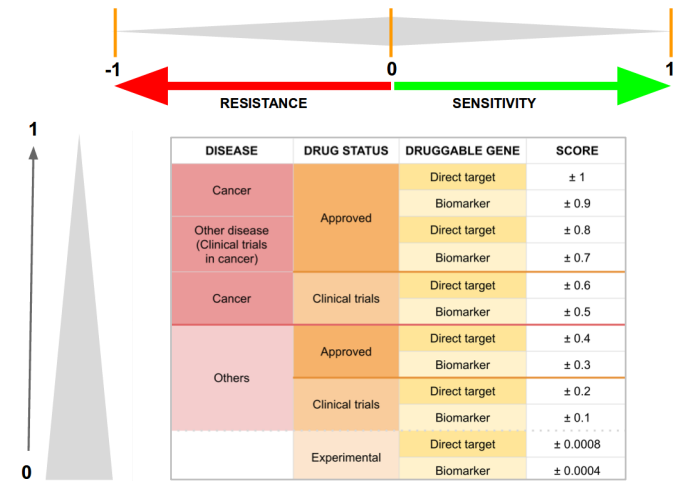
**Supplementary Figure S1.** PanDrugs database integration schema.



A)

$$\begin{aligned}
 \text{GScore} = & \left[ 0.4 * \text{Essentiality Score} \right] + \left[ 0.3 * (\max \{ \max \text{ OncoScape score for oncogene in different tumor types, max OncoScape score for tumor suppressor gene in different tumor types} \} \text{ normalized between 0 and 1}) \right] \\
 & + \left[ \begin{array}{l} \text{Presence in cancer (Tumor Portal)} \\ 0.1 \text{ (If Highly significantly mutated)} \\ 0.05 \text{ (If Significantly mutated)} \\ 0.025 \text{ (If Near Significance)} \end{array} \right] + \left[ \begin{array}{l} \text{Presence in cancer (CGC)} \\ 0.1 \text{ (If present in CGC)} \end{array} \right] + \left[ \begin{array}{l} \text{cancer driver} \\ 0.1 \text{ (If High Confidence Driver)} \\ 0.05 \text{ (If Candidate Driver)} \end{array} \right]
 \end{aligned}$$

B)



C)

**Approved and Clinical trials drugs:**

**Pre-computed DScore value** = [Cancer + Drug Status + Druggable gene type]\*[if resistance (-1)]

**Collective gene impact** = # genes (max. 9) + [-1(if pathway member)]

**Database factor** = # expert curated sources (max. 9)

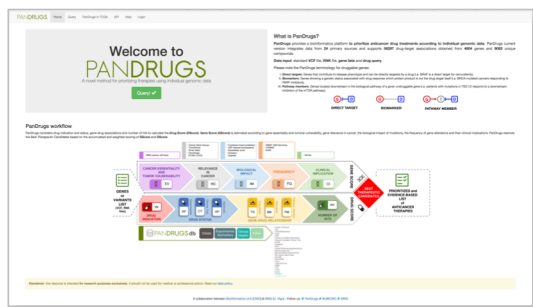
DScore = max{Pre-computed DScore value} - 0.1 + (0.01 \* Collective gene impact) + 0.001 + (0.001 \* Database factor)

**Experimental:**

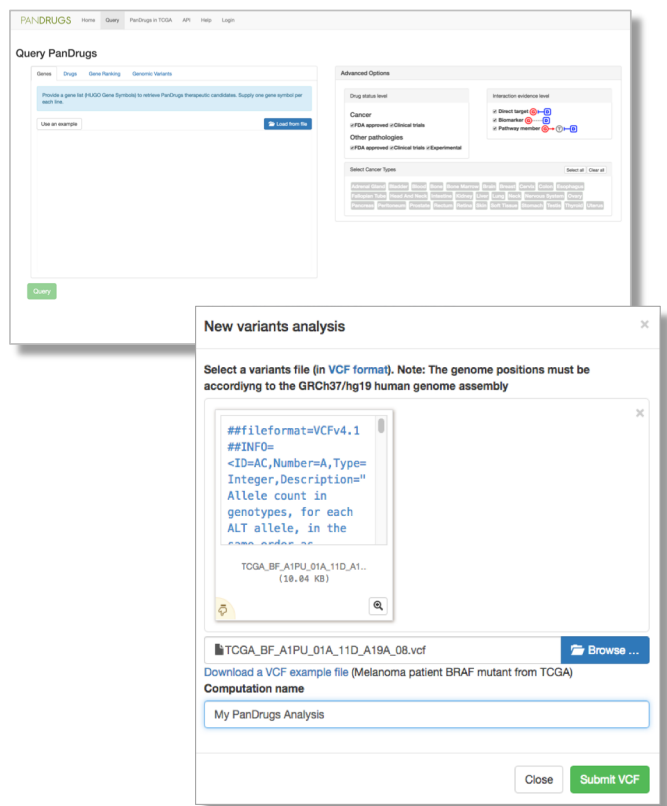
DScore = max{Pre-computed DScore value} - 0.0002 (if indirect)

**Supplementary Figure S3.** A) GScore calculation. B) Features contributing to pre-computed DScore calculation. C) DScore calculation.

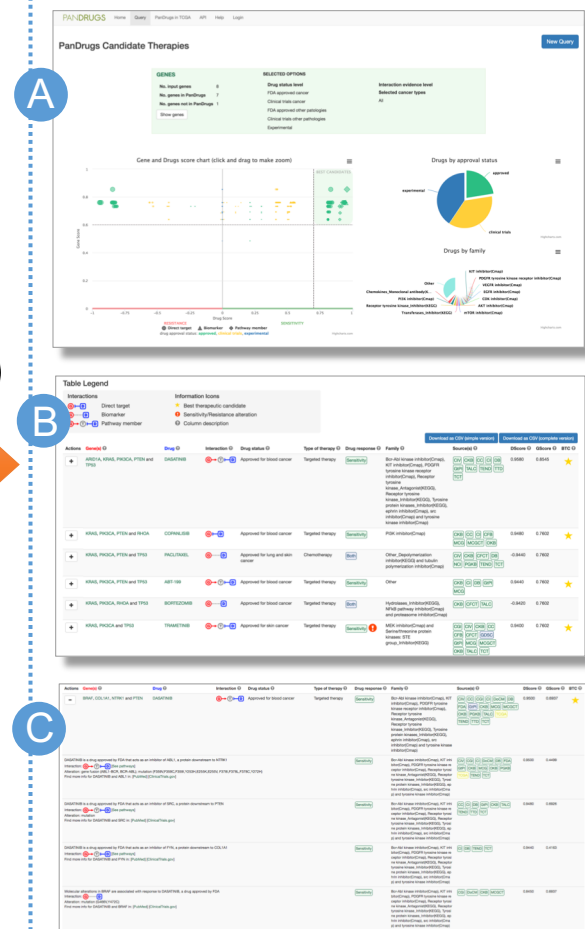
# MAIN INTERFACE



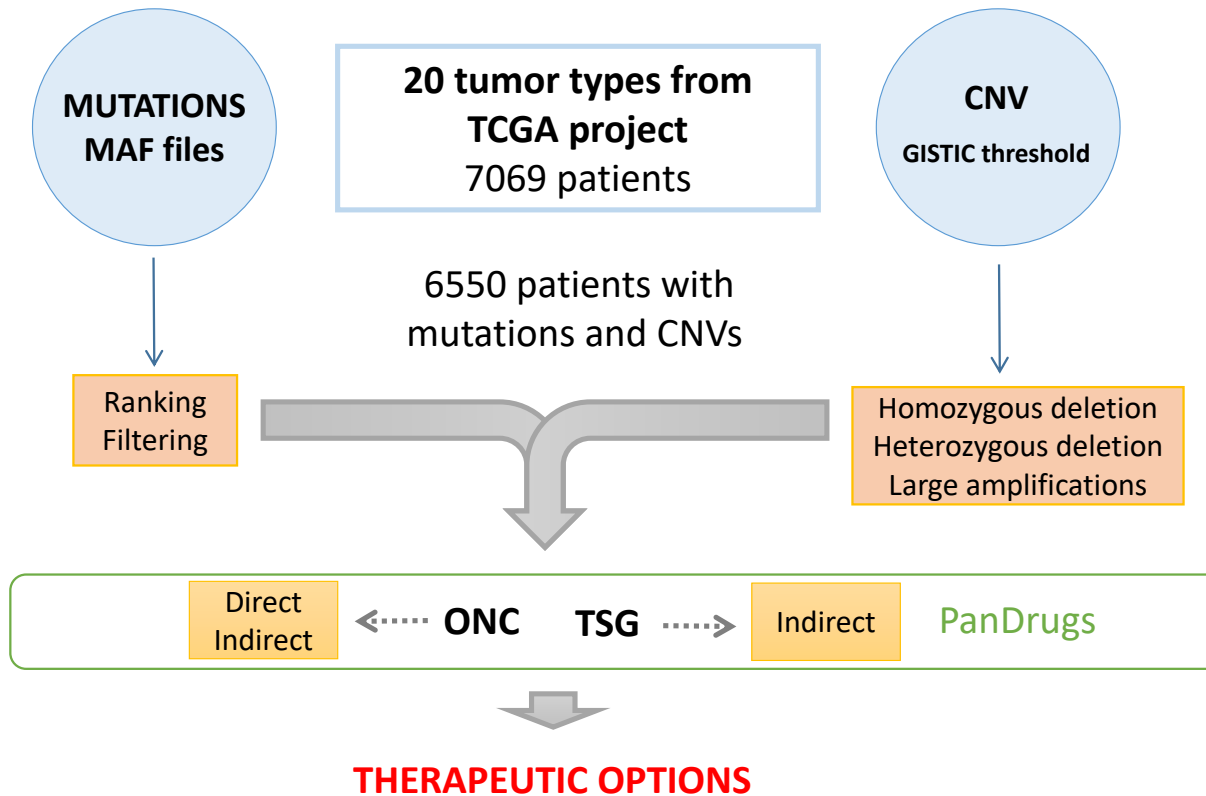
# QUERY INTERFACE



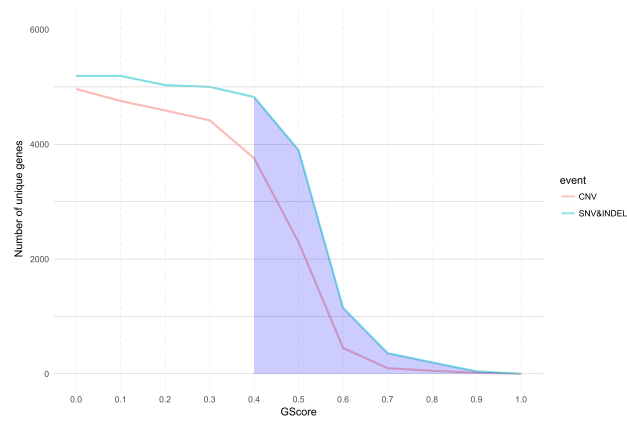
# RESULTS INTERFACE



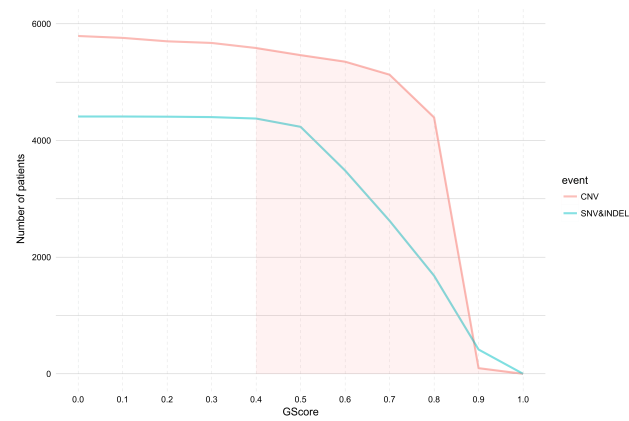
**Supplementary Figure S4.** PanDrugs workflow for standard VCF file containing variants from a BRAF-mutated melanoma patient from TCGA (TCGA-BF-A1PU-01A-11D-A19A-08). PanDrugs results interface includes: A) Score chart displaying the best candidates drugs proposed and B) summary table listing the treatments suggested. C) Summary table may be deployed to show detailed information for treatments (i.e. Dasatinib). Additional information about genes, annotation sources, drug characteristics and type of gene-drug interaction is displayed for each drug-gene association.



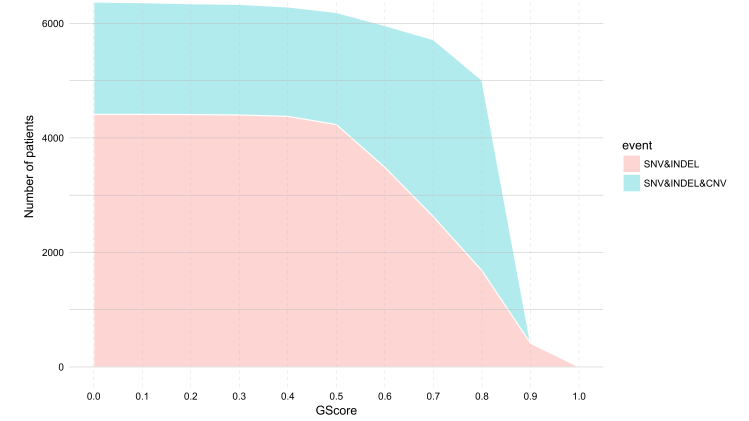
**Supplementary Figure S5.** Flow chart of the TCGA analysis. (ONC: Oncogenes; TSG: Tumor Suppressor Genes.)



A)

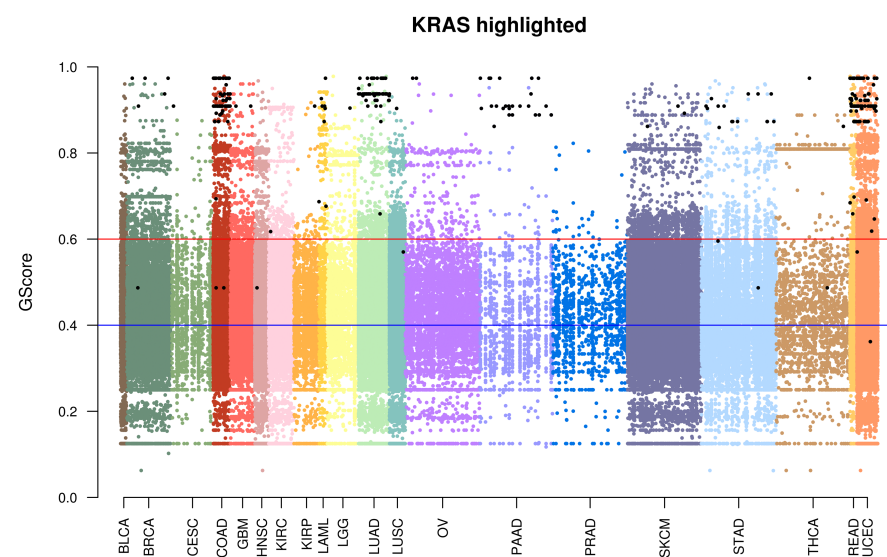
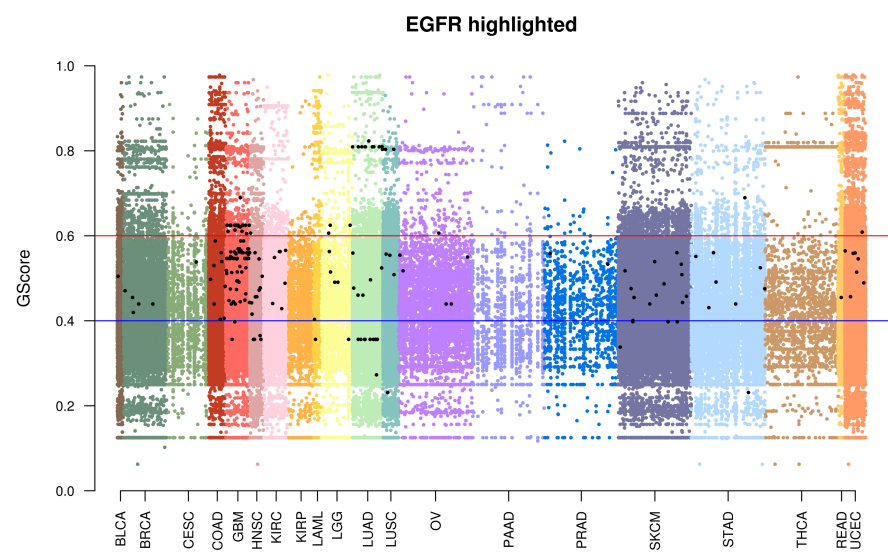


B)



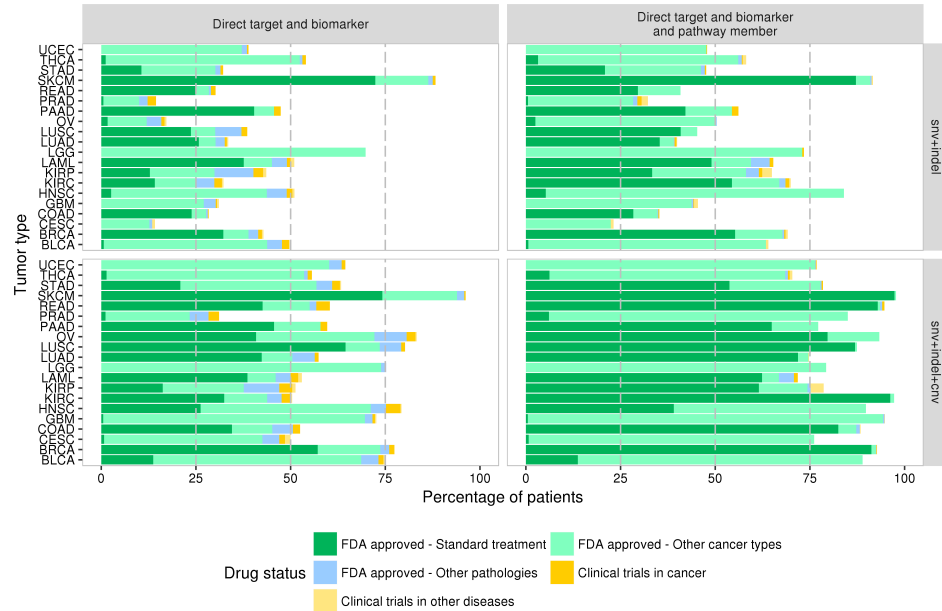
C)

**Supplementary Figure S6.** A) Distribution of altered genes and their corresponding GScore value in TCGA data (SNVs, indels and CNVs). B) Distribution of patients and the corresponding accumulative GScore value in TCGA data (SNV, indels and CNVs). C) Distribution of patients and the corresponding accumulative GScore value in TCGA data (SNV+indels and SNV+indels+CNV).

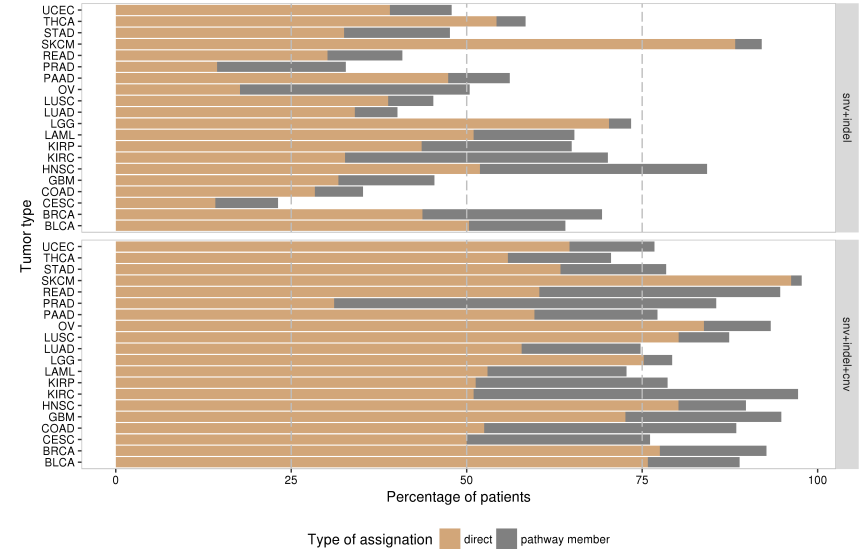


**Supplementary Figure S7.** Manhattan plots for GScore values across 20 TCGA tumor types. EGFR and KRAS GScores are highlighted in black.

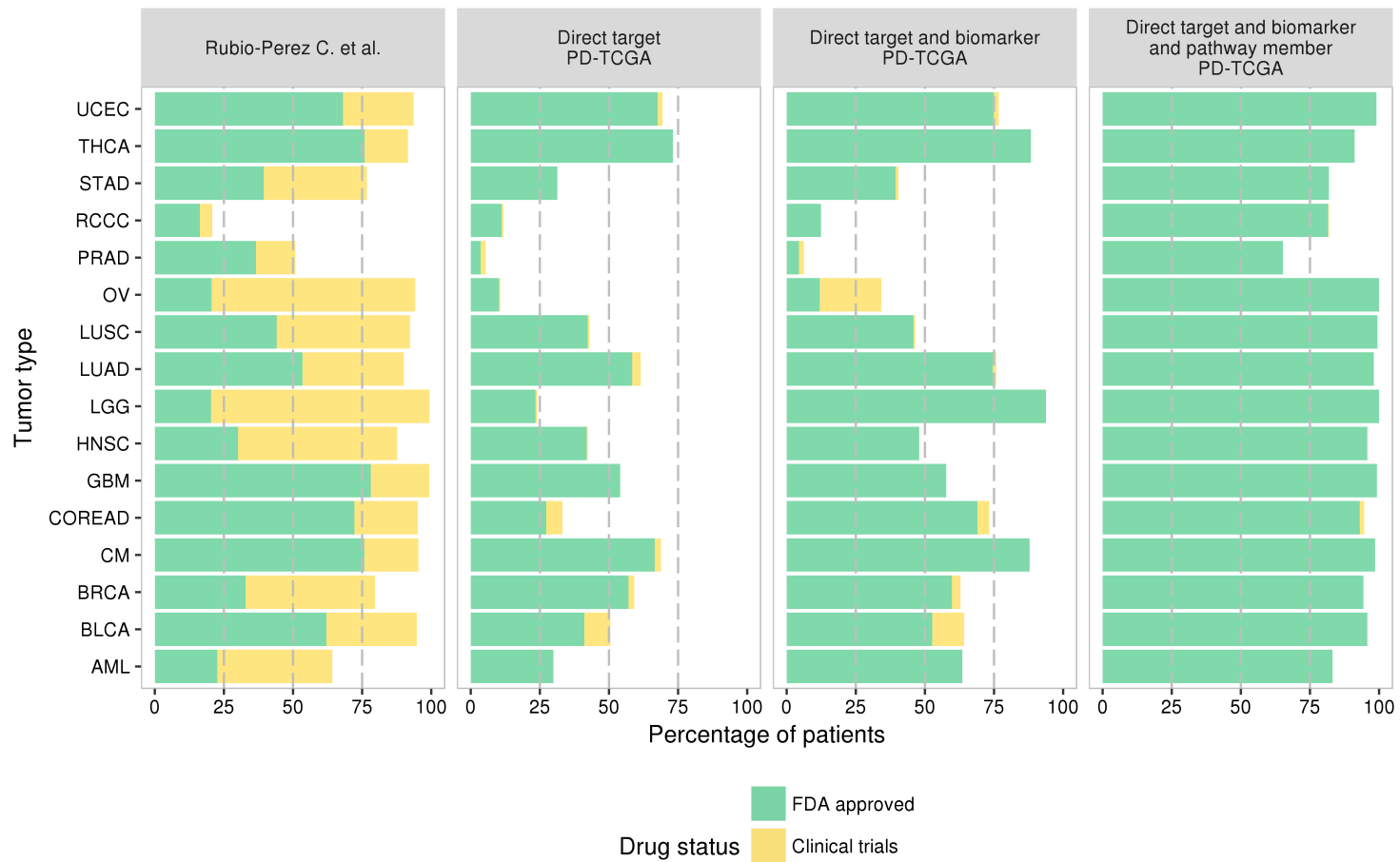
A)



B)



**Supplementary Figure S8.** A) TCGA patients under clinical trials or treated with approved drugs. Three different levels of evidence for these treatments are shown: (i) treatments that directly target the affected gene, (ii) treatments indicated to direct gene targets and biomarkers and, (iii) treatments indicated to direct gene targets, biomarkers and downstream pathway members. The bar-chart considers separately genes affected only by single nucleotide variants and indels (top panel) and genes affected by single nucleotide variants, indels and CNVs (bottom panel). B) TCGA patients treated with drugs that directly targets an altered gene (brown bars). Grey bars represent the increase in the number of TCGA patients treated when pathway members are also included. The bar-chart considers separately genes affected only by single nucleotide variants and indels (top panel) and genes affected by single nucleotide variants, indels and CNVs (bottom panel).



**Supplementary Figure S9.** Current in silico prescription methods based on the genomic analysis of known cancer genes may be enriched by PanDrugs pathway member approach. TCGA comparative analysis showed that PanDrugs path member approach clearly expands the number of cancer patients who can be potentially benefited with FDA approved treatments (green bars).

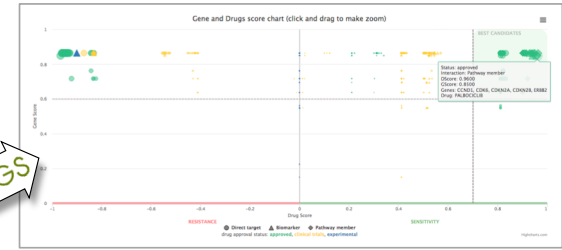
A)

EGFR FGF19  
MET ERBB2  
AKT1 FGF3  
RNF43 CDKN2B  
CCND1 CDKN2A  
PTPRD TP53  
CDK6



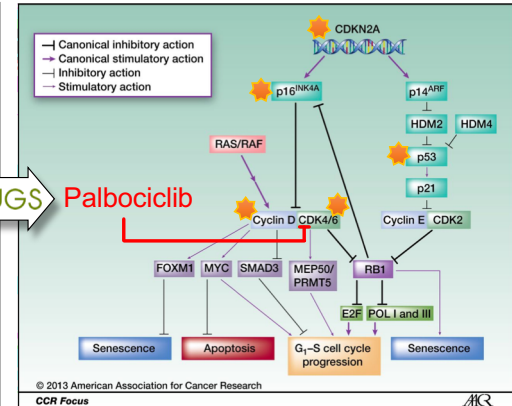
**NSCLC, TCGA-38-4629**

Gender: Male  
Diagnosis Age: 68M  
Days to Last Followup: 864  
Death from Initial Pathologic Diagnosis Date: 864  
Stage T: IIB  
Stage N: n0  
Stage M: m0



Actions	Gene(s)	Drug	Drug status	Type of therapy	R/S	Interaction	Family	Source(s)	DScore	GScore	BTC
	CCND1, CDK6, CDKN2A, CDKN2B and PTPRD	PALBOCICLIB	Approved for breast cancer	Targeted therapy	Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CancerCommons, ClearityFoundationBiomarkers, ClearityFoundationClinicalTrial, DrugBank, GuideToPharmacologyInteractions, MyCancerGenome, MyCancerGenomeClinicalTrial and TTD	0.9570	0.8500	★
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CancerCommons, ClearityFoundationBiomarkers, ClearityFoundationClinicalTrial, DrugBank, GuideToPharmacologyInteractions, MyCancerGenome, MyCancerGenomeClinicalTrial and TTD	0.9570	0.8500	
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CancerCommons, ClearityFoundationClinicalTrial, DrugBank, GuideToPharmacologyInteractions, MyCancerGenome, MyCancerGenomeClinicalTrial and TTD	0.9560	0.5620	
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	ClearityFoundationBiomarkers	0.8520	0.8500	
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	ClearityFoundationBiomarkers and GDSC	0.8520	0.7829	
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	GDSC	0.8510	0.2250	
					Sensitivity	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	CDK inhibitor(Cmag) and Serine/threonine protein kinases. CMGC group. Inhibitor(KEGG)	GDSC	0.8510	0.1500	

PANDRUGS



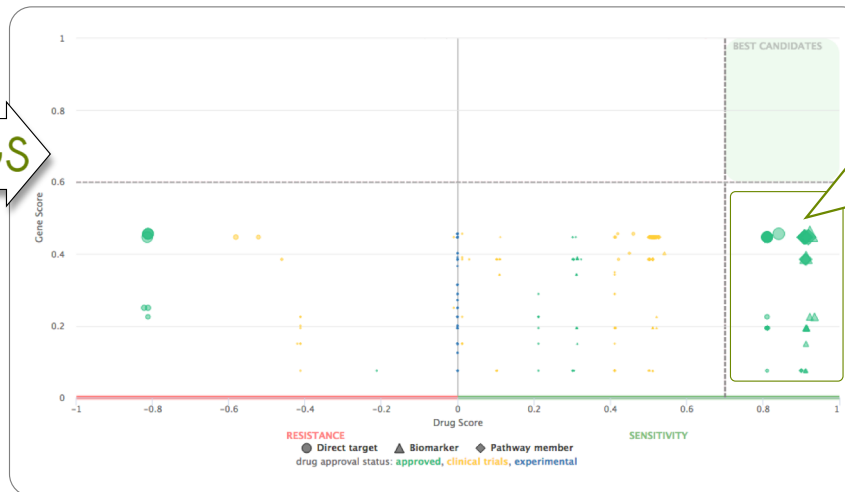
B)

Martincorena I. et al. *Cell*, 2017.

**83**  
**Novel Cancer Genes**  
✓ CGCv73 = FALSE  
✓ Cancer5000-S = FALSE

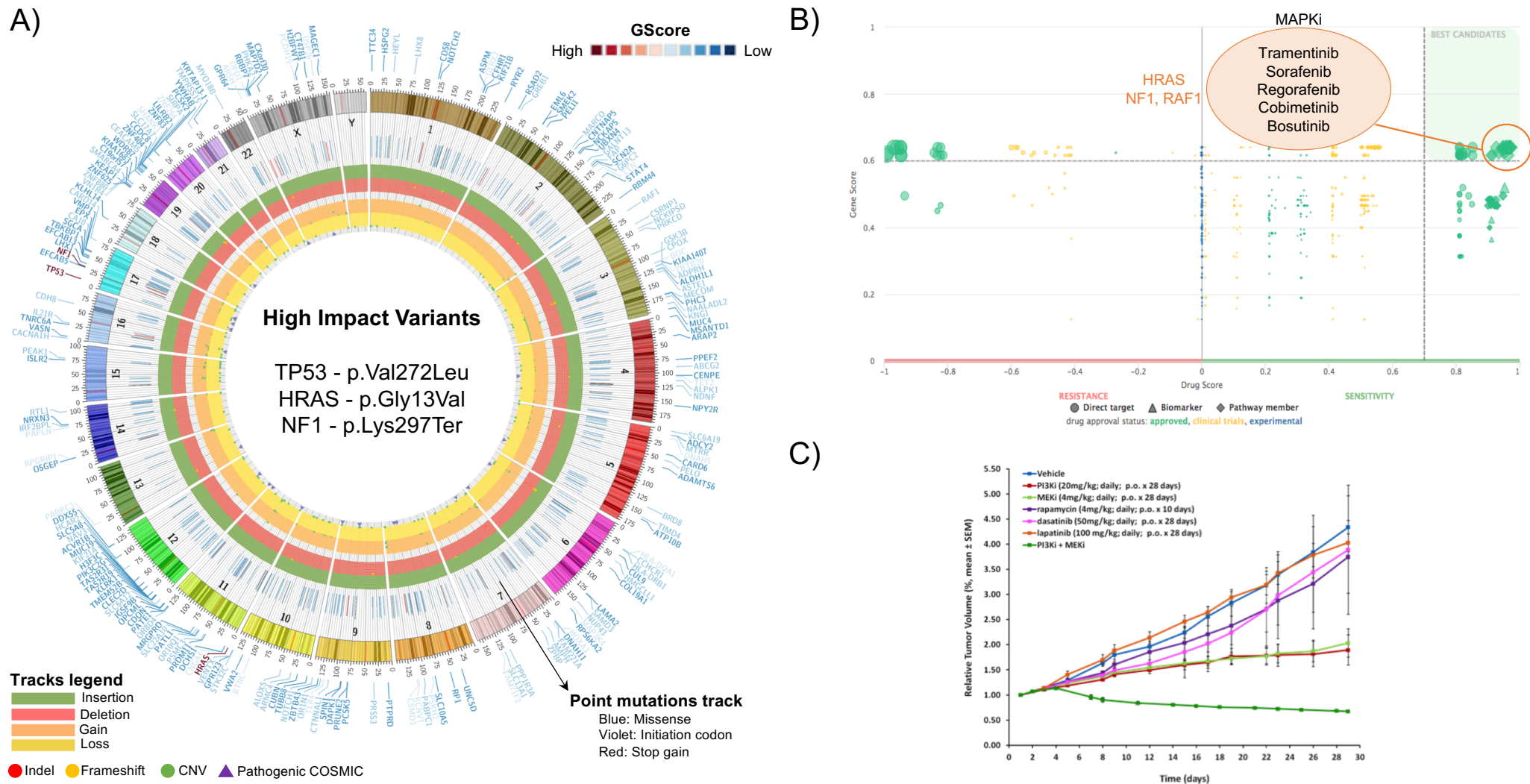
PANDRUGS

**Supplementary Figure S10.** (A) PanDrugs analysis for NSCLC patient (TCGA-38-4629). Oncogenes (red) and tumor suppressors genes (blue) annotated in cBioportal were selected to make tool comparison viable. PanDrugs prescribes Palbociclib since (i) CDK6 is direct target, (ii) CCND1, CDKN2A and CDKN2B are biomarkers and, (iii) CDK4 is a downstream pathway member gene. (B) PanDrugs analysis for novel cancer genes proposed by Martincorena and colleagues. 13 genes showed DScore > 0.7 and were proposed as candidate targets. Low GScore in these genes is explained by the lack of clinical evidence and cancer annotations associated to them.



CDK4 pathway adapted from Sheppard KE and McArthur GA. *Clin Cancer Res* 2013; 19(19): 5320-8.

**DScore >0.7**  
**32 gene-drugs associations**  
**13 genes:**  
NIPBL, TG, TOP2A, ERBB4, POM121L12, ACVR2A, RARG, MAP2K7, PPP3CA, CYP11B1, LATS2, RPS6KA3, DAZAP1



**Supplementary Figure S11.** PanDrugs has been employed to predict treatments in a PDX model of brain metastasis in advanced squamous cell lung carcinoma (Stage IV). A) Landscape of small genomic alterations detected in the PDX model. B) Drug assignments obtained from PanDrugs execution. C) Patient-derived xenograft tumor growth inhibition measure under drug efficacy test.

Source	Source provider	# initial records	# processed records	Direct Target /Biomarker	Sensitivity/Resistance	Alteration type	Expert curated
CancerCommons	DGldb	104	104	yes			yes
CGI	DGldb	309	309	yes	yes	yes	yes
ChEMBLInteractions	DGldb	7695	7558				
CIViC	DGldb	534	534		yes	yes	yes
CKB	DGldb	1412	1403		yes	yes	yes
ClarityFoundationBiomarkers	DGldb	148	148	yes	yes	yes	yes
ClarityFoundationClinicalTrial	DGldb	178	175				yes
DoCM	DGldb	72	72		yes	yes	yes
DrugBank	DGldb	7805	7723	yes			
FDA	DGldb	245	244	yes	yes	yes	yes
GuideToPharmacologyInteractions	DGldb	7672	7613	yes			yes
MyCancerGenome	DGldb	814	782	yes	yes		yes
MyCancerGenomeClinicalTrial	DGldb	319	303		yes		yes
NCI	DGldb	4298	4287	yes	yes	yes	yes
OncoKB	DGldb	155	155			yes	yes
PharmGKB	DGldb	1274	1245				
TALC	DGldb	492	486	yes	yes		yes
TdgClinicalTrial	DGldb	4155	4085	yes			yes
TTD	DGldb	1829	2210	yes			
TEND	DGldb	2233	1822	yes			yes
moAb	moAb	605	605	yes			yes
TARGET-CGA	TARGET	74	72	yes	yes	yes	yes
CTRP	CTRP	397270	13041		yes		
GDSC	GDSC	1323	1321		yes		
<b>Total</b>		<b>441015</b>	<b>56297</b>				

**Supplementary Table 1.** Gene-drug records in PanDrugsdb for each annotation source (redundant records are included). The table shows the number of initial records, the number of records after PanDrugsdb processing and the additional information provided.

<b>Feature</b>	<b>Weight</b>	<b>Value</b>	<b>Score</b>
<b>Essentiality Score</b>	40%	Computed Essentiality Score	[0 - 1]
<b>OncoScape Score</b>	30%	max{max{OncoScape score for oncogene in different tumor types}, max{OncoScape score for tumor suppressor gene in different tumor types}} normalized between 0 and 1	[0 - 1]
<b>Gene annotated in TumorPortal</b>	10%	Highly significantly mutated	1
		Significantly mutated	0.5
		Near significance	0.25
		No annotation	0
<b>Gene annotated in Cancer Gen Census (COSMIC)</b>	10%	Yes	1
		No	0
<b>Driver Gene</b>	10%	High confidence driver	1
		Candidate driver	0.5
		No annotation	0

**Supplementary Table 2.** Features involved and their corresponding weights in PanDrugs GScore calculation for non-ranked lists of genes.

Feature	Value	Score addition ONC	Score addition TSG
Score prediction by PolyPhen	> 0.435	+ 0.125/3	+ 0.125/3
Score prediction by Sift	<= 0.05	+ 0.125/3	+ 0.125/3
Score prediction by CONDEL	> 0.468	+ 0.125/3	+ 0.125/3
COSMIC	Pathogenic by FATHMM prediction	+ 0.125/3	+ 0.03125
Frequency of mutation in COSMIC	>= 100	+ 0.125/3	
	< 100	+ (0.125 / 3) * (log(mutation frequency) / log(maximum mutation frequency))	
Frequency of gene in COSMIC	>= 100	+ 0.125/3	+ 0.03125
	< 100	+ (0.125 / 3) * (log(gene frequency) / log(maximum gene frequency))	+ 0.03125 * (log(gene frequency) / log(maximum gene frequency))
VEP consequence	stop gain frameshift missense inframe insertion inframe deletion	+ 0.125	+ 0.125
GMAF	< 1	+ 0.125/2	+ 0.125/2
EXAC	< 1	+ 0.125/2	+ 0.125/2
DOMAINS	Listed as relevant in cancer <sup>1</sup> or previous last protein domain (in stop-gained or frameshift)	+ 0.125	+ 0.125
CLINVAR	Within a domain in other circumstances	+ 0.125/2	+ 0.125/2
	Pathogenic with zygosity data	+ 0.125	+ 0.125
ZYGOSITY (when available)	Pathogenic without zygosity data	+ 0.250	+ 0.3125
	Homozygous	+ 0.125	+ 0.1875
ESSENTIALITY SCORE		+ 0.125 * ES	+ 0.125 * ES

<sup>1</sup>Yang F, et al. Proteindomain-level landscape of cancer-type-specific somatic mutations. *PLoS Comput Biol*. 2015 Mar 20;11(3):e1004147.

**Supplementary Table 3.** PanDrugs GScore weight assignation for lists of gene variants (VCF files). For each variant PanDrugs calculates a variant score (VScore). Highest VScore for each gene is selected as the GScore. (ONC: Oncogene; TSG: Tumor suppressor gene.)

Tumor type	TCGA Code	Mutations (Synapse IDs)	CNV (Synapse IDs)
Bladder Urothelial Carcinoma	BLCA	syn1729383	syn1687592
Breast invasive carcinoma	BRCA	syn1729383	syn395566
Cervical squamous cell carcinoma and endocervical adenocarcinoma	CESC	syn1729383	syn1687594
Colon adenocarcinoma	COAD	syn1729383	syn1687596
Rectum adenocarcinoma	READ	syn1729383	syn1687628
Glioblastoma multiforme	GBM	syn1729383	syn1687604
Head and Neck squamous cell carcinoma	HNSC	syn1729383	syn1687600
Kidney renal clear cell carcinoma	KIRC	syn1729383	syn1687602
Kidney renal papillary cell carcinoma	KIRP	syn1729383	syn1687614
Acute Myeloid Leukemia	LAML	syn1729383	syn1714787
Brain Lower Grade Glioma	LGG	syn1729383	syn1687616
Lung adenocarcinoma	LUAD	syn1729383	syn1687610
Lung squamous cell carcinoma	LUSC	syn1729383	syn1687612
Ovarian serous cystadenocarcinoma	OV	syn1729383	syn1687638
Pancreatic adenocarcinoma	PAAD	syn1729383	syn1687626
Prostate adenocarcinoma	PRAD	syn1729383	syn1687640
Skin Cutaneous Melanoma	SKCM	syn1729383	syn1687618
Stomach adenocarcinoma	STAD	syn1729383	syn1687622
Thyroid carcinoma	THCA	syn1729383	syn1687634
Uterine Corpus Endometrioid Carcinoma	UCEC	syn1729383	syn1687636

**Supplementary Table 4.** File sources for TCGA data employed by PanDrugs analysis.

<b>Resource</b>	<b>Version</b>
ensembl	version 90
COSMIC	Release 84
Pfam	31.0
UniProt	2018_02
InterPro	66.0
Clinvar	2018_02
APPRIS	gencode19/ensembl74

**Supplementary Table 5.** Database versions employed in the TCGA analysis.

Tumor type	Initial number of patients	Alteration type	Number of patients (SNV&Indels/CNV/Both)
BLCA	153	MUT/CNV/BOTH	99/135/96
BRCA	934	MUT/CNV/BOTH	771/866/752
CESC	134	MUT/CNV/BOTH	39/102/36
COAD	423	MUT/CNV/BOTH	155/413/153
READ	169	MUT/CNV/BOTH	69/162/68
GBM	599	MUT/CNV/BOTH	291/563/281
HNSC	343	MUT/CNV/BOTH	306/306/302
KIRC	502	MUT/CNV/BOTH	417/493/415
KIRP	117	MUT/CNV/BOTH	100/103/100
LAML	202	MUT/CNV/BOTH	196/194/190
LGG	222	MUT/CNV/BOTH	170/180/169
LUAD	543	MUT/CNV/BOTH	230/356/172
LUSC	389	MUT/CNV/BOTH	178/343/178
OV	599	MUT/CNV/BOTH	316/559/311
PAAD	57	MUT/CNV/BOTH	34/48/34
PRAD	180	MUT/CNV/BOTH	83/171/82
SKCM	264	MUT/CNV/BOTH	253/236/225
STAD	292	MUT/CNV/BOTH	151/237/115
THCA	435	MUT/CNV/BOTH	323/401/318
UCEC	512	MUT/CNV/BOTH	248/492/242
<b>Total</b>	<b>7069</b>		<b>4429/6360/4239</b>

**Supplementary Table 6.** Patients and genomic alterations considered in the TCGA data analysis sorted by tumor type.

**Supplementary Table 7.** Most frequently altered genes suggested for treatment by PanDrugs. The table shows the top-5 genes for each TCGA tumor type considering snv (point mutations and indels) and for CNVs separately.

Tumor	Gene	# cases	Event	Driver gene
BLCA	TP53	90	cnv	yes
BLCA	CDKN2A	81	cnv	yes
BLCA	CDKN2B	81	cnv	no
BLCA	LPL	80	cnv	no
BLCA	PTK2B	80	cnv	no
BLCA	TP53	50	snv	yes
BLCA	SYNE1	22	snv	yes
BLCA	PIK3CA	18	snv	yes
BLCA	RB1	14	snv	yes
BLCA	CDKN1A	13	snv	yes
BRCA	TP53	593	cnv	yes
BRCA	MC1R	571	cnv	no
BRCA	CYBA	569	cnv	no
BRCA	PLCG2	563	cnv	no
BRCA	PARD6A	550	cnv	no
BRCA	PIK3CA	261	snv	yes
BRCA	TP53	256	snv	yes
BRCA	CDH1	57	snv	yes
BRCA	MAP3K1	57	snv	yes
BRCA	MAP2K4	32	snv	yes
CESC	ETS1	60	cnv	no
CESC	TP53AIP1	60	cnv	no
CESC	ARHGEF12	59	cnv	yes
CESC	CHEK1	59	cnv	no
CESC	THY1	59	cnv	no
CESC	PIK3CA	9	snv	yes
CESC	NFE2L2	6	snv	yes
CESC	MYH9	5	snv	yes
CESC	CREBBP	4	snv	yes
CESC	SYNE1	4	snv	yes
COAD	SMAD4	270	cnv	yes
COAD	DCC	269	cnv	yes
COAD	MALT1	264	cnv	yes
COAD	TNFRSF11A	260	cnv	no
COAD	NFATC1	259	cnv	no
COAD	TP53	76	snv	yes
COAD	KRAS	59	snv	yes
COAD	SYNE1	38	snv	yes
COAD	PIK3CA	32	snv	yes
COAD	LRP2	29	snv	no
GBM	PTEN	505	cnv	yes
GBM	DOCK1	501	cnv	no
GBM	PLCE1	500	cnv	no
GBM	BLNK	499	cnv	no
GBM	CASP7	497	cnv	no
GBM	PTEN	90	snv	yes
GBM	TP53	84	snv	yes
GBM	EGFR	77	snv	yes

GBM	NF1	32	snv	yes
GBM	PIK3CA	32	snv	yes
HNSC	PRKCD	234	cnv	yes
HNSC	RHOA	234	cnv	yes
HNSC	TLR9	234	cnv	no
HNSC	WNT5A	234	cnv	no
HNSC	APPL1	233	cnv	no
HNSC	TP53	215	snv	yes
HNSC	CDKN2A	66	snv	yes
HNSC	PIK3CA	64	snv	yes
HNSC	SYNE1	56	snv	yes
HNSC	FAM135B	30	snv	yes
KIRC	VHL	445	cnv	yes
KIRC	CDC25A	437	cnv	no
KIRC	CTNNB1	435	cnv	yes
KIRC	CXCR6	435	cnv	no
KIRC	MYD88	435	cnv	yes
KIRC	VHL	218	snv	yes
KIRC	MTOR	25	snv	yes
KIRC	PTEN	18	snv	yes
KIRC	SYNE1	16	snv	yes
KIRC	FBN2	12	snv	yes
KIRP	MAPK12	30	cnv	no
KIRP	MAPK8IP2	29	cnv	no
KIRP	ADORA2A	28	cnv	yes
KIRP	BCR	28	cnv	yes
KIRP	CSNK1E	28	cnv	no
KIRP	MET	8	snv	yes
KIRP	SYNE1	6	snv	yes
KIRP	BRAF	4	snv	yes
KIRP	KAT6A	4	snv	yes
KIRP	LRP2	4	snv	no
LAML	FLT3	54	cnv	yes
LAML	CDK5	24	cnv	no
LAML	CUL1	23	cnv	yes
LAML	IRF5	23	cnv	no
LAML	LEP	23	cnv	no
LAML	FLT3	53	snv	yes
LAML	IDH2	20	snv	yes
LAML	IDH1	19	snv	yes
LAML	RUNX1	18	snv	yes
LAML	TP53	16	snv	yes
LGG	IDH1	131	cnv	yes
LGG	GP6	101	cnv	no
LGG	KIR2DL1	100	cnv	no
LGG	KIR3DL1	100	cnv	no
LGG	KIR3DL2	100	cnv	no
LGG	IDH1	131	snv	yes
LGG	TP53	88	snv	yes
LGG	PIK3CA	15	snv	yes
LGG	NF1	11	snv	yes
LGG	EGFR	8	snv	yes
LUAD	TP53	265	cnv	yes

LUAD	CDKN2A	217	cnv	yes
LUAD	CDKN2B	206	cnv	no
LUAD	NLRP1	206	cnv	no
LUAD	SERPINF1	206	cnv	no
LUAD	TP53	122	snv	yes
LUAD	KRAS	60	snv	yes
LUAD	FBN2	40	snv	yes
LUAD	KEAP1	40	snv	yes
LUAD	SYNE1	39	snv	yes
LUSC	RASSF1	297	cnv	no
LUSC	TLR9	297	cnv	no
LUSC	WNT5A	297	cnv	no
LUSC	APPL1	296	cnv	no
LUSC	PRKCD	295	cnv	yes
LUSC	TP53	145	snv	yes
LUSC	SYNE1	52	snv	yes
LUSC	FAM135B	33	snv	yes
LUSC	LRP2	32	snv	no
LUSC	SI	30	snv	no
OV	APC2	495	cnv	no
OV	S1PR4	492	cnv	no
OV	GNA11	491	cnv	yes
OV	MKNK2	491	cnv	no
OV	STK11	491	cnv	yes
OV	TP53	301	snv	yes
OV	CSMD3	18	snv	yes
OV	LRP2	15	snv	no
OV	NF1	12	snv	yes
OV	SYNE1	9	snv	yes
PAAD	SMAD4	31	cnv	yes
PAAD	TP53	31	cnv	yes
PAAD	CDKN2A	30	cnv	yes
PAAD	DCC	29	cnv	yes
PAAD	PIAS2	28	cnv	no
PAAD	KRAS	24	snv	yes
PAAD	TP53	22	snv	yes
PAAD	SMAD4	8	snv	yes
PAAD	CDKN2A	6	snv	yes
PAAD	MYH9	6	snv	yes
PRAD	GNRH1	97	cnv	no
PRAD	LPL	97	cnv	no
PRAD	PTK2B	90	cnv	no
PRAD	GATA4	87	cnv	no
PRAD	HTR2A	74	cnv	no
PRAD	SYNE1	5	snv	yes
PRAD	TP53	5	snv	yes
PRAD	HSPG2	4	snv	no
PRAD	BCL6	3	snv	yes
PRAD	CTNNB1	3	snv	yes
READ	DCC	143	cnv	yes
READ	SMAD4	143	cnv	yes
READ	MALT1	142	cnv	yes
READ	TNFRSF11A	142	cnv	no

READ	PMAIP1	141	cnv	no
READ	TP53	45	snv	yes
READ	KRAS	38	snv	yes
READ	PIK3CA	13	snv	yes
READ	SYNE1	11	snv	yes
READ	SMAD4	8	snv	yes
SKCM	CDKN2A	187	cnv	yes
SKCM	CDKN2B	182	cnv	no
SKCM	CER1	171	cnv	no
SKCM	IFNA1	171	cnv	no
SKCM	IFNB1	171	cnv	no
SKCM	BRAF	132	snv	yes
SKCM	NRAS	70	snv	yes
SKCM	SYNE1	61	snv	yes
SKCM	PREX2	59	snv	yes
SKCM	DCC	56	snv	yes
STAD	TP53	120	cnv	yes
STAD	DCC	107	cnv	yes
STAD	SMAD4	101	cnv	yes
STAD	TNFRSF11A	101	cnv	no
STAD	MALT1	100	cnv	yes
STAD	TP53	69	snv	yes
STAD	SYNE1	48	snv	yes
STAD	PREX2	28	snv	yes
STAD	PIK3CA	25	snv	yes
STAD	LRP2	24	snv	no
THCA	BRAF	187	cnv	yes
THCA	MAPK8IP2	71	cnv	no
THCA	CSF2RB	70	cnv	no
THCA	MAPK12	70	cnv	no
THCA	PPARA	70	cnv	no
THCA	BRAF	183	snv	yes
THCA	NRAS	26	snv	yes
THCA	HRAS	12	snv	yes
THCA	MT-ND5	9	snv	no
THCA	LRP1	6	snv	no
UCEC	PTEN	230	cnv	yes
UCEC	PLCG2	163	cnv	no
UCEC	PHLPP2	160	cnv	no
UCEC	PIK3CA	160	cnv	yes
UCEC	CDH1	157	cnv	yes
UCEC	PTEN	161	snv	yes
UCEC	PIK3CA	132	snv	yes
UCEC	CTNNB1	74	snv	yes
UCEC	TP53	69	snv	yes
UCEC	KRAS	53	snv	yes

**Supplementary Table 8.** Summary of 46 deleterious variants detected in SCLC patient enrolled for personalized medicine protocol.

chr	Position	Mutation	Consequence	Gene Symbol	Amino Acid Change	dbSNP	Lung Squamous Cell Carcinoma (TCGA) % Cases Altered	COSMIC ID	COSMIC Gene frequency
1	215960153	A/C	missense variant	USH2A	C3416G		36.50%	130095	649 / 606592
1	120612003	GG/-	frameshift variant, feature truncation	NOTCH2	6		9%		
3	119634983	G/C	missense variant	GSK3B	I172M		7.30%		18 / 606592
3	171417616	C/A	missense variant	PLD1	W382C		38.20%		73 / 606592
3	48723095	G/A	missense variant	NCKIPSD	P49L		1.70%		12 / 606592
3	53220704	C/T	missense variant	PRKCD	R449C		1.70%		18 / 606592
3	12626663	C/A	missense variant	RAF1	M562I		0.60%		32 / 606592
3	127800173	C/T	missense variant	RUVBL1	E431K		7.30%	172449	28 / 606592
3	46414969	-/GCTCTCAT	frameshift variant, feature elongation	CCR5	192		1.70%		
4	113356408	A/G	missense variant	ALPK1	R969G		1.70%		61 / 606592
4	106156522	ATGCTTTCTGAAAGGCCTCAGAATA/-	frameshift variant, feature truncation	TET2	475-483		2.80%		
5	156381503	C/A	missense variant	TIMD4	G108V		4.50%	402725	49 / 606592
5	140052407	G/A	missense variant	DND1	P76L	rs72800920	0.60%	1130919	6 / 606592
5	37044488	A/C	missense variant	NIPBL	I2050L		13.50%		134 / 606592
5	13864613	C/A	stop gained	DNAH5	E1497*		26.40%	232179	
6	107008770	G/T	missense variant	AIM1	R1575L		5.60%		67 / 606592
6	161508864	C/G	missense variant	MAP3K4	L901V		3.90%		81 / 606592
6	166827293	G/A	missense variant	RPS6KA2	H600Y		2.80%		43 / 606592
7	113558424	G/T	missense variant	PPP1R3A	R210S	rs141223649	8.40%		134 / 606592
8	88885088	C/T	missense variant	DCAF4L2	R371H		5.60%	1102264	97 / 606592
8	101719121	G/A	missense variant	PABPC1	R436C	rs79986761	1.10%	748078	43 / 606592
8	56922484	G/T	missense variant	LYN	V431L		3.90%		23 / 606592
8	87076765	A/T	missense variant	PSKH2	M94K		3.40%		46 / 606592
8	113516090	G/C	stop gained	CSMD3	S1671*		44.40%	603817	
9	90114003	T/A	missense variant	DAPK1	F4Y		1.70%		79 / 606592
9	8317878	G/A	missense variant	PTPRD	T1890M	rs151311972	12.40%		184 / 606592
9	120476517	A/T	missense variant	TLR4	Q640L		5.60%		117 / 606592

10	134059432	C/A	missense variant	STK32C	R36L		1.10%		20 / 606592
10	103221746	-/AG	frameshift variant, feature elongation	BTRC	19		1.70%		
11	534285	C/A	missense variant	HRAS	G13V	rs104894226	3.40%	489	21 / 606592
11	123777861	T/A	stop gained	OR8D4	C241*	rs61748875	2.80%	147344	
12	52369253	A/G	missense variant	ACVR1B	D47G		1.70%		41 / 606592
12	78513222	T/A	missense variant	NAV3	S154R		20.20%		261 / 606592
12	18439806	T/A	missense variant	PIK3C2G	V235E		5.10%		63 / 606592
12	56488226	G/T	missense variant	ERBB3	G523V		2.20%		2 / 606592
12	53605639	T/C	missense variant	RARG	R324G		2.20%		20 / 606592
14	33291955	T/A	missense variant	AKAP6	S1646T		9%		140 / 606592
14	21785937	C/T	stop gained	RPGRIP1	Q54*		1.10%	552574	
16	74502926	C/A	missense variant	GLG1	R774L		2.80%		61 / 606592
17	7577124	C/A	missense variant	TP53	V272L	rs121912657	90.40%	133678	1365 / 606592
17	78172215	G/T	missense variant	CARD14	R322L		1.70%		25 / 606592
17	29546102	C/G	stop gained	NF1	S202*		11.80%	96520	
17	29527440	A/T	stop gained, splice region variant	NF1	K297*		11.80%	41809	
19	10602314	C/T	missense variant	KEAP1	D422N		14%	710198	100 / 606592
19	11129683	T/A	missense variant	SMARCA4	V830E		6.20%		136 / 606592
22	26219574	T/A	missense variant	MYO18B	L875H		11.20%		165 / 606592