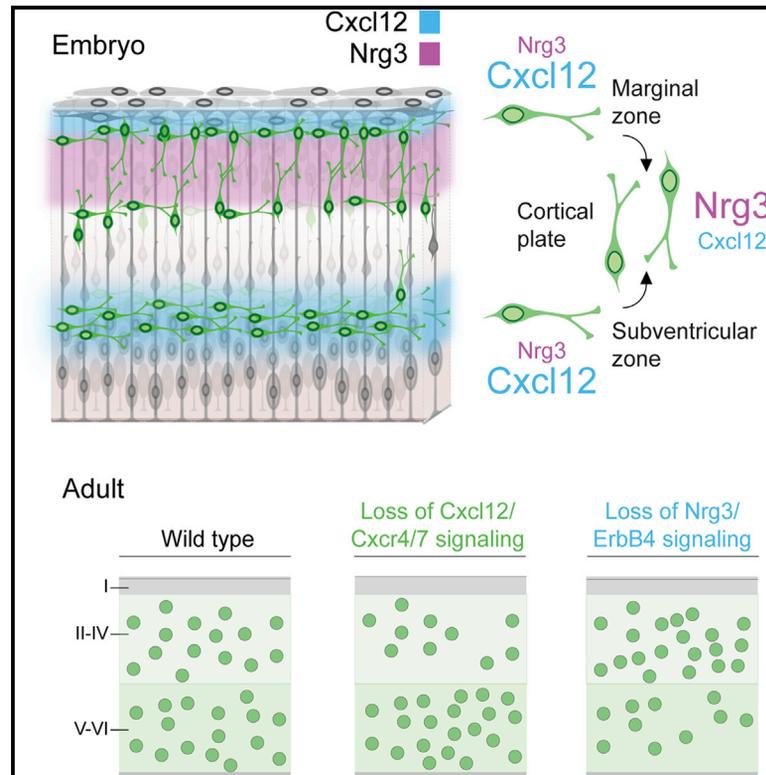


Neuregulin 3 Mediates Cortical Plate Invasion and Laminar Allocation of GABAergic Interneurons

Graphical Abstract



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In Brief

The integration of interneurons in the developing cerebral cortex depends on signals generated by pyramidal cells. In this study, Bartolini et al. identify Neuregulin 3 as a developmentally regulated factor that controls the migration of interneurons into the cortical plate and influences their final laminar distribution.

Highlights

- Nrg3 acts as a short-range chemoattractive molecule for cortical interneurons
- Nrg3 functions through ErbB4 to attract interneurons into the cortical plate
- Interneurons prefer Cxcl12 over Nrg3 during tangential migration
- Disruption of Nrg3 signaling causes abnormal interneuron lamination in the cortex



Neuregulin 3 Mediates Cortical Plate Invasion and Laminar Allocation of GABAergic Interneurons

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<http://dx.doi.org/10.1016/j.celrep.2016.12.089>

SUMMARY

Neural circuits in the cerebral cortex consist of excitatory pyramidal cells and inhibitory interneurons. These two main classes of cortical neurons follow largely different genetic programs, yet they assemble into highly specialized circuits during development following a very precise choreography. Previous studies have shown that signals produced by pyramidal cells influence the migration of cortical interneurons, but the molecular nature of these factors has remained elusive. Here, we identified Neuregulin 3 (Nrg3) as a chemoattractive factor expressed by developing pyramidal cells that guides the allocation of cortical interneurons in the developing cortical plate. Gain- and loss-of-function approaches reveal that Nrg3 modulates the migration of interneurons into the cortical plate in a process that is dependent on the tyrosine kinase receptor ErbB4. Perturbation of Nrg3 signaling in conditional mutants leads to abnormal lamination of cortical interneurons. Nrg3 is therefore a critical mediator in the assembly of cortical inhibitory circuits.

INTRODUCTION

The balance of excitation and inhibition is crucial for the normal functioning of the cerebral cortex (Vogels and Abbott, 2009; Xue et al., 2014; Yizhar et al., 2011), yet the mechanisms controlling the assembly of excitatory pyramidal cells and inhibitory gamma-aminobutyric acid-containing (GABAergic) interneurons into common circuits are poorly understood. Although most neural circuits consist of different classes of neurons, the cerebral cortex is relatively unique in that excitatory and inhibitory neurons are born in distinct progenitor regions of the embryonic telencephalon (Marín and Rubenstein, 2001; Wonders and An-

derson, 2006), and therefore their assembly requires the precise coordination of two largely different migration programs (Marín and Rubenstein, 2003).

Cortical GABAergic interneurons are highly heterogeneous and comprise distinct functional classes with unique morphological, electrophysiological, and molecular features (Tremblay et al., 2016). Recent transcriptomic analyses in the mouse suggest that the neocortex contains over 20 molecularly distinct classes of interneurons (Tasic et al., 2016; Zeisel et al., 2015), a number comparable to those identified in the hippocampus through morphological and electrophysiological studies (Klausberger and Somogyi, 2008). Nevertheless, neocortical interneurons can be classified into several cardinal groups based on their developmental origin and the expression of key molecular markers (Tremblay et al., 2016). The large majority of cortical interneurons derives from the embryonic medial ganglionic eminence (MGE) and comprises two major classes: parvalbumin-expressing (PV⁺) fast-spiking interneurons such as chandelier cells and basket cells, and somatostatin-expressing (SST⁺) interneurons comprising mostly Martinotti cells.

Our understanding of the mechanisms controlling the development of cortical interneurons has increased exponentially since Anderson et al. (1997) discovered the origin of cortical interneurons in the embryonic subpallium. However, although the mechanisms controlling the tangential migration of interneurons from the ganglionic eminences toward the cortex have been widely explored (Baudoin et al., 2012; Flames et al., 2004; Hernández-Miranda et al., 2011; Marín, 2013; Marín et al., 2001; Nóbrega-Pereira et al., 2008; Polleux et al., 2002; Pozas and Ibáñez, 2005; Rakić et al., 2015; van den Berghe et al., 2013), the cellular and molecular events governing the final allocation of interneurons in their corresponding cortical layers remain largely unknown. Interneurons follow two main routes of migration as they disperse tangentially throughout the cortex, the marginal zone (MZ) and the subventricular zone (SVZ) (Lavdas et al., 1999; Marín and Rubenstein, 2001; Miyoshi and Fishell, 2011; Tanaka et al., 2009). Interneurons initially avoid migrating into the cortical plate (CP), where pyramidal cells

differentiate and form the different cortical layers (López-Bendito et al., 2008). Chemokines are responsible for maintaining migrating interneurons in their migratory streams (Tiveron et al., 2006), and deficits in this signaling system lead to laminar and regional defects in the distribution of cortical interneurons (Li et al., 2008; López-Bendito et al., 2008; Sánchez-Alcañiz et al., 2011; Wang et al., 2011). These results revealed that the timing of CP invasion is crucial for the final allocation of interneurons in cortical layers, but the molecular factors controlling this process have not yet been identified.

Neuregulins comprise a large family of widely expressed epidermal growth factor (EGF)-like proteins that regulates multiple aspects of neurodevelopment, including neuronal migration, axon guidance, myelination, and synapse formation (Mei and Nave, 2014; Mei and Xiong, 2008). Neuregulins are EGF domain-containing proteins encoded by six individual genes that activate receptor tyrosine kinases of the ERBB family, and some of the members of this signaling network have been shown to play crucial roles in the assembly of GABAergic circuits (Rico and Marín, 2011). In particular, ErbB4 is highly expressed by interneurons migrating from the MGE (Rakić et al., 2015; Villar-Cerviño et al., 2015; Yau et al., 2003), most notably among PV⁺ interneurons (Fazzari et al., 2010; Vullhorst et al., 2009). ErbB4 is required for the tangential migration of GABAergic interneurons from the subpallium to the developing cortex, and this effect is mediated by different isoforms of Nrg1 (Flames et al., 2004; Rakić et al., 2015). Here, we have identified Nrg3 as a chemoattractive factor regulating the final allocation of GABAergic interneurons in the cortex. Our results demonstrate that ErbB4-mediated Nrg3 signaling controls CP entry by GABAergic interneurons and influences their final laminar allocation. Together with previous results, our observations unravel a hierarchical system of chemoattractive signals that regulate the distribution of interneurons in the cerebral cortex.

RESULTS

A Candidate Gene Approach to Identify Factors Regulating Cortical Plate Invasion by Interneurons

Different studies have shown that the chemokine Cxcl12 is responsible for maintaining the migration of interneurons within the MZ and the SVZ as they disperse tangentially throughout the cortex (Li et al., 2008; López-Bendito et al., 2008; Sánchez-Alcañiz et al., 2011; Stumm et al., 2007; Tiveron et al., 2006; Wang et al., 2011). Genetic deletion of chemokine receptors (Abe et al., 2014; Sánchez-Alcañiz et al., 2011; Tiveron et al., 2006; Vogt et al., 2014; Wang et al., 2011) or acute inhibition of chemokine signaling (Lysko et al., 2014) (Figure S1; Movies S1 and S2) lead to the rapid accumulation of interneurons in the CP, which suggests the existence of chemoattractive signals in this region regulating the final allocation of interneurons.

We took a candidate gene approach to identify molecules regulating the migration of interneurons into the CP. We hypothesized that factors relevant for this process might be upregulated in pyramidal cells at the time interneurons invade the CP. To analyze gene expression in equivalent cohorts of pyramidal cells, we performed in utero electroporation experiments with a plasmid encoding GFP targeting the dorsal pallium of E14.5

mice (Figure 1A). We then used fluorescence-activated cell sorting (FACS) to isolate GFP⁺ pyramidal cells from these experiments at two different stages: embryonic day 17.5 (E17.5), when pyramidal cells are still migrating toward the CP, and postnatal day 4 (P4), when pyramidal cells have already reached their final position (Figures 1B–1D). To examine the differential gene expression at these two stages in the development of pyramidal cells, we customized a TaqMan array with ~100 genes known to be involved in neuronal migration, adhesion, and axon guidance during corticogenesis (Table S1), including members of the eph, ephrin, semaphorin, plexin, cadherin, protocadherin, and neuregulin families. We also included several genes known to encode proteins expressed in pyramidal cells, such as *Cux1* and *Tbr1*, as positive controls, and genes that encode proteins exclusively expressed in interneurons, such as *Cxcr7*, *Cxcr4*, *Sst*, *Lhx6*, and *Htr3a*, as negative controls.

We identified 42 genes that are differentially expressed between the two stages (Figure 1E; Table S2). We focused our attention on 19 genes that are significantly more expressed by pyramidal cells at P4 compared to E17.5, because these are more likely to be involved in the chemoattraction of interneurons into the CP. To examine the pattern of expression of these genes, we performed in situ hybridization at E17.5 and P4 for this later list of candidate genes (*Cdh6*, *Cdh20*, *Epha6*, *Epha10*, *Ephb2*, *Ephb3*, *Ephb6*, *Efna3*, *Efna5*, *Efnb3*, *Plxndc2*, *Plxna1*, *Plxnd1*, *Pcdh9*, *Pcdh15*, *Pcdh20*, *Nrg3*, and *Sema7a*). Analysis of the expression of candidate genes revealed different patterns. For example, some genes were preferentially expressed in superficial layers of the cortex, and their expression increased during early postnatal stages (Figures S2A–S2F). In other cases, candidate genes were expressed throughout all layers of the neocortex (Figures S2G and S2H).

Nrg3 Is Expressed in the Developing Cortical Plate

We noticed that one of the genes that is expressed throughout the CP and that is significantly more expressed by pyramidal cells at later stages is *Neuregulin 3* (*Nrg3*), a member of the neuregulin family. In vitro experiments have shown that Nrg3 binds preferentially to ErbB4 receptors (Zhang et al., 1997), which are highly enriched in migrating cortical interneurons and excluded from pyramidal cells (Fazzari et al., 2010; Flames et al., 2004; Vullhorst et al., 2009; Yau et al., 2003). Based on this evidence, we hypothesized that Nrg3 might regulate the intracortical migration of GABAergic interneurons and focused our subsequent work on this molecule.

We investigated the pattern of expression of *Nrg3* in the developing cortex from mid-embryonic until early postnatal stages using in situ hybridization (Figure 2). We observed that *Nrg3* is highly expressed in the developing CP since its inception, and that Nrg3 expression is maintained in pyramidal cells as they mature and start forming differentiated layers. Pyramidal cells therefore express *Nrg3* as soon as they reach the CP, and its expression is subsequently maintained throughout all layers of the neocortex, including the subplate. *Nrg3* is, however, largely absent from the MZ and the SVZ at all stages examined (Figure 2). These results support the hypothesis that Nrg3 might be involved in the regulation of interneuron migration into the CP.

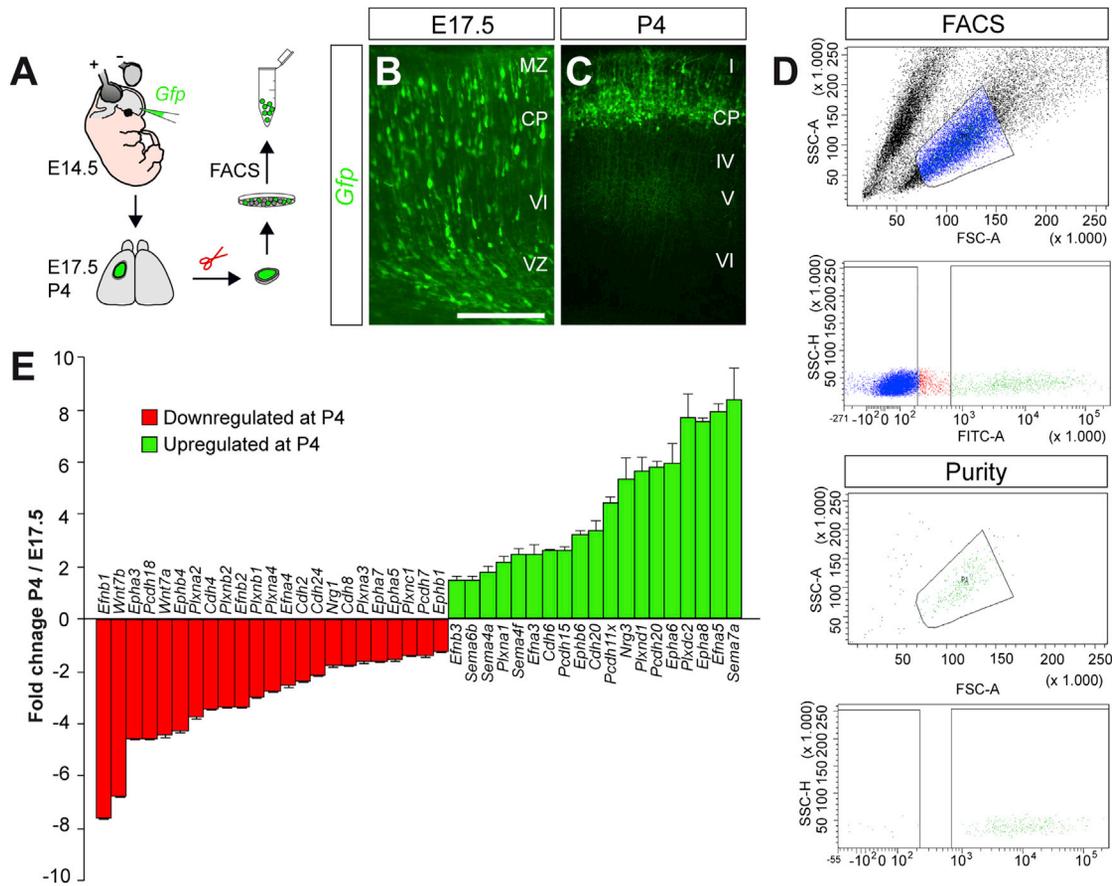


Figure 1. Identification of Factors Regulating the Invasion of the Cortical Plate by Interneurons

(A) Schematic of the experimental design. A plasmid encoding *Gfp* was electroporated in the neocortex of E14.5 mouse embryos. The electroporated region was then isolated at E17.5 or P4, and GFP⁺ cells were dissociated and recovered via FACS.

(B and C) Distribution of GFP⁺ pyramidal cells at E17.5 (B) and P4 (C), prior to isolation.

(D) FACS of GFP⁺ cells.

(E) Among the pool of candidate genes (Table S1), we found 42 that were differentially expressed between the two stages. The corresponding p values from *t*-tests are shown in Table S2. Histograms show average \pm SEM. Scale bar represents 250 μ m.

Nrg3 Is a Short-Range Chemoattractant for MGE-Derived Interneurons

We have previously shown that different isoforms of *Nrg1* act both as short- and long-range chemoattractive molecules for tangentially migrating interneurons (Flames et al., 2004). To examine whether *Nrg3* may exert a similar effect on cortical interneurons, we performed confrontation assays in three-dimensional matrices in which we cultured MGE explants obtained from E13.5 GFP-expressing embryos together with aggregates of COS cells transfected with control or *Nrg3* encoding plasmids (Figure 3A1). In parallel experiments, we carried out co-cultures in which COS cells were transfected with *Ig-Nrg1*, which encodes for a diffusible form of *Nrg1*. As described before (Flames et al., 2004), we observed that Ig-*Nrg1* exerts a prominent chemoattractive response in MGE-derived cells (Figures 3B, 3C, and 3H). In contrast, we found no significant differences in the response of MGE-derived cells to *Nrg3* compared to controls (Figures 3B, 3D, and 3H). Thus, *Nrg3* does not seem to function as a long-range chemoattractant for MGE-derived interneurons.

To investigate whether *Nrg3* may function as a short-range chemoattractant for migrating interneurons, we carried out a new set of co-culture experiments in which COS cell aggregates, transfected with control or *Nrg3*-encoding plasmids, were placed at a relatively short distance from MGE explants (Figure 3A2). In this new set of experiments, we used COS cells transfected with *CRD-Nrg1* as a positive control, because this membrane-bound form of *Nrg1* has been shown to induce short-range chemoattraction during the migration of cortical interneurons through the subpallium (Flames et al., 2004). We found that both *CRD-Nrg1* and *Nrg3* evoke a potent chemoattractive effect on migrating interneurons, which can be visualized by the great abundance of cells accumulating around the proximal side of COS cell aggregates (Figures 3E–3G and 3I). Thus, *Nrg3* induces a potent short-range chemoattractive effect on MGE-derived interneurons.

Nrg3 Chemoattraction Requires ErbB4 Function

Because *Nrg3* is thought to bind preferentially to ErbB4 receptors (Zhang et al., 1997), we next examined whether ErbB4

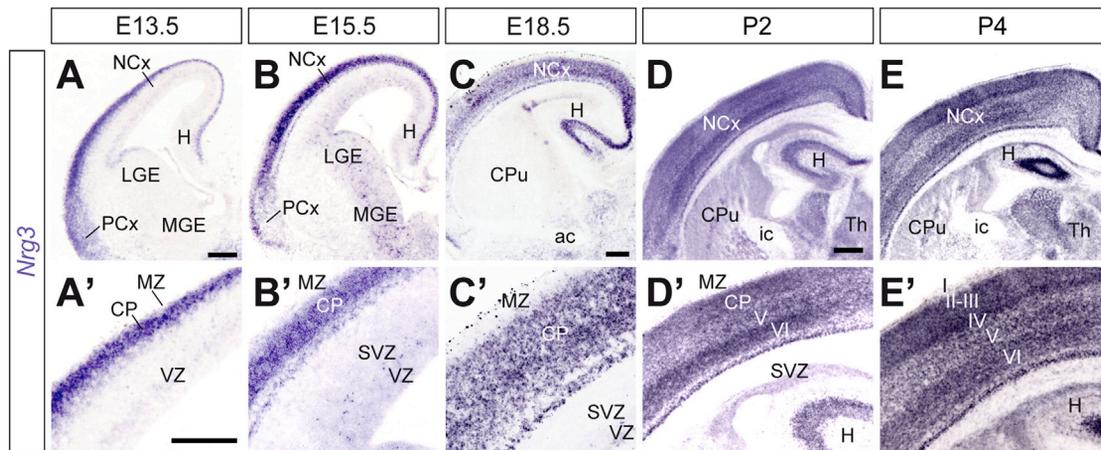


Figure 2. Expression of *Nrg3* mRNA in the Developing Mouse Cortex

(A–E') Coronal sections through the telencephalon of E13.5 (A and A'), E15.5 (B and B'), E18.5 (C and C'), P2 (D and D'), and P4 (E and E') embryos and neonates showing mRNA expression for *Nrg3*. Ac, anterior commissure; CP, cortical plate; H, Hippocampus; ic, internal capsule; LGE, lateral ganglionic eminence; MGE, medial ganglionic eminence; MZ, marginal zone; NCx, neocortex; PCx, piriform cortex; Str, striatum; SVZ, subventricular zone; Th, thalamus; VZ, ventricular zone; I–VI, cortical layers I–VI. Scale bars represent 250 μ m.

function mediates the chemoattractive responses elicited by *Nrg3* in MGE-derived interneurons. To this end, we carried out a new set of co-culture experiments with MGE explants obtained from control and *ErbB4* mutant embryos (Figure 3J). Because loss of *ErbB4* causes early lethality due to cardiac defects, *ErbB4* mutants carried a human transgene under a cardiac-specific myosin promoter (*HER4^{heart}*) to circumvent this problem (Tidcombe et al., 2003). In contrast to controls, we observed that *Nrg3* does not exert any effect on MGE-derived cells obtained from *ErbB4* mutant embryos (Figures 3K–3O). Thus, *ErbB4* is necessary for the migration of cortical interneurons in response to *Nrg3*. Altogether, these experiments indicate that the chemoattractive effect elicited by *Nrg3* on migrating interneurons is mediated by *ErbB4* function.

Tangentially Migrating MGE-Derived Interneurons Prefer *Cxcl12* to *Nrg3*

It has been previously shown that the chemokine *Cxcl12* strongly promotes the tangential migration of MGE-derived cells throughout the embryonic cortex. *Cxcl12* is expressed by the meninges and in the SVZ of the pallium during embryonic development (Borrell and Marín, 2006; Stumm et al., 2003; Tiveron et al., 2006). Because *Nrg3* is expressed in the CP from early stages of development (Figures 2A and 2A'), tangentially migrating interneurons encounter both cues as they reach the embryonic cortex. Both *Cxcl12* and *Nrg3* seem to function as chemoattractive factors for migrating MGE-derived interneurons, so we next explored whether interneurons display a preference for any of these molecules. To this end, we cultured MGE explants obtained from E13.5 embryos from GFP-expressing mice together with aggregates of COS cells placed at a short distance and transfected with either a mock plasmid, *Nrg3* or *Nrg3* and *Cxcl12* together (Figure 4A1). As expected, we observed that *Cxcl12* enhances the migration of MGE-derived interneurons (Figures 4B, 4D, and 4E). In addition, we found that *Cxcl12* does not block the chemoattractive effect elicited by

Nrg3 (Figures 4B–4E). These results suggested that tangentially migrating MGE-derived interneurons are equipped to respond to both *Cxcl12* and *Nrg3* simultaneously.

We next wondered whether MGE-derived interneurons display any preference for *Cxcl12* or *Nrg3*. To answer this question, we performed stripe choice assays using recombinant proteins, as described previously (Walter et al., 1987). In brief, E13.5 MGE explants obtained from GFP-expressing embryos were placed on top of stripes coated with a control peptide (GST), *Nrg3*-GST or recombinant *Cxcl12* in alternate combinations (Figure 4A2), and their lane preference was scored after 48 hr. As expected, MGE-derived cells showed no migratory preference when alternative stripes were coated with the same recombinant protein (GST/GST, *Nrg3*/*Nrg3* or *Cxcl12*/*Cxcl12*) in control experiments (Figure 4I; data not shown). In contrast, MGE-derived interneurons displayed a strong preference for *Nrg3*-coated stripes compare to control lanes (Figures 4G and 4I). Moreover, MGE-derived cells exhibited a strong preference toward *Cxcl12* when they were given the possibility to migrate on alternating stripes containing *Nrg3* and *Cxcl12* (Figures 4H and 4I). Taken together, these experiments suggest that tangentially migrating interneurons can respond simultaneously to *Cxcl12* and *Nrg3*, but they display stronger affinity for the chemokine. These observations are consistent with the *in vivo* behavior of MGE-derived interneurons, which initially disperse through the cortex via *Cxcl12*-rich territories (MZ and SVZ) without accumulating in the CP (López-Bendito et al., 2008).

Nrg3 Overexpression Promotes Interneuron Invasion of the CP *In Vivo*

We next wondered whether unbalancing the normal levels of *Nrg3* in the developing CP would interfere with the migration of cortical interneurons *in vivo*. To this end, we electroporated the ventricular zone of the pallium in E14.5 *Nkx2-1Cre;Ai9* embryos with either *Gfp*-expressing plasmids or a combination of *Gfp* and *Nrg3* (Figure 5A), and examined the distribution of

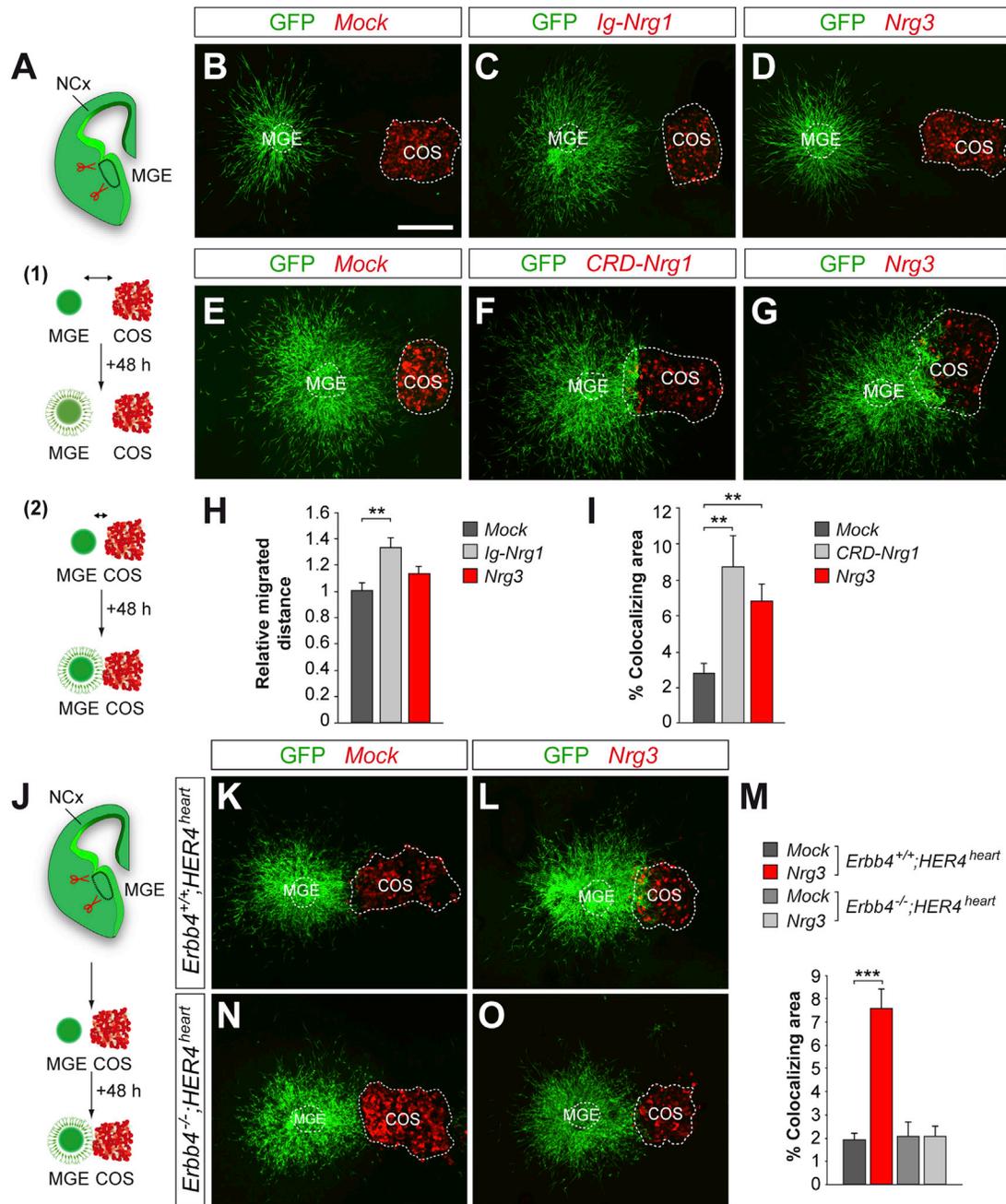


Figure 3. Nrg3 Functions as a Short-Range Chemoattractant for MGE-Derived Interneurons and Requires ErbB4 Receptors

(A) Schematic of the experimental design. MGE explants were confronted to transfected COS cells located at relative long (A₁) or short (A₂) distances. (B–G) Migration of MGE-derived interneurons in response to mock- (B and E), *Ig-Nrg1*- (C), *CRD-Nrg1*- (D), or *Nrg3*-transfected (D and G) COS cells located at relatively long (B–D) and short (E–G) range. Dotted lines indicate the limits of the explants and COS cell aggregates. (H) Quantification of long-distance confrontation assays. Control: MGE versus mock, n = 19; MGE versus *Ig-Nrg1*, n = 24; one-way ANOVA, **p < 0.01. MGE versus *Nrg3*, n = 20; one-way ANOVA, p > 0.05. (I) Quantification of short-distance confrontation assays. Control: MGE versus mock, n = 29; MGE versus *CRD-Nrg1*, n = 24, MGE versus *Nrg3*, n = 27; one-way ANOVA, **p < 0.01. (J) Schematic of the experimental design. MGE explants were confronted to control and *Nrg3*-transfected COS cells located at relative short distances. (K, L, N, and O) Migration of MGE-derived cells derived from *Erb4*^{+/+}; *HER4*^{heart} (K and L) and *Erb4*^{-/-}; *HER4*^{heart} (N and O) mice in response to mock- (K and N) or *Nrg3*-transfected (L and O) COS cell aggregates cultured in matrigel matrices for 48 hr. (M) Quantification of confrontation assays. *Erb4*^{+/+}; *HER4*^{heart} versus mock, n = 33; *Erb4*^{+/+}; *HER4*^{heart} versus *Nrg3*, n = 25; *Erb4*^{-/-}; *HER4*^{heart} versus mock, n = 15; *Erb4*^{-/-}; *HER4*^{heart} versus *Nrg3*, n = 14; one-way ANOVA, ***p < 0.001. Histograms show average ± SEM. NCx, neocortex; MGE, medial ganglionic eminence. Scale bar represents 200 μm.

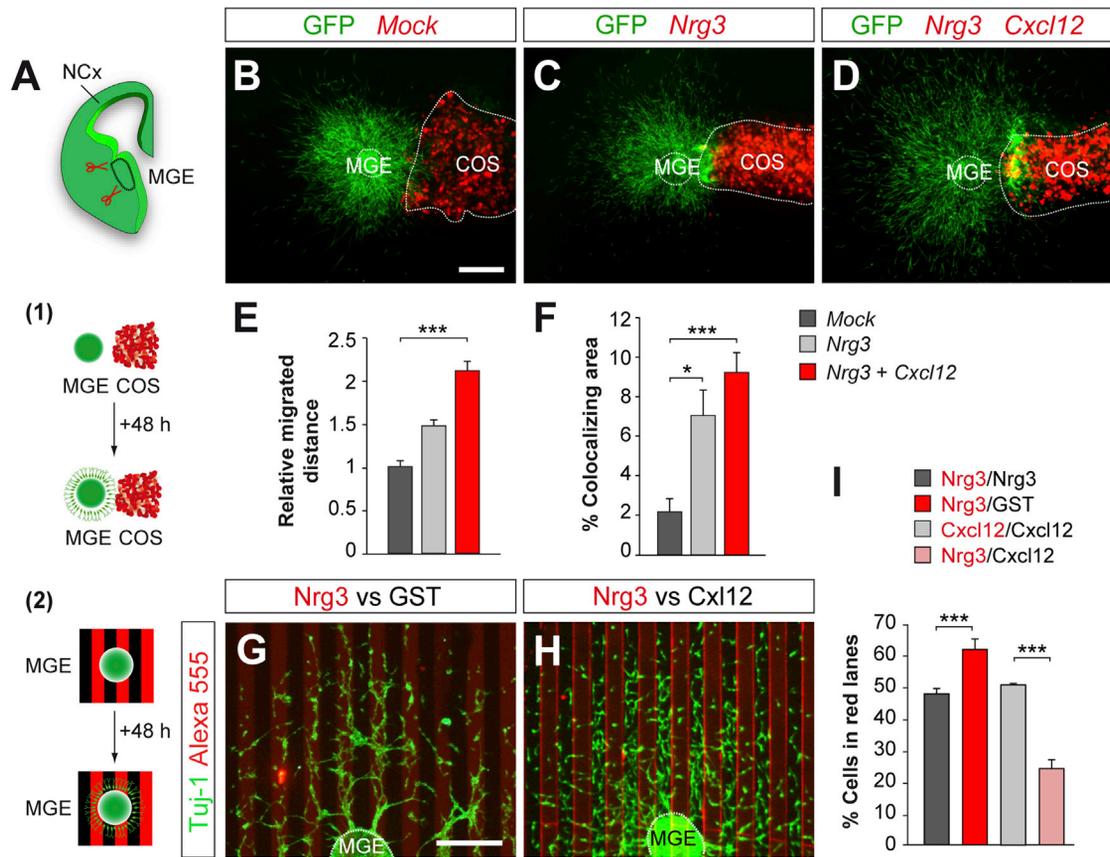


Figure 4. MGE-Derived Interneurons Display Preferential Responses to Cxcl12 and Nrg3

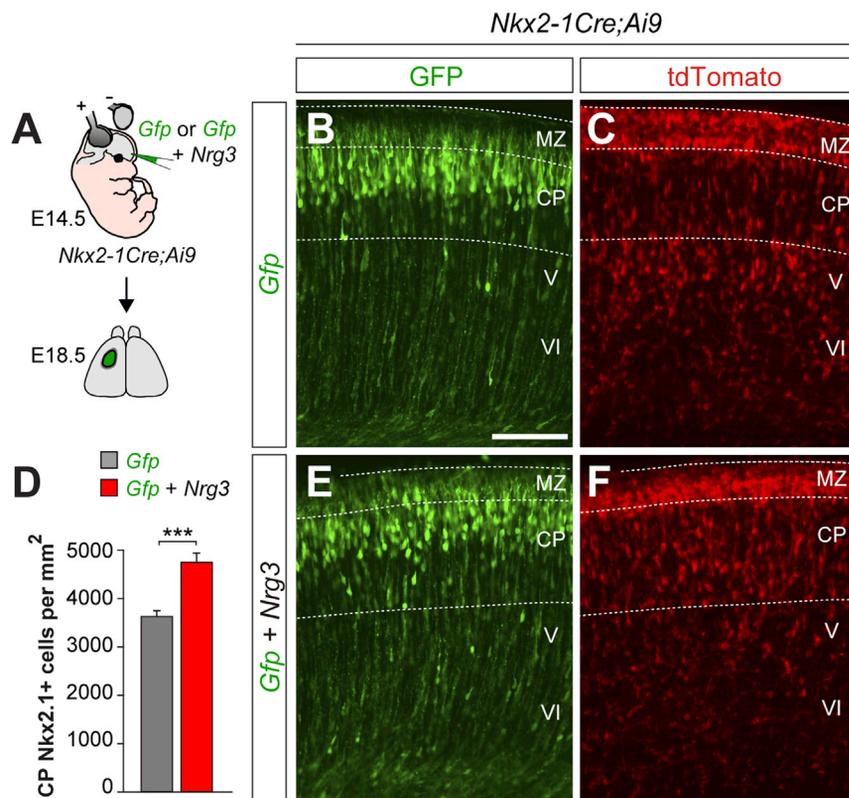
(A) Schematic of the experimental designs. MGE explants were confronted to transfected COS cells located at relative short distances (A₁) or placed on coated matrices in stripe choice assays (A₂). (B–D) Migration of MGE-derived interneurons in response to mock- (B), *Nrg3*- (C), or *Nrg3*- and *Cxcl12*-transfected (D) COS cells. Dotted lines indicate the limits of the explants and COS cell aggregates. (E) Quantification of MGE-derived interneurons migrating away from explants. MGE versus mock, n = 18; MGE versus *Nrg3*, n = 15; MGE versus *Nrg3* + *Cxcl12*, n = 27; one-way ANOVA, ***p < 0.001. (F) Quantification of short-distance confrontation assays. MGE versus mock, n = 18; MGE versus *Nrg3*, n = 15; MGE versus *Nrg3* + *Cxcl12*, n = 27; one-way ANOVA, ***p < 0.001, *p < 0.05. (G and H) Migration of MGE-derived cells in the stripe choice assay, with control and *Nrg3*-coated (G) or *Nrg3*- and *Cxcl12*-coated alternating stripes (H). (I) Quantification of stripe choice assays. *Nrg3*/*Nrg3* (n = 11) versus *Nrg3*/GST (n = 20); one-way ANOVA, ***p < 0.001. *Cxcl12*/*Cxcl12* (n = 13) versus *Nrg3*/*Cxcl12* (n = 18); one-way ANOVA, ***p < 0.001. Histograms show average ± SEM. NCx, neocortex; MGE, medial ganglionic eminence. Scale bar represents 200 μm.

MGE-derived interneurons (labeled with tdTomato) at E18.5. We observed that overexpression of *Nrg3* does not seem to disrupt the migration of pyramidal cells (Figures 5B and 5E). In contrast, overexpression of *Nrg3* accelerates the invasion of the CP by MGE-derived interneurons (Figures 5C, 5D, and 5F). Thus, these results suggested that *Nrg3* promotes the intracortical migration of MGE-derived interneurons in vivo.

In Vivo Loss of *Nrg3*/*ErbB4* Signaling Disrupts Interneuron Lamination

To examine the long-term consequences of disrupting *Nrg3* signaling in vivo, we generated mice carrying conditional *Nrg3* alleles (Figure S3). To delete *Nrg3* specifically from developing pyramidal cells, we used *Nex-Cre* mice, in which exon 2 of the *NeuroD6* locus has been replaced by Cre recombinase (Goebels et al., 2006). We found that early deletion of *Nrg3* in pyrami-

dal cells does not alter their laminar position in the postnatal cortex (Figure S4). Analysis of the distribution of *ErbB4*-expressing neurons in the somatosensory cortex of control and conditional *Nrg3* mutants at P30 revealed no differences in the total density of *ErbB4*⁺ cells (controls: 783.28 ± 48.18 cells/mm²; *Nrg3* mutants: 923.69 ± 56.15 cells/mm²; n = 4; t test, p > 0.05). However, we observed that the laminar location of *ErbB4*-expressing neurons was significantly different between control and conditional *Nrg3* mutants, with a deep to superficial layer shift in the distribution of *ErbB4*-expressing cells in mutants compared to controls (Figures 6A–6C). Because *ErbB4* is primarily expressed by PV⁺ interneurons in the cortex (Fazzari et al., 2010; Neddens and Buonanno, 2010), we also analyzed their distribution at P30. Similar to the analysis of *ErbB4*-expressing neurons, we found no differences in the total density of PV⁺ interneurons (controls: 281.88 ± 25.29 cells/mm²; *Nrg3*



mutants: 258.68 ± 36.1 cells/mm²; $n = 5$; t test, $p > 0.05$), but a subtle shift in their laminar distribution (Figures 6D–6F). To determine whether these differences in the laminar distribution of cortical interneurons were already present in the early postnatal cortex, we examined the distribution of *Lhx6*-expressing neurons in the somatosensory cortex of control and conditional *Nrg3* mutants at P4. We found a significant shift toward superficial layers in the distribution of *Lhx6*-expressing cells in conditional *Nrg3* mutants compared to controls (Figures 6G–6I). Altogether, these results revealed that conditional deletion of *Nrg3* from developing pyramidal cells disrupts the laminar distribution of cortical interneurons.

Because *Nrg3* signaling requires *ErbB4* function in cortical interneurons (Figures 3J–3O), we next wondered whether similar layering defects exist in conditional *ErbB4* mutants. To this end, we bred *Lhx6-Cre* mice with mice carrying loxP-flanked (*F*) *ErbB4* alleles and analyzed the distribution of PV⁺ interneurons in the somatosensory cortex. We have previously shown that the total number of interneurons in the cortex in *Lhx6-Cre;ErbB4^{F/F}* mice is similar to controls at P30 (Del Pino et al., 2013). However, detailed analysis of the laminar distribution of PV⁺ interneurons revealed a clear shift in conditional *ErbB4* mutants, with fewer cells in deep layers and more cells in superficial layers than in controls (Figures 7A–7F). To elucidate whether this defect reflects an overall shift in the allocation of cortical interneurons, we examined the laminar distribution of specific cohorts of PV⁺ cells by injecting BrdU in control and conditional *ErbB4* mutants at E12.5 or E15.5 (Figure 7A). We found that inter-

Figure 5. Nrg3 Overexpression in Pyramidal Cells Enhances Interneuron Invasion of the CP In Vivo

(A) Schematic of the experimental design. The pallial ventricular zone of *Nkx2-1Cre;Ai9* embryos was electroporated at E14.5 with *Gfp*-expressing or *Gfp* and *Nrg3*-expressing plasmids, and the distribution of MGE-derived interneurons was analyzed at E18.5.

(B, C, E, and F) Distribution of MGE-derived interneurons (labeled with *tdTomato*) in the somatosensory cortex of E18.5 *Nkx2-1Cre;Ai9* embryos following in utero electroporation of *Gfp* (B and C) or *Gfp* and *Nrg3* (E and F) at E14.5.

(D) Density of MGE-derived cells in the CP; $n = 6$; t test, $***p < 0.001$. Histograms show average \pm SEM. CP, cortical plate; MZ, marginal zone; V and VI, cortical layers V and VI, respectively. Scale bar represents 200 μm .

neurons born at E12.5 and E15.5 tend to occupy deep and superficial layers of the cortex, respectively, in both controls and *ErbB4* mutants (Figures 7B–7E, 7G, and 7H). However, the distribution of PV⁺ interneurons was shifted toward progressively more superficial layers for both cohorts of cells in conditional *ErbB4* mutants compared to controls (Figures 7G and 7H), a result that is consistent with

the overall shift in the distribution of PV⁺ interneurons toward superficial layers (Figure 7F). Altogether, these experiments indicate that *ErbB4* mediates the function of *Nrg3* in the intracortical migration of interneurons, and that this signaling system is required for the appropriate laminar allocation for these cells.

DISCUSSION

We have identified *Nrg3* as a factor regulating the assembly of inhibitory neurons in the developing cerebral cortex. We found that *Nrg3* is highly expressed by developing pyramidal cells during the entire period of interneuron allocation in the cerebral cortex. Gain- and loss-of-function experiments suggest that *Nrg3* functions specifically as a chemoattractive signal regulating CP invasion by interneurons, and that this process is mediated by the tyrosine kinase receptor *ErbB4*. *Nrg3* is therefore a critical mediator in the assembly of inhibitory circuits, controlling the orderly entry of interneurons in the CP prior to their subsequent sorting into specific cortical layers.

Nrg3 in Cortical Interneuron Development

Nrg3 was identified as a neural tissue-enriched molecule nearly 20 years ago (Zhang et al., 1997), yet very little is known about its role in brain development and function. An isoform of NRG3 that is specifically expressed in the human embryonic CNS has been shown to influence oligodendrocyte survival in vitro (Carteron et al., 2006). In addition, recent in vitro experiments revealed that MGE-derived interneurons are able to respond to *Nrg3*

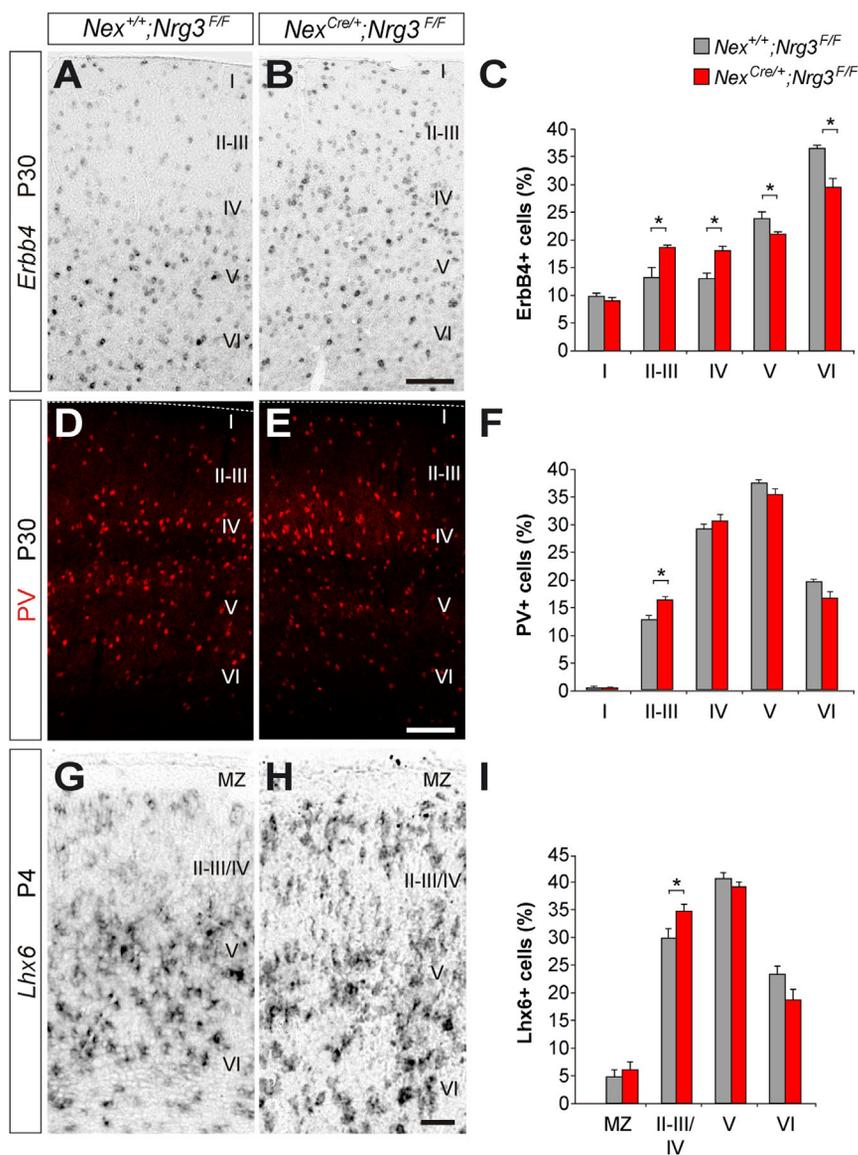


Figure 6. Abnormal Lamination of Cortical Interneurons in Conditional *Nrg3* Mutants

(A and B) Distribution of *ErbB4*-expressing interneurons in the somatosensory cortex of control (A) and conditional *Nrg3* (B) mice at P30. (C) Quantification of the distribution of *ErbB4*-expressing cells; n = 4; χ^2 test, *p < 0.05. (D and E) Distribution of PV⁺ interneurons in the somatosensory cortex of control (D) and conditional *Nrg3* (E) mice at P30. (F) Quantification of the distribution of PV⁺ interneurons; n = 5; χ^2 test, *p < 0.05. (G and H) Distribution of *Lhx6*-expressing cells in the somatosensory cortex of control (G) and conditional *Nrg3* (H) mice at P4. (I) Quantification of the laminar distribution of *Lhx6*-expressing cells; n = 5; *p < 0.05, χ^2 test. Histograms show average \pm SEM. Scale bars represent 200 μ m.

released in vivo through post-translational proteolysis, our experiments suggest that its diffusion is likely restricted to near the CP. In this context, it is worth noting that *Nrg3* contains a mucin-like Ser/Thr-rich region containing abundant sites for O-linked glycosylation (Zhang et al., 1997), which may function to limit its diffusion.

Constitutive *Nrg3* mutant mice have decreased impulsivity, increased activity in a novel open field, and deficient pre-pulse inhibition of the acoustic startle response and fear conditioning (Hayes et al., 2016; Loos et al., 2014). Interestingly, these defects are not rescued by overexpression of *Nrg3* in the adult medial prefrontal cortex—the region responsible for impulsive responding (Loos et al., 2014). In contrast, systemic overexpression of *Nrg3* during early postnatal development via subcutaneous injections

(Li et al., 2012; Rakić et al., 2015). This is consistent with the observation that MGE-derived interneurons express ErbB4 (Flames and Marín, 2005; Yau et al., 2003), and that ErbB4 is the preferred receptor for *Nrg3* (Zhang et al., 1997). Our experiments confirm and extend these observations by demonstrating a specific role for *Nrg3* in the entry of GABAergic interneurons in the developing CP in vivo.

Our results suggest that *Nrg3* functions as a short-range chemoattractive factor for cortical interneurons. Analysis of the amino acid sequence of human *Nrg3* reveals important homologies with *Nrg1*, but contrary to many *Nrg1* family members the extracellular domain of *Nrg3* lacks Ig-like domains (Zhang et al., 1997). Although the structure of *Nrg3* resembles more closely that of type I/II *Nrg1*, our experiments indicate that *Nrg3* functions largely as a short-range chemoattractant, most similar to membrane-bound forms of *Nrg1*. Even if *Nrg3* is

leads to alterations in anxiety and social behaviors in adulthood (Paterson and Law, 2014). It is conceivable that the behavioral deficits reported in these mice are due to defects in the distribution of cortical interneurons, reinforcing the notion that *Nrg3* function during development is required for normal brain function.

Neuregulins have been also implicated in other steps in the development of MGE-derived cortical interneurons (Rico and Marín, 2011). In particular, *Nrg1* has been shown to regulate the wiring of PV⁺ interneurons in the postnatal cortex (Fazzari et al., 2010; Ting et al., 2011). Because *Nrg3* expression is maintained in pyramidal cells into adulthood, it remains to be explored whether this neuregulin may contribute to the regulation of the connectivity of PV⁺ interneurons. *Nrg3* is also expressed in cortical interneurons in the postnatal brain (Loos et al., 2014), which suggests that its function in cortical

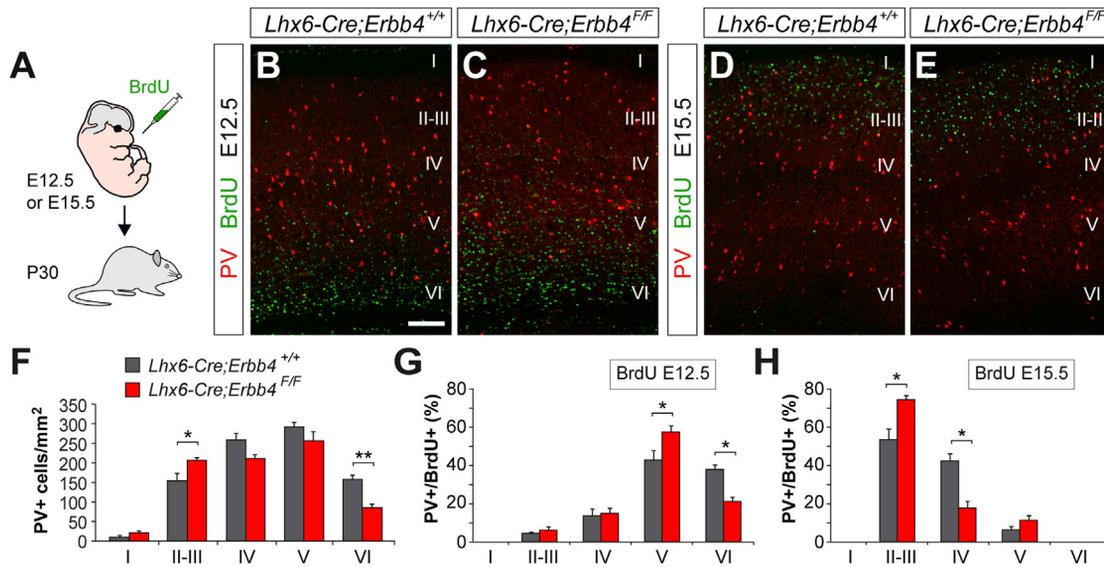


Figure 7. Abnormal Lamination of Cortical Interneurons in Conditional *Erbb4* Mutants

(A) Schematic of the experimental design.

(B–E) Lamina distribution of PV⁺ interneurons (red) and specific cohorts of PV⁺ interneurons labeled after BrdU injections at E12.5 (B and C) and E15.5 (D and E) in the somatosensory cortex of control (B and D) and conditional *Erbb4* mutant (C and E) mice at P30.

(F) Quantification of the distribution of PV⁺ interneurons; n = 5; χ^2 test, *p < 0.05.

(G and H) Quantification of the distribution of specific cohorts of PV⁺ interneurons born at E12.5 (G) and E15.5 (H); n = 5; χ^2 test, *p < 0.05. Histograms show average \pm SEM. Scale bar represents 200 μ m.

circuit assembly might be more complex than previously anticipated.

A Hierarchical Organization of Guidance Cues for Cortical Interneurons

Our experiments suggest that the transition from tangential to radial migration and the allocation of interneurons inside the CP depends on a delicate balance between two chemoattractive factors, *Ngr3* and *Cxcl12*, which seems to be hierarchically organized. Our experiments indicate that tangentially migrating interneurons show a prominent preference for *Cxcl12* during their tangential dispersion through the cortex, and it is only when they lose their responsiveness to this molecule that they steer toward the source of *Nrg3* in the CP. One caveat of the *in vitro* experiments is that the tested MGE interneurons are younger than they would normally be when they encounter both molecules in the cortex *in vivo*. Our *in vivo* analyses, however, strongly suggest that interneurons respond to both of these cues during the migration through the cortex.

Cxcl12 is strongly expressed by the meninges and by intermediate progenitor cells transiently present in the SVZ (Stumm et al., 2003; Tham et al., 2001; Tiveron et al., 2006) and is also expressed by cells in the SP (Stumm et al., 2007). *Cxcl12* is a potent long-range chemoattractant for MGE-derived interneurons *in vitro* (Li et al., 2008; López-Bendito et al., 2008), but its limited diffusion *in vivo* would explain the relative confinement of interneurons to the migratory streams found in the cortex. Consistent with this idea, mouse mutants with altered expression of *Cxcl12* in the meninges or in the SVZ have defects in the intracortical migration of interneurons that are specific to the affected migra-

tory route (Abe et al., 2015; Sessa et al., 2010; Tiveron et al., 2006; Zarbališ et al., 2012).

The molecular mechanisms regulating the tangential to radial switch in the migration of cortical interneurons and the subsequent CP invasion remains unknown. The exit of interneurons from the migratory streams is coordinated with the loss of responsiveness to *Cxcl12* (Li et al., 2008), and this is ultimately linked with the entry of interneurons into the CP. The analysis of *Cxcr4* and *Cxcr7* mutants revealed many interneurons inside CP at relatively early embryonic stages (Li et al., 2008; López-Bendito et al., 2008; Sánchez-Alcañiz et al., 2011; Tanaka et al., 2010; Tiveron et al., 2006; Wang et al., 2011), a finding that can now be interpreted as a premature response to *Nrg3*.

The timing of interneuron responses to the cues guiding their migration into the cortex seems tightly regulated, and it is important for the final distribution of interneurons in different regions and layers of the cortex. Premature entry in the cortical plate, as observed in *Cxcr4* and *Cxcr7* mutants, causes defects in the regional and laminar distribution of interneurons in the adult cortex (Li et al., 2008; López-Bendito et al., 2008; Sánchez-Alcañiz et al., 2011; Tanaka et al., 2010; Wang et al., 2011). Similarly, removal of *Nrg3*, a factor that facilitates CP entry, also disrupts laminar distribution in the adult cortex. Of note, the laminar defects observed in *Cxcr4/Cxcr7* and *Nrg3* mutants are complementary (accumulation of interneurons in deep or superficial layers for *Cxcr4/Cxcr7* and *Nrg3* mutants, respectively). These observations reinforce the notion that *Cxcl12* and *Nrg3* play sequential and largely complementary roles in the migration of cortical interneurons.

Pyramidal Cells and Microglia Regulate the Distribution of Interneurons

Studies over the last decade have revealed some important aspects on the regulation of layer acquisition by cortical interneurons. Most notably, several studies have suggested that pyramidal cells regulate the laminar distribution of cortical interneurons (Hevner et al., 2004; Lodato et al., 2011; Pla et al., 2006). In *reeler* mice, for example, the distribution of interneurons follow that of pyramidal cells, which is roughly inverted compared to control mice (Hevner et al., 2004; Pla et al., 2006). This phenotype does not depend on Reelin signaling, but rather on the abnormal distribution of pyramidal cells (Pla et al., 2006). Subsequent work revealed that experimental manipulations causing the formation of clusters of misplaced pyramidal cells under the white matter are sufficient to misguide interneurons into these ectopic locations (Lodato et al., 2011). These results reinforced the notion that pyramidal cells produce factors that guide the migration of cortical interneurons, and our experiments now reveal that *Nrg3* is one of these molecules. In addition, recent work has shown that *Cxcl12* may also play additional functions in the postnatal differentiation of cortical interneurons, affecting their laminar distribution and synapse development (Vogt et al., 2014; Wu et al., 2016).

Recent work suggests that microglia may also regulate the laminar positioning of cortical interneurons (Squarzoni et al., 2014). Microglia invade the cortex following a gradient similar to interneurons (Cunningham et al., 2013; Squarzoni et al., 2014; Swinnen et al., 2013). In the absence of microglia, or when microglia is abnormally activated, MGE-derived interneurons enter the CP prematurely and adopt abnormal laminar distribution patterns (Squarzoni et al., 2014). It remains to be tested whether microglia exert their function through the regulation of chemokines or by directly acting on signals produced by pyramidal cells.

Nrg3, Cortical Interneurons, and Psychiatric Disorders

Human genetic studies have associated variation in the *NRG3* gene with attention deficit/hyperactivity disorder (ADHD) and schizophrenia (Chen et al., 2009; Kao et al., 2010; Meier et al., 2013; Morar et al., 2011; Sonuga-Barke et al., 2008). Although some of these studies await replication in larger genome-wide association studies (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014), increasing evidence suggests that pathological variants in the neuregulin signaling pathway are present in at least a subset of phenotypically distinct patients (Hatzimanolis et al., 2013). In mice, *Nrg3* overexpression in the adult prefrontal cortex increases impulsivity (Loos et al., 2014), a feature that is observed in ADHD and schizophrenia patients (Aron and Poldrack, 2005; Nolan et al., 2011). Loss of *Nrg3* in mice has also been linked with behavioral deficits observed in psychotic disorders (Hayes et al., 2016). Considering that the balance between excitatory and inhibitory circuits seems compromised in schizophrenia patients, and that this might be caused by subtle defects in the function of GABAergic interneurons (Lewis et al., 2012), our results suggest a possible mechanism through which changes in *NRG3* expression may influence the assembly of inhibitory circuits in the cerebral cortex.

EXPERIMENTAL PROCEDURES

Animals

This study was performed in strict accordance with Spanish, United Kingdom, and European Union regulations. The local ethical committees at the Instituto de Neurociencias and King's College London approved all experimental procedures involving animals.

Nkx2-1Cre (Xu et al., 2008) and *Ai9* (*Rosa26 Reporter* CAG-boosted *tdTomato*) mice (Madisen et al., 2010) were maintained in a C57BL/6 background, whereas wild-type and ubiquitously expressing GFP mice (Hadjantonakis et al., 1998) were maintained in a CD1 background. *HER4^{heart}* transgenic mice, which express a human ErbB4 (*HER4*) cDNA under the control of the cardiac-specific myosin heavy-chain (HMC) promoter, were maintained in a mixed C57b/6 and 129/SvJ background. The generation of *ErbB4* mutant mice (Gassmann et al., 1995) and *HER4^{heart}* transgenic mice (Tidcombe et al., 2003) has been previously described. *Lhx6-Cre;ErbB4^{F/F}* mice were generated by breeding *Lhx6-Cre* mice (Fogarty et al., 2007) with mice carrying loxP-flanked (F) *ErbB4* alleles (Golub et al., 2004). For birth-dating experiments, pregnant females received intraperitoneal injections at E12.5 (three injections in 18 hr) or E15.5 (three injections in 12 hr) with 50 mg/kg 5-bromo-2'-deoxyuridine (BrdU) (B5002; Sigma-Aldrich).

We generated a pre-conditional allele for *Nrg3* by crossing mice carrying a knockout first *Nrg3* allele (Knockout Mouse Program of the University of California, Davis) (Skarnes et al., 2011) with mice ubiquitously expressing Flp (Rodríguez et al., 2000). We then produced conditional *Nrg3* mutants by crossing *Nex-Cre* (Goebbels et al., 2006) or *Nestin-Cre* (Tronche et al., 1999) with mice carrying conditional *Nrg3* alleles. To genotype these mice, we used the following primer sequences: CSD-loxP, 5'-GAGATGGCGCAACGCAATTAA TG-3'; CSD-*Nrg3*-SR1: 5'-AGTGCTGGAATAAAGCATGGTGGG-3'; CSD-*Nrg3*-wtF: 5'-CATATTACATACAGAATTCGAAGATAGGC-3'; CSD-*Nrg3*-wtR: 5'-CCAGTGCTGGAATTTGAATACAA-3'. CSD-loxP and CSD-*Nrg3*-SR1 primers were used to detect the knockout first allele. CSD-*Nrg3*-wtF and CSD-*Nrg3*-wtR were used to detect the wild-type allele and the wild-type pre-conditional allele generated after Flp-mediated recombination.

In Situ Hybridization and Immunohistochemistry

For in situ hybridization, postnatal mice were perfused transcardially with 4% paraformaldehyde (PFA) in PBS, and the dissected brains were fixed overnight at 4°C in the same solution. Brains were then cut at 40 μm on a freezing microtome, and free-floating coronal sections were subsequently hybridized with digoxigenin-labeled probes as described before (Flames et al., 2007). The following cDNA probes were used in this study: *Nrg3* and *ErbB4* (kindly provided by Cary Lai, Indiana University, Bloomington), *GAD67* (kindly provided by John Rubenstein, UCSF), *Lhx6* (kindly provided by V. Pachnis, The Crick Institute, London), *Cxcr4* (Invitrogen; 174412), *Cxcl12* (Invitrogen; 3483088), *Cdh6* and *Cdh9* (kindly provided by C. Redies, University of Jena), *Ephb6* (Source BioScience; EST clone IMAGp998L1511952Q), *Epha6* (kindly provided by V. Borrell, Instituto de Neurociencias, Alicante), and *Sema7a* (Source BioScience; EST clone IMAGp998I188236Q).

For immunohistochemistry, postnatal mice were perfused transcardially with 4% PFA in PBS, and the dissected brains were fixed for 2 hr at 4°C in the same solution. Brains were sectioned at 60 μm on a vibratome or 40 μm on a freezing microtome, and free-floating coronal sections were then subsequently processed for immunohistochemistry as previously described (Pla et al., 2006). The following primary antibodies were used: rat anti-BrdU (1:200; ab6326; Abcam), rat anti-Ctip2 (1:500; ab18465; Abcam), rabbit anti-Cux1 (CDP-M222; 1:100; Santa Cruz), rabbit anti-DsRed (1:500; 632496; Clontech), chicken anti-GFP (1:1,000; GFP-1020; Aves Labs), and rabbit anti-PV (1:3,000; Swant). The following secondary antibodies were used: goat anti-chicken 488, donkey anti-rabbit 555, donkey anti-mouse 488, and goat anti-rat IgG (H+L) Alexa Fluor 555 conjugate (Molecular Probes). Cell nuclei were stained with 5 μM DAPI in PBS, and sections were mounted with Mowiol (Sigma) with NPG (Calbiochem).

In Utero Electroporation

E14.5 timed-pregnant ICR or *Nkx2-1Cre;Ai9* females were deeply anesthetized, and the abdominal cavity was cut open. Embryos were exposed in the

uterus, and 1 $\mu\text{g}/\mu\text{L}$ pCAG-Gfp or *Nrg3* (kindly provided by C. Lai, Indiana University, Bloomington, and subcloned into pCAGGS) plasmids were injected into the lateral ventricle of the telencephalon through the uterine wall. Square electric pulses of 45 V and 50 ms were passed through the uterus five times, spaced 950 ms, using a square pulse electroporator. The uterine horns were placed back in the abdominal cavity, which was then suture closed, and the female was allowed to recover.

Explant Cultures

For COS cell confrontation assays, COS7 cells were transfected with plasmids encoding *Rfp* alone, *Rfp* and *Cxcl12*, *Rfp* and *Nrg3*, *Rfp* and *CRD-Nrg1*, or *Rfp* and *Ig-Nrg1*, and cell aggregates were prepared by diluting transfected cells with Matrigel in a 1:1 proportion. After jellification, COS cell aggregates were cut with a scalpel in small rectangular prisms of approximately 400 \times 400 \times 800 μm and confronted to MGE explants obtained from GFP-expressing transgenic mice in 3D Matrigel pads. The cDNA used for expression of *Cxcl12* was obtained from Invitrogen (clone number: 3483088; accession number: BC006640). *Nrg3* was kindly provided by Cary Lai (Indiana University, Bloomington). The sequences used for expression of type I NRG1 (*Ig-Nrg1*) and type III NRG1 (*CRD-Nrg1*) correspond to the accession numbers AY648976 and AY648975, respectively. For *Cxcl12* chemokine-blocking experiments, SU6656 (Sigma; 330161-87-0) was added to the medium at a final concentration of 15 μM . Previous work has shown that Src functions downstream of *Cxcr4* activation (Cabioglu et al., 2005).

In Vitro Focal Electroporation

Coronal slice cultures were obtained as described previously (Anderson et al., 1997). A pCAGG-based *dsRed* plasmid was pressure injected focally into the MGE of coronal slice cultures by a Pneumatic PicoPump through a glass micropipette. Slices were then electroporated within a setup of two horizontally oriented platinum electrodes powered by a Electro-Square-Porator, as described before (Flames et al., 2004).

Time-Lapse Videomicroscopy

Slices were transferred to the stage of an upright Leica DMLFSA or inverted Leica DMIRE2 microscope coupled to a confocal spectral scanning head (Leica; TCS SL) and viewed through 10–60 \times water immersion or 20 \times oil objectives. Slices were continuously superfused with warmed (32 $^{\circ}\text{C}$) artificial cerebrospinal fluid at a rate of 1 mL/min or maintained in supplemented Neurobasal medium. To block *Cxcl12* function, SU6656 (Sigma; 330161-87-0) was added to the medium at a final concentration of 15 μM .

Stripe Assay

Purified CXCL12 protein was obtained from PeproTech (250-20A) and used at 1 ng/ μL . GST and EGF-Nrg3-GST were purified using standard protocols and used at 10 $\mu\text{g}/\text{mL}$. Alternating lanes, 50 μm wide, were laid down on a polylysine-coated plastic dish. Alexa 555-labeled anti-rabbit IgGs were added to the GST, EGF-Nrg3-GST, and CXCL12 protein solution for lane identification. The lanes were further coated with laminin. MGE explants were dissected out of GFP $^{+}$ brain slices, plated on top of the protein stripes, and incubated in methylcellulose-containing Neurobasal medium for 48 hr.

FACS

We dissected the sensorimotor cortex of E17.5 embryos and P4 pups following in utero electroporation at E14.5. Cortical tissue was dissociated as described previously (Catapano et al., 2001). GFP $^{+}$ cells were purified using fluorescent activated cell sorting (FACSARIA III; BD Biosciences), and the resulting pellet was kept at -80°C .

TaqMan Gene Expression Assays

We isolated GFP $^{+}$ pyramidal cells by FACS at E17.5 and P4 after in utero electroporation at E14.5. mRNA was then extracted using the RNeasy Micro Kit (QIAGEN) according to the manufacturer's instructions. RNA quality was assessed using a bioanalyzer (Agilent Technologies) and then retro-transcribed into single-stranded cDNA. The RNA was sent to Unidad Genómica (Fundación Parque Científico de Madrid) for quality control and retro-transcription. Relative gene expression levels from three independent samples were

analyzed using custom designed TaqMan low-density array (TLDA) plates (Micro Fluidic Cards; Applied Biosystems). Each plate contained duplicates for all the genes shown in Table S1. Data were collected and analyzed using the threshold cycle (Ct) relative quantification method. The housekeeping gene 18 RNA was included in the array for assessing RNA quality and sample normalization.

Western Blot

Cortical lysates were prepared from P30 control and *Nestin-Cre;Nrg3^{F/F}* and *Nex-Cre;Nrg3^{F/F}* mutants as described before (Fazzari et al., 2010; Vullhorst et al., 2009) and blotted using mouse anti- β -Actin (1:4,000; Sigma) and rabbit anti-Nrg3 (1:500; Abcam). Signals were detected with a luminescent image analyzer (LAS-1000PLUS; Fujifilm) and quantified with Quantity One 1D Analysis Software (Bio-Rad Laboratories).

Image Analysis and Quantification

Images were acquired using fluorescence microscopes (DM5000B, CTR5000, and DMIRB from Leica, or Apotome.2 from Zeiss) coupled to digital cameras (DC500 or DFC350FX, Leica; OrcaR2, Hamamatsu), or in an inverted Leica TCS SP8 confocal microscope. All images were analyzed with ImageJ (Fiji). For the quantification of migration in MGE explants, the distance migrated by the 30 furthest cells was measured. For the quantification of short-range chemoattraction, the colocalizing area between MGE and COS cells was measured. For the analysis of the interneuron angle of migration, we draw a grid of virtual radial lines (lines perpendicular to the ventricular zone and the pial surface) and oriented each cell in relation to the most adjacent "radial line." Cells that deviated less than 25 $^{\circ}$ from radial lines were considered as radially oriented; those that deviate more than 25 $^{\circ}$ were designated as tangentially oriented. We systematically exclude from this analysis those cells located in the more lateral or medial regions of the cortex, so that the curvature of the slice in those regions would not interfere with our analysis (Martini et al., 2009). For the quantification of cell migration in MGE explants, we measured the distance migrated by the 30 furthest cells and normalized the average migrated distance to the distance between MGE and COS explants. For the quantification of the colocalizing area between migrating interneurons and COS cells in the short-distance confrontation assays, we quantified the colocalizing area using ImageJ (Fiji). Stripes were quantified by counting the number of neurons contained in a virtual grid containing five black and five red lines. The same area was used for all explants. Sections from control and mutant mice were imaged during the same imaging session. Data acquisition was performed using the same laser power, photomultiplier gain, pinhole, and detection filter settings (1,024 \times 1,024 resolution; 12 bits). Quantifications were done using ImageJ (Fiji). Layers were drawn following nuclear staining. For in situ hybridization, the area quantified was divided in ten equal bins, and the percentage of cells in each bin was calculated. The bins were then matched to the appropriate layers.

Statistical Analyses

Statistical analysis was carried out in SPSS (SPSS, Inc.). The p values below 0.05 were considered statistically significant. Data are presented as mean and SEM throughout the manuscript (Table S3). Normality and variance tests were first applied to all experimental data. When data followed a normal distribution, paired comparisons were analyzed with t test, whereas multiple comparisons were analyzed using either ANOVA with post hoc Bonferroni correction (equal variances) or the Welch test with post hoc Games-Howell (different variances). A χ^2 test was applied to analyze the distribution of cells in either bins or layers.

SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures, three tables, and two movies and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2016.12.089>.

AUTHOR CONTRIBUTIONS

Conception and Design, Acquisition of Data, Analysis and Interpretation of Data, Drafting or Revising the Article, and Contribution of Unpublished

Essential Data or Reagents, G.B., J.A.S.-A., and O.M.; Acquisition of Data, Analysis and Interpretation of Data, and Drafting or Revising the Article, C.O., C.G.-F., and M.V.

ACKNOWLEDGMENTS

We thank I. Andrew, S. Bae, M.A. Casillas, M. Fernández, and T. Gil for excellent technical assistance and laboratory support; A. Caler for excellent support with FACS experiments; G. Expósito for support with imaging; L. Lim for help with quantitative methods; V. Borrell, R. Hevner, C. Lai, V. Pachnis, C. Redies, B. Rico, J.L.R. Rubenstein, and M. Tessier-Lavigne for plasmids and antibodies; and A. Barco, M.A. Nieto, and K. Nave for mouse strains. We are grateful to members of the Flames, O.M., and Rico laboratories for stimulating discussions and ideas. This work was supported by grants from European Research Council (ERC-2011-AdG 293683) and the Spanish Government (CSD2007-00023 and SAF2011-28845) to O.M. O.M. is a Wellcome Trust Investigator.

Received: November 2, 2016

Revised: December 8, 2016

Accepted: December 27, 2016

Published: January 31, 2017

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