Target product profiles: leprosy diagnostics

Petra Kukkaro,^a Sundeep Chaitanya Vedithi,^b David J Blok,^c Wim H van Brakel,^d Annemieke Geluk,^e Aparna Srikantam, David Scollard, Linda B Adams, Mathias Duck, Sunil Anand, Andie Tucker, Israel Cruz, VRR Pemmaraju, Daniel Argaw Dagne, Kingsley Asiedu & Christopher Hanna

Abstract The World Health Organization (WHO) aims to reduce new leprosy cases by 70% by 2030, necessitating advancements in leprosy diagnostics. Here we discuss the development of two WHO's target product profiles for such diagnostics. These profiles define criteria for product use, design, performance, configuration and distribution, with a focus on accessibility and affordability. The first target product profile outlines requirements for tests to confirm diagnosis of leprosy in individuals with clinical signs and symptoms, to guide multidrug treatment initiation. The second target product profile outlines requirements for tests to detect Mycobacterium leprae or M. lepromatosis infection among asymptomatic contacts of leprosy patients, aiding prophylactic interventions and prevention. Statistical modelling was used to assess sensitivity and specificity requirements for these diagnostic tests. The paper highlights challenges in achieving high specificity, given the varying endemicity of M. leprae, and identifying target analytes with robust performance across leprosy phenotypes. We conclude that diagnostics with appropriate product design and performance characteristics are crucial for early detection and preventive intervention, advocating for the transition from leprosy management to prevention.

Abstracts in عربى, 中文, Français, Русский and Español at the end of each article.

Introduction

Leprosy, a neglected tropical disease, is caused by Mycobac*terium leprae* or less often by *M. lepromatosis.*¹ The disease is a chronic, moderately infectious condition affecting mostly skin, peripheral nerves, mucosa of upper respiratory tract and eyes.^{2,3} Approximately 200 000 new cases are reported annually from nearly 120 countries.4 In 2021, the World Health Organization (WHO) launched the Towards zero leprosy: global leprosy (Hansen's disease) strategy 2021-2030, aiming for a global reduction of 70% in new leprosy cases by 2030.5 Continued investment in leprosy diagnostics is crucial if we are to achieve the proposed targets.

Leprosy is an important public health problem due to its potential for causing lasting physical impairments and adverse socioeconomic consequences if left undiagnosed.^{6,7} M. leprae is moderately contagious and infections can become chronic. Infected contacts can remain asymptomatic for up to 20 years, 8-10 and indirect evidence suggests that these individuals with subclinical infection could transmit M. leprae to close contacts. 10-13 Therefore, conducting regular contact tracing and testing, and administering prophylactic treatment is crucial to interrupt transmission cycles. 11 Despite multidrug treatment availability and global advancements in leprosy treatment, delayed diagnosis remains a substantial concern as it can lead to grade 2 disabilities (ulcers, contractures, foot drop, lagophthalmos, and muscle wasting). 14,15 The global reduction in leprosy cases, ¹⁶ along with less active engagement from health-care professionals in managing the disease, has led to a decline in clinical public health expertise in diagnosing leprosy, further causing delays in diagnosis. 15,17,18

Currently, leprosy diagnosis primarily relies on defined clinical criteria. 19 Microscopic or laboratory-based diagnosis using acid-fast bacilli identification in a slit-skin smear or skin biopsy is used in numerous leprosy programmes and tertiary care settings.¹⁹ Additionally, various point-of-care tests and laboratory assays have been developed to detect M. leprae infection directly or indirectly.²⁰⁻²² These include enzyme-linked immunosorbent assays and lateral flow assays for detection of immunoglobulins and polymerase chain reaction for pathogen detection. 23,24 Both immunodiagnostics and molecular assays are sensitive enough to diagnose multibacillary leprosy²³ as well as some paucibacillary cases.²⁵ Although direct diagnosis of paucibacillary leprosy is challenging, in vitro stimulation followed by detection of immunity against M. leprae antigens, increases diagnostic potential.^{26,27}

Variability in leprosy presentation, patient type and diagnostic targets complicate accurate testing. 28,29 Furthermore,

Correspondence to Andie Tucker (email: atucker@taskforce.org).

(Submitted: 18 September 2023 – Revised version received: 22 November 2023 – Accepted: 6 December 2023 – Published online: 22 February 2024)

^a Novartis Pharma AG, Basel, Switzerland.

^b Department of Biochemistry, University of Cambridge, Cambridge, England.

^c Department of Public Health, Erasmus University Medical Center, Rotterdam, Kingdom of the Netherlands.

^d Until No Leprosy Remains, Amsterdam, Kingdom of the Netherlands.

^e Department of Infectious Diseases, Leiden University Medical Center, Leiden, Kingdom of the Netherlands.

f Clinical and Laboratory Research Division, Blue Peter Public Health and Research Centre-LEPRA Society, Hyderabad, India.

⁹ National Hansen's Disease Program, Baton Rouge, United States of America (USA).

^h The Leprosy Mission International, Brentford, England.

ⁱ American Leprosy Missions, Hyderabad, India.

¹ The Task Force for Global Health, Inc, Decatur, USA.

^k National School of Public Health, Instituto de Salud Carlos III, Madrid, Spain.

¹ Global Leprosy Programme, WHO Regional Office for South-East Asia, New Delhi, India.

Department of Control of Neglected Tropical Diseases, World Health Organization, Geneva, Switzerland.

ⁿ Global Project Partners, Oakland, USA.

limited awareness about leprosy among health-care workers poses a diagnostic challenge.³⁰ Hence, diagnostic tests that support rapid contact tracing and screening are essential for efficient and comprehensive leprosy control programmes. Easy-to-use diagnostic tests are therefore needed to help reduce delays.

In efforts to achieve better performance, some tests lean on complex instrumentation and expertise that limit their field use, especially in lowresource settings. Additionally, tests requiring invasive sampling are challenging to deploy in a field setting.³¹ These problems underscore the current need to develop diagnostic tests designed for settings where they are most needed.32

To facilitate early prophylactic interventions to disrupt the chain of leprosy transmission, the Global Partnership for Zero Leprosy, under WHO's diagnostic technical advisory group guidance,33 developed two target product profiles for high-priority leprosy diagnostics. These profiles ensure that the diagnostic products not only meet the necessary performance criteria but also consider the specificities of the intended health-care context and the patient demographic. The first target product profile covers confirmatory diagnostic tests for individuals presenting with clinical manifestations indicative of leprosy, with the goal of initiating multidrug therapy. The second target product profile covers diagnostic assays for the detection of *M. leprae* infection in asymptomatic households or familial contacts of individuals with confirmed clinical leprosy.

Methods

Development process

To create target product profiles and guide product developers, the Global Partnership for Zero Leprosy formed a leprosy-focused diagnostic expert working group to assist WHO's diagnostic technical advisory group's skin neglected tropical disease subgroup. The working group assisted by clarifying unmet public health needs; determining whether existing available target product profiles or pipeline products are addressing current needs; defining the scope of needed new target product profiles; and serving as a scientific group to develop new target product profiles.

The Global Partnership for Zero Leprosy included leprosy experts working in laboratory, field research and clinical capacities, as well as community stakeholders who developed target product profiles. The group collaborated with the WHO diagnostic technical advisory group, WHO Technical Advisory Group on Leprosy Control, and consulted experts at the Bill & Melinda Gates Foundation. The original draft version criteria were chosen by the Global Partnership for Zero Leprosy's diagnostic working group using methods such as landscape assessments, use case needs analyses and diagnostic performance modelling, all designed through an internal consultative process.

The Global Partnership for Zero Leprosy's diagnostic working group reviewed the need for a leprosy diagnostic test using WHO reports, literature and outcomes of discussions with the experts. We created the first version of target product profile using the quality by design planning method,34,35 with performance characteristics based on statistical analysis and modelling by expert group members (online repository).36 Based on feedback from diagnostic technical advisory group members, we adapted the first version before publishing the document on the WHO website for public consultation for one month (30 November to 30 December 2021). In addition to the online public consultation, the WHO Technical Advisory Group on Leprosy Control also reviewed the first version. To finalize the target product profile, we addressed all comments received on the first version, and subsequently the chair of the diagnostic technical advisory group's skin neglected tropical diseases subgroup and a WHO technical staff member reviewed the document.

Target product profiles^{9,10}

WHO finalized and disseminated final version of the target product profiles on 24 July 2023. 9,10 Target product profile 1 describes a test to confirm leprosy in individuals presenting with clinical signs and symptoms (hereafter confirmatory test). Target product profile 2 describes a point-of-care test for the detection of analytes specific to M. leprae or host response to M. leprae to enable detection

of subclinical M. leprae infections (hereafter test for subclinical infection).10

The WHO diagnostic target product profiles define minimal and ideal targets for each profile and organize them into five categories: (i) product use summary; (ii) design; (iii) performance; (iv) product configuration; and (v) product costs and distribution channels. Minimal refers to the lowest acceptable output for a characteristic for the test to be suitable for the intended use, and ideal reflects targets that may be harder to achieve but would accelerate access, adoption and clinical outcomes.

Product use summary

For a confirmatory test, the intended application is at minimum a laboratorybased assay for the qualitative and quantitative detection of biomarkers specific to M. leprae and, ideally, M. lepromatosis. This test should be able to confirm diagnosis of clinical leprosy in individuals exhibiting clinical manifestations. In contrast, the subclinical test delineates specifications for a point-of-care, rapid diagnostic tool aimed at identifying biomarkers pertinent to M. leprae or the host immune response to M. leprae or M. lepromatosis. Such a test should be applicable in contact tracing scenarios, and facilitate detection of asymptomatic M. leprae infections among contacts of leprosy patients. The ideal intended use for both profiles is deployment as pointof-care diagnostics.

A confirmatory test should require minimal infrastructure, characterized by a laboratory setting where technicians with less than one week of additional formal training can perform the assay. The ideal scenario for such a test is a pointof-care format executable in health-care settings without any laboratory infrastructure. The ideal intended user profile is health-care professionals, community health workers and volunteers; requiring only a one-day training, complemented by easy and accessible usage instructions. For tests for subclinical infection, the prerequisites for infrastructure, end-user capability and training are consistent with the ideal conditions described for confirmatory tests.

Design

For confirmatory tests, at the minimal level, portability requisites for a laboratory-based assay stipulate that transport and portability conditions should not exceed those of standard laboratory apparatus. The test should be designed to use electricity supplied by main lines and laboratory-grade water resources (such as distilled water). Should instrumentation require periodic maintenance and calibration, it should be feasible within the recipient countries and not more than once per calendar year. Acceptable specimen types include capillary blood via fingerstick, venous blood, collected urine, nasal swabs, slit-skin smears and punch biopsies, with the latter permitting sub-millimetre tissue collection. Sample processing and transfer should be simple, necessitating a single holding tube with a 500 µL capacity and disposable transfer pipette for one-time use. The maximum sample volume should not surpass 100 µL. Confirmatory tests should aim to detect biomarkers uniquely associated with M. leprae and provide semiquantitative analysis of bacterial load or immune response. An instrument-based detection method should incorporate an external process control indicator. All necessary reagents and operational supplies must comply with the basic importation restrictions and ensure the safety of the operator.

In an ideal scenario, confirmatory tests should be done on a highly portable point-of-care device without specialized transport requirements. The device should be battery powered or otherwise not depend on mains power and availability of water, obviating the need for regular maintenance or calibration. Sample collection is confined to capillary blood via fingerstick, urine or nasal swabs, with straightforward processing and single-use pipette transfer. Ideally, requisite sample volume is less than 10 μL. The assay should quantitatively determine biomarkers specific to both M. leprae and M. lepromatosis, assessing bacterial load or immune status. Results should be discernible to the unaided eye, marked by stark contrast and clarity. The required provisions for quality control, necessary supplies and safety protocols mirror those at the minimal level.

For a test for subclinical infection, the prerequisites for portability, and power and water independence are consistent for both minimal and ideal characteristics. The test's portability should negate the need for specialized transportation, mains electricity and water supply. At the minimum level, any field-compatible equipment employed (e.g. sample incubator, reader) should require only basic maintenance or cali-

bration, potentially facilitated through return to the manufacturer or execution of a standard procedure. In the ideal scenario, the reader should require neither maintenance nor calibration. Both minimal and ideal acceptable sample types include capillary blood, collected urine and nasal swabs, with venous blood and slit-skin smear included at the minimal level only. Sample volumes are confined to less than 100 µL for the minimal scenario and less than 10 µL for the ideal scenario. The minimal requirement is identification of biomarkers indicative of latent M. leprae infection, whereas in the ideal scenario the test also identifies M. lepromatosis. Both in minimal and ideal scenarios a qualitative output is favoured, with results clearly visible to the naked eye, and the test must have an internal process control indicator.

Performance

For confirmatory tests, a minimal diagnostic assay should display a clinical sensitivity of ≥90% and a specificity of \geq 99% for the detection of *M. leprae*. The assay should yield results within 4 hours, and these results must maintain their stability for at least 30 minutes post-analysis. Operational throughput should exceed 100 tests per technician per day. Assay stability should be ≥ 18 months when stored at temperatures ranging from 4 °C to 40 °C and at 75% relative humidity. The testing procedure should be limited to maximum 15 user steps, of which a maximum of five steps should be timed.

An ideal confirmatory test would be capable of detecting both M. leprae and M. lepromatosis, maintaining a clinical sensitivity of \geq 90% but with a specificity of ≥ 99.9%. A field-deployable version of the assay should deliver results in less than 30 minutes, with the stability of results extending to at least 24 hours. Operational throughput should surpass 10 tests per technician per hour. The stability criterion for the ideal test should extend to ≥24 months under the aforementioned temperature and humidity conditions. Conducting the analysis should be possible by performing maximum five steps, out of which no more than one should be timed.

For a test for subclinical infection, the minimal test should have a clinical sensitivity of \geq 81% and a specificity of \geq 99.5% for the detection of *M. leprae*. The time to results should be less than 2 hours, and these results should main-

tain their stability for at least 30 minutes post-analysis. The expected throughput is more than seven tests per tester per hour. The stability of these assays should be no less than 18 months within the temperature range of 4 °C to 40 °C and at maximum 75% relative humidity. The analysis should be maximum two timed steps and maximum eight user steps.

An ideal test for subclinical infection would detect both M. leprae and M. lepromatosis with a clinical sensitivity of \geq 94% and a specificity of \geq 99.9%. The test should produce results in less than 30 minutes, with results remaining stable for at least 24 hours. The throughput should be more than 10 tests per tester per hour, with stability guaranteed for \geq 24 months under the above specified storage conditions. The test should be designed for ease of use, with maximum one timed step and maximum five user steps. Both minimal and ideal tests should yield binary outcomes.

Product configuration

A minimal confirmatory test must adhere to the relevant standards, such as ASTM International (ASTM) D4169–05 and international standard organization (ISO) 11607–1:2006, or their equivalents. Test components or consumables for laboratory use should be able to be stored and shipped at temperatures ranging from 0 °C to 4 °C. Cold storage is an acceptable condition for any laboratory-based assays. For laboratory-based tests, support should be available from the equipment manufacturer for problemsolving and use of the equipment.

All materials included in the assay should be universally compatible with standard laboratory biohazard waste management protocols. Labelling and instructions must comply with the pertinent CE Mark under In Vitro Diagnostic Regulation stipulations (or other recognized regulatory authorities, such as the United States Federal Drug Administration under 21 CFR 820), alongside guidelines set forth by the WHO prequalification processes.³⁷

The ideal product configuration for confirmatory tests and both the minimal and ideal scenarios for tests for subclinical infection have the same requirements. These assays should be point-of-care tests that adhere to the specified ASTM and ISO standards or their accepted equivalents, eliminating the need for cold-chain transport. They should be storable at ambient

Fig. 1. Comparative analysis matrix for target product profiles for leprosy diagnostics

Category

Product use summary	Design	Performance	Product configuration Product cost and channels
Intended use	Portability	Species differentiation or detection	Shipping conditions Target pricing per test
Target population	Instrument or power requirement	Diagnostic or clinical sensitivity	Storage conditions Capital cost
Lowest infrastructure level	Water requirement	Diagnostic or clinical specificity	Service and support Product lead times
Lowest level user	Maintenance and calibration	Time to results	Waste disposal Target launch countries
Training requirements	Sample type/collection	Result stability	Labelling and instructions for use Product registration (substantiation to regulatory body of product claims)
	Sample preparation or transfer device	Throughput	Level of agreement between profiles Unique requirements for each type of test and performance level Same requirements for both tests for one of the performance levels Same requirements for ideal confirmatory test and both minimal and ideal subclinical test
	Sample volume	Target shelf-life or stability	
	Target analyte	Ease of use	
	Type of analysis	Ease of results interpretation	
	Detection	Operating temperature	
	Quality control		
	Supplies needed		Same requirements for both tests across performance levels
	Safety		Same requirements for both performance levels for at least one of the tests, but different requirements between tests

Note: Target product profile 1 is a diagnostic test to confirm leprosy in individuals presenting with clinical signs and symptoms, and target product profile 2 is a diagnostic test to identify Mycobacterium leprae infection in asymptomatic household and family contacts of diagnosed leprosy patients.¹⁰

temperatures ranging from 2 °C to 40 °C without requiring service interventions. The assays must not include any materials that are not compatible with standard biohazard waste disposal procedures in a laboratory environment. Packaging must consider daily throughput to minimize unnecessary waste. Finally, the labelling and usage instructions should align with those established for the minimal requirements for confirmatory tests.

Product cost and channels

The minimal requirements for the cost of a confirmatory test are below 3 United States dollars (US\$). Capital expenditure for the deployment of such tests should remain within a threshold of US\$ 5000. The anticipated lead time for product availability should be less than eight weeks. The market introduction should be focused on countries with endemic leprosy, with regulatory prerequisites encompassing: (i) compliance with CE Mark under In Vitro Diagnostic Regulation or other relevant stringent regulatory authorities; (ii) export certifications from the country of manufacture; (iii) WHO prequalification, contingent upon necessity and relevance; and (iv) national registration in accordance with the regulatory demands of target countries.

The pricing for an ideal confirmatory test is set at below US\$ 1, not accounting for additional expenses like logistics, storage and other operational costs related to national procurement for neglected tropical diseases programmes. Capital costs for these laboratory-based tests should not exceed US\$ 5000 in a minimal scenario; however, ideally, given the point-of-care nature of the test, no capital investment would be required. The expected lead time for the product should be less than six weeks. The target markets and required registrations for launch should match those outlined for the minimal test.

For a test for subclinical infection, the financial and distribution criteria remain consistent with those for the ideal confirmatory test, with the stipulation that capital costs can go up to US\$ 2000 for both the minimal and ideal versions. Minimally, the contact-tracing test is specifically designed for countries that are actively involved in leprosy contact tracing and post-exposure prophylaxis programmes.

Comparative analysis

When comparing 38 requirements across the five categories for both minimal and ideal requirements, we found that 10.5% (4/38) of the requirements were identical for both profiles, regardless of whether we were looking at the

minimal or ideal criteria. When comparing the ideal confirmatory test with both levels of test for subclinical infection, 36.8% (14/38) of the requirements were the same. Furthermore, 34.2% (13/38) of the requirements were alike for both types of tests when considering either the minimal or ideal scenario. Only 18.4% (7/38) of the requirements were different between the two types of tests (Fig. 1).

Discussion

Here we have outlined the minimal and ideal requirements listed in the target product profiles for a confirmatory test and contact-tracing test for leprosy.9,10 Development of a new test complying with the requirements could improve testing outcomes, avoiding complications such as unethical treatment, emotional and physical effects, social exclusion and higher costs.38

Three high-risk factors must be considered to ensure the successful development of a diagnostic test for leprosy, as highlighted for both target product profiles. First, the diagnostic tests aim to identify biomarkers indicative of active infection with M. leprae and M. lepromatosis. However, M. leprae may remain dormant for years and reactivate under certain immune conditions, complicating the identification of recent infections. Despite the challenges and time required to qualify and validate new markers, it is crucial to advance test development using the currently available analytes to prevent delays in development. Second, due to the similar clinical presentations of other mycobacterial infections (like M. tuberculosis) and skin disorders prevalent in the same areas as leprosy is prevalent, the tests must differentiate between these conditions. The development of multiplex assays, which can detect multiple diseases simultaneously, could streamline patient care and diagnostics. Third, even in highly endemic areas, the prevalence of leprosy cases is so low that it poses unique diagnostic challenges; the specificity requirements are high;³⁹ and the tests must be highly specific to avoid false positives and unnecessary treatment. While test developers may find these performance targets challenging, they are crucial to achieve and maintain low rates of both false positives and false negatives, especially as diseases approach elimination. Perfect accuracy is rarely possible with a single test. However, using multiple tests together, either serial testing or parallel testing, can improve the overall accuracy. Serial testing uses a sequence of more precise tests to confirm a diagnosis, while parallel testing checks for several disease indicators at once to enhance the detection process.39

The target product profile outlines that one of the minimal requirements for a confirmatory test is high specificity for all types of leprosy, including manifestations with low bacilli level. While such specificity can be difficult to achieve, some studies demonstrated adequate sensitivity across paucibacillary and multibacillary leprosy. 40-43 To ensure adequate performance across leprosy manifestations, clinical validation studies should be set up at multiple sites worldwide to demonstrate that

acceptance criteria are met for all types of leprosy.

As leprosy incidence decreases, and confirmation of diagnosis happens in environments that can support moderately complex evaluations, laboratory-based tests can satisfy the minimal requirements for a confirmatory diagnosis of leprosy. Given the strict clinical sensitivity and specificity requirements, laboratory-based tests might be more suitable than point-ofcare tests to meet these performance criteria. On the contrary, to perform WHO-recommended contact tracing for individuals diagnosed with M. leprae infection, a test that can be implemented at the point-of-care is essential to ensure usefulness in field settings. In all cases, ensuring immediate availability of appropriate medical interventions following the detection of leprosy or M. leprae infection is crucial for ethical reasons.

Currently, WHO only recommends use of single-dose rifampicin for postexposure-prophylaxsis.11 However, in household contacts of newly diagnosed leprosy cases, a single dose of rifampicin may not suffice, as the observed risk reduction for developing leprosy is only 50%-60% and this protection lasts for merely two years after administration.44,45 Thus, additional tools to detect M. leprae infection, together with improved post-exposure prophylaxis, are desirable. Close collaboration, coordination and alignment are required with teams working on other post-exposureprophylaxis regimens, to ensure concurrent availability of these regimens along with the appropriate diagnostic tools. Two trials are currently ongoing: one in Bangladesh, Brazil, India and Nepal, and another in the Comoros. 46,47 In these trials, field teams closely collaborate with researchers developing and evaluating immunodiagnostic and molecular tests for monitoring the direct and longitudinal effects and efficacy of various

forms of post-exposure prophylaxis on development of leprosy.46,47 Clinical validation studies of both diagnostics and treatment interventions may depend on and benefit from each other.

A factor not covered in this research is the impact of stigma associated with leprosy, leading to discrimination against affected individuals and their families, which can hinder timely diagnosis and treatment. Long-standing stigmas associated with leprosy necessitate diagnostic approaches that also consider social factors such as privacy and discretion, similar to contact-tracing efforts undertaken in human immunodeficiency virus testing.

In conclusion, investing in diagnostics for both disease and infection is critical to significantly reduce new cases of leprosy worldwide. WHO target product profiles for leprosy diagnostics can help guide development of appropriate tools. The goal is to not only manage leprosy but also to prevent it, thereby reducing its global burden.

Acknowledgements

Petra Kukkaro and Sundeep Chaitanya Vedithi contributed equally to this work. Israel Cruz is also affiliated with Centro de Investigación Biomédica en Red de Enfermedades Infecciosas (CIBERINfEC). We extend our sincere gratitude to the late Ozorio Moraes, our colleague who passed away before the publishing of this manuscript. We thank Shweta Pitre (SIRO Clinpharm, England), Sangita Patil, (SIRO Clinpharm Pvt Ltd, India) and the Global Partnership for Zero Leprosy.

Funding: The Global Partnership for Zero Leprosy provided funds for writing and publication support.

Competing interests: PK is employed by Novartis Pharma AG and DS is a consultant with Janssen Pharmaceuticals.

© 2024 The authors; licensee World Health Organization.

This is an open access article distributed under the terms of the Creative Commons Attribution IGO License (http://creativecommons.org/licenses/by/3.0/igo/legalcode), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. In any reproduction of this article there should not be any suggestion that WHO or this article endorse any specific organization or products. The use of the WHO logo is not permitted. This notice should be preserved along with the article's original URL.

ملخص

الكشف عن عدوى المتفطرة الجذامية أو متفطرة الجذام بين مخالطي مرضى الجذام، ولا تظهر عليهم الأعراض، مما يساعد في التدخلاتُ الوقائية والعلاجية. تم اللجوء لوضع نموذج إحصائية لتقييم متطلبات الحساسية والخصوصية لهذه الاختيارات التشخيصية. تركز هذه الورقة على التحديات التي تواجه تحقيق خصوصية عالية، في ظل الاستشراء المتباين للمتفطرة الجذامية، وتحديد التحليلات الستهدفة ذات الأداء القوى عبر الناذج الظاهرية للجذام. وننتهي إلى أن التشخيص باستخدام التصميم المناسب للمنتج وخصائص الأداء، هو أمر حيوي للكشف المبكر والتدخل الوقائي، والدعوة إلى الانتقال من إدارة مرض الجذام إلى الوقاية.

ملفات تعريف المنتج المستهدف: تشخيص مرض الجذام تهدف منظمة الصحة العالمية (WHO) إلى خفض حالات الإصابة الجديدة بالجذام بنسبة 70% بحلول عام 2030، مما يستلزم إحراز خطوات للتقدم في تشخيص مرض الجذام. نحن نناقش هنا تطوير اثنين من ملفات تعريف المنتجات المستهدفة لدى منظمة الصحة العالمية لمثل هذه التشخيصات. تحدد هذه الملفات التعريفية معايس استخدام المنتج، وتصميمه، وأداؤه، وتكوينه، وتوزيعه، مع التركيز على إمكانية الحصول عليه، والقدرة على تحمل تكاليفه. يحدد الملف التعريفي الأول للمنتج المستهدف متطلبات الاختبارات لتأكيد تشخيص الإصابة بالجذام لدى الأفراد الذين يعانون من علامات وأعراض إكلينيكية، وذلك لإدارة بدء العلاج بأدوية متعددة. ويحدد ملف تعريف المنتج المستهدف الثاني متطلبات اختبارات

摘要

目标产品简介:麻风病诊断方法

世界卫生组织 (WHO) 的目标是到 2030 年将新发麻风 病病例减少70%,这需要改进麻风病的诊断方法。在本 文,我们将讨论世卫组织针对此类诊断产品制定的两 个目标产品简介。这些产品简介定义了产品使用、设 计、性能、配置和分发的标准,重点关注了产品的可 及性和可负担性。第一份目标产品简介概述了对有临 床体征和症状的个体进行检测以确诊麻风病的要求, 以在启动多药治疗时提供指导。第二份目标产品简介 概述了对麻风病患者的无症状接触者进行检测以查明

麻风分枝杆菌或弥散型麻风分枝杆菌感染情况的要求, 这有助于采取预防性干预和预防措施。统计模型用于 评估这些诊断试验的敏感性和特异性要求。本文考虑 到不同地方麻风分枝杆菌的流行状况,强调了实现高 特异性和识别在各种麻风表型中表现稳定的靶分析物 方面所面临的挑战。我们得出结论,使用具有适当产 品设计和性能特征的诊断产品对于早期检测和采取预 防性干预措施至关重要,倡导从麻风病的诊治转向麻 风病的预防。

Résumé

Profil de produit cible: diagnostic de la lèpre

L'Organisation mondiale de la Santé (OMS) vise à réduire le nombre de nouveaux cas de lèpre de 70% d'ici 2030, ce qui nécessite un meilleur diagnostic de la maladie. Dans le présent document, nous évoquons le développement de deux profils de produit cible établis par l'OMS à cette fin. Ces profils définissent des critères en matière d'utilisation, de conception, de performances, de configuration et de distribution du produit, en accordant une attention particulière à l'accessibilité et à l'abordabilité. Le premier profil de produit cible décrit les exigences pour les tests servant à confirmer le diagnostic de la lèpre chez les individus qui présentent des signes cliniques et des symptômes, afin d'orienter l'instauration d'un traitement à base de plusieurs médicaments. Le second profil de produit cible décrit les exigences pour les tests servant à détecter une infection à Mycobacterium leprae ou M. lepromatosis parmi les contacts asymptomatiques de patients lépreux, ce qui contribue à l'adoption de mesures prophylactiques et à la prévention. Nous avons eu recours à une modélisation statistique pour évaluer les exigences de sensibilité et de spécificité de ces tests diagnostiques. Cet article met en évidence les obstacles à l'atteinte d'un niveau élevé de spécificité en raison de l'endémicité variable de M. leprae, et à l'identification d'analytes cibles offrant de bons résultats chez les phénotypes lépreux. Nous concluons qu'un diagnostic reposant sur des caractéristiques de performance et de conception appropriées est essentiel pour détecter rapidement la maladie et intervenir en amont, et nous plaidons pour une prévention plutôt qu'une gestion de la lèpre.

Резюме

Профиль целевого продукта: диагностика лепры

Всемирная организация здравоохранения (ВОЗ) ставит перед собой задачу сократить число новых случаев лепры на 70% к 2030 году, что требует совершенствования методов диагностики заболевания. В этой статье представлена информация о разработке двух профилей целевых продуктов ВОЗ для таких диагностических средств. На основе этих профилей определяются критерии использования, дизайна, эффективности, конфигурации и распространения продукции с акцентом на физическую и экономическую доступность. В первом профиле целевого продукта изложены требования к тестам для подтверждения диагноза лепры у лиц с клиническими проявлениями и симптомами с целью определения начала лечения несколькими препаратами. Второй профиль целевого продукта определяет требования к тестам для выявления инфекции Mycobacterium leprae или M. lepromatosis среди бессимптомных контактов больных лепрой, что способствует проведению профилактических мероприятий и предотвращению распространения заболевания. Для оценки требований к чувствительности и специфичности этих диагностических тестов использовался метод статистического моделирования. В статье освещаются проблемы достижения высокой специфичности с учетом различной эндемичности М. leprae, а также выявления целевых аналитов, позволяющих надежно определять заболевание при различных фенотипах лепры. Авторы пришли к выводу, что диагностическая продукция с соответствующим дизайном и эксплуатационными характеристиками имеет решающее значение для выявления заболевания на ранних стадиях и проведения профилактических мероприятий, способствуя переходу от лечения лепры к ее профилактике.

Resumen

Perfiles de productos objetivo: diagnóstico de la lepra

La Organización Mundial de la Salud (OMS) pretende reducir los nuevos casos de lepra en un 70% para 2030, lo que requiere avances en el diagnóstico de la lepra. Aquí se analiza el desarrollo de dos perfiles de productos objetivo de la OMS para este tipo de diagnósticos. Estos perfiles definen los criterios de uso, diseño, rendimiento, configuración y distribución de los productos, centrándose en su accesibilidad y asequibilidad. El primer perfil de producto objetivo describe los requisitos de las pruebas para confirmar el diagnóstico de la lepra en personas con signos y síntomas clínicos, con el fin de orientar el inicio del tratamiento con múltiples fármacos. El segundo perfil de producto objetivo describe los requisitos de las pruebas para detectar la infección por Mycobacterium leprae o M. lepromatosis entre los contactos asintomáticos de los pacientes con lepra, para facilitar las intervenciones profilácticas y la prevención. Se utilizaron modelos estadísticos para evaluar los requisitos de sensibilidad y especificidad de estas pruebas diagnósticas. El artículo destaca las dificultades para lograr una alta especificidad, dada la diferente endemicidad de M. leprae, y para identificar analitos diana con un rendimiento sólido en todos los fenotipos de lepra. Concluimos que los diagnósticos con un diseño de producto y unas características de rendimiento adecuados son fundamentales para la detección precozy la intervención preventiva, lo que favorece la transición del manejo de la lepra a la prevención.

References

- 1. Collin SM, Lima A, Heringer S, Sanders V, Pessotti HA, Deps P. Systematic review of Hansen disease attributed to Mycobacterium lepromatosis. Emerg Infect Dis. 2023 Jul;29(7):1376–85. doi: http://dx.doi.org/10.3201/eid2907 .230024 PMID: 37347507
- Das NK, De A, Naskar B, Sil A, Das S, Sarda A, et al. A quality of life study of patients with leprosy attending the dermatology OPD of a tertiary care center of eastern India. Indian J Dermatol. 2020 Jan-Feb;65(1):42-6. doi: http://dx.doi.org/10.4103/ijd.IJD_729_18 PMID: 32029939
- 3. What is Hansen's Disease? [internet]. Atlanta: Centers for Disease Control and Prevention; 2017. Available from: https://www.cdc.gov/leprosy/about/ about.html [cited 2023 Jun 23].
- Leprosy [internet]. Geneva: World Health Organization; 2023. Available from: https://www.who.int/news-room/fact-sheets/detail/leprosy [cited 2023 Jun
- Towards zero leprosy: global leprosy (Hansen's disease) strategy 2021–2030. Geneva: World Health Organization; 2021. Available from: https://iris.who .int/handle/10665/340774 [cited 2023 Jun 23].
- Somar P, Waltz MM, van Brakel WH. The impact of leprosy on the mental wellbeing of leprosy-affected persons and their family members - a systematic review. Glob Ment Health (Camb). 2020 Jun 9;7. doi: http://dx .doi.org/10.1017/gmh.2020.3 PMID: 32742673
- 7. Koschorke M, Al-Haboubi YH, Tseng P-C, Semrau M, Eaton J. Mental health, stigma, and neglected tropical diseases: a review and systematic mapping of the evidence. Front Trop Dis. 2022;3:808955. doi: http://dx.doi.org/10 .3389/fitd.2022.808955
- Froes LAR, Sotto MN, Trindade MAB. Leprosy: clinical and immunopathological characteristics. An Bras Dermatol. 2022 May-Jun;97(3):338-47. doi: http://dx.doi.org/10.1016/j.abd.2021.08.006 PMID: 35379512
- Target product profile for a diagnostic test to confirm leprosy in individuals with clinical signs and symptoms. Geneva: World Health Organization; 2023. Available from: https://www.who.int/publications/i/item/9789240073739 [cited 2023 Jun 23].
- 10. Target product profile for a diagnostic test to detect Mycobacterium leprae infection among asymptomatic household and familial contacts of leprosy patients. Geneva: World Health Organization; 2023. Available from: https:// www.who.int/publications/i/item/9789240074231 [cited 2023 Jun 23].
- 11. Leprosy/Hansen disease: contact tracing and post-exposure prophylaxis. New Delhi: World Health Organization Regional Office for South-East Asia; 2020. Available from: https://iris.who.int/handle/10665/336679 [cited 2023 Jun 23].
- 12. Bakker MI, Hatta M, Kwenang A, Van Mosseveld P, Faber WR, Klatser PR, et al. Risk factors for developing leprosy—a population-based cohort study in Indonesia. Lepr Rev. 2006 Mar;77(1):48-61. doi: http://dx.doi.org/10.47276/ Ir.77.1.48 PMID: 16715690

- 13. Sales AM, Ponce de Leon A, Düppre NC, Hacker MA, Nery JA, Sarno EN, et al. Leprosy among patient contacts: a multilevel study of risk factors. PLoS Negl Trop Dis. 2011 Mar 15;5(3). doi: http://dx.doi.org/10.1371/journal.pntd .0001013 PMID: 21423643
- 14. Richardus JH. Leprosy remains an important public health challenge in India. Indian J Med Res. 2013 May;137(5):878-9.https://www.ncbi.nlm.nih .gov/pubmed/23760371 PMID: 23760371
- 15. Dharmawan Y, Fuady A, Korfage IJ, Richardus JH. Delayed detection of leprosy cases: A systematic review of healthcare-related factors. PLoS Negl Trop Dis. 2022 Sep 6;16(9):e0010756. doi: http://dx.doi.org/10.1371/journal .pntd.0010756 PMID: 36067195
- Saunderson P. WHO global leprosy (Hansen's disease) update, 2022: new paradigm- control to elimination. Lepr Rev. 2023;94(4):262-3. doi: http://dx .doi.org/10.47276/lr.94.4.262
- 17. Gómez L, Rivera A, Vidal Y, Bilbao J, Kasang C, Parisi S, et al. Factors associated with the delay of diagnosis of leprosy in north-eastern Colombia: a quantitative analysis. Trop Med Int Health. 2018 Feb;23(2):193-8. doi: http://dx.doi.org/10.1111/tmi.13023 PMID: 29230912
- 18. Post E, Brandsma W, Wagenaar I, Alam K, Shetty V, Husain S, et al. Delay in leprosy diagnosis – a multi-center, multi-country Asian study. Lepr Rev. 2021;92(1):29-37. doi: http://dx.doi.org/10.47276/lr.92.1.29
- Guidelines for the diagnosis, treatment and prevention of leprosy. New Delhi: World Health Organization Regional Office for South-East Asia; 2018. Available from: https://iris.who.int/handle/10665/274127 [cited 2023 Jun 23].
- Romero CP, Castro R, do Brasil PEA, Pereira DR, Pinheiro RO, Toscano CM, et al. Accuracy of rapid point-of-care serological tests for leprosy diagnosis: a systematic review and meta-analysis. Mem Inst Oswaldo Cruz. 2022 Apr 8;117:e220317. doi: http://dx.doi.org/10.1590/0074-02760220317 PMID: 35416839
- 21. van Hooij A, Tjon Kon Fat EM, de Jong D, Khatun M, Soren S, Chowdhury AS, et al. Prototype multi-biomarker test for point-of-care leprosy diagnostics. iScience. 2020 Dec 29;24(1):102006. doi: http://dx.doi.org/10.1016/j.isci .2020.102006 PMID: 33490914
- 22. Saar M, Beissner M, Gültekin F, Maman I, Herbinger KH, Bretzel G. RLEP LAMP for the laboratory confirmation of leprosy: towards a point-of-care test. BMC Infect Dis. 2021 Nov 25;21(1):1186. doi: http://dx.doi.org/10.1186/ s12879-021-06882-2 PMID: 34823479
- 23. van Hooij A, Tjon Kon Fat EM, van den Eeden SJF, Wilson L, Batista da Silva M, Salgado CG, et al. Field-friendly serological tests for determination of M. leprae-specific antibodies. Sci Rep. 2017 Aug 21;7(1). doi: http://dx.doi.org/ 10.1038/s41598-017-07803-7 PMID: 28827673
- 24. Gurung P, Gomes CM, Vernal S, Leeflang MMG. Diagnostic accuracy of tests for leprosy: a systematic review and meta-analysis. Clin Microbiol Infect. 2019 Nov;25(11):1315-27.doi: http://dx.doi.org/10.1016/j.cmi.2019.05.020 PMID: 31158516

- 25. Pierneef L, Malaviya P, van Hooij A, Sundar S, Singh AK, Kumar R, et al. Fieldfriendly anti-PGL-I serosurvey in children to monitor Mycobacterium leprae transmission in Bihar, India. Front Med (Lausanne). 2023 Sep 27;10:1260375. doi: http://dx.doi.org/10.3389/fmed.2023.1260375 PMID: 37828950
- 26. van Hooij A, Tjon Kon Fat EM, de Jong D, Khatun M, Soren S, Chowdhury AS, et al. Prototype multi-biomarker test for point-of-care leprosy diagnostics. iScience. 2021 Dec 29;24(1). doi: http://dx.doi.org/10.1016/j.isci.2020 .102006 PMID: 33490914
- 27. Tió-Coma M, Kiełbasa SM, van den Eeden SJF, Mei H, Roy JC, Wallinga J, et al. Blood RNA signature RISK4LEP predicts leprosy years before clinical onset. EBioMedicine. 2021 Jun;68. doi: http://dx.doi.org/10.1016/j.ebiom.2021 .103379 PMID: 34090257
- Pierneef L, van Hooij A, Taal A, Rumbaut R, Nobre ML, van Brakel W, et al. Detection of anti-M. leprae antibodies in children in leprosy-endemic areas: a systematic review. PLoS Negl Trop Dis. 2021 Aug 27;15(8. doi: http://dx.doi .org/10.1371/journal.pntd.0009667 PMID: 34449763
- 29. Scollard DM, Adams LB, Gillis TP, Krahenbuhl JL, Truman RW, Williams DL. The continuing challenges of leprosy. Clin Microbiol Rev. 2006 Apr;19(2):338-81. doi: http://dx.doi.org/10.1128/CMR.19.2.338-381.2006 PMID: 16614253
- 30. Singh A, Ambujam S, Kumar NS. No hypopigmented lesion, no nerve thickening, but its leprosy! Indian J Dermatol. 2012 Jan;57(1):73-4. doi: http://dx.doi.org/10.4103/0019-5154.92689 PMID: 22470220
- 31. GenoType LepraeDR VER 1.0. Nehren: Hain Lifescience GmbH; 2023. Available from: https://www.hain-lifescience.de/en/products/microbiology/ mycobacteria/lepra/genotype-lepraedr.html [cited 2023 Jun 23].
- 32. Naaz F, Mohanty PS, Bansal AK, Kumar D, Gupta UD. Challenges beyond elimination in leprosy. Int J Mycobacteriol. 2017 Jul-Sep;6(3):222-8. doi: http://dx.doi.org/10.4103/ijmy.ijmy_70_17 PMID: 28776519
- 33. Souza AA, Ducker C, Argaw D, King JD, Solomon AW, Biamonte MA, et al. Diagnostics and the neglected tropical diseases roadmap: setting the agenda for 2030. Trans R Soc Trop Med Hyg. 2021 Jan 28;115(2):129-35. doi: http://dx.doi.org/10.1093/trstmh/traa118 PMID: 33169166
- 34. Ficalora J, Cohen L, St C. A QFD handbook quality function deployment and six sigma: a QDF handbook. 2nd ed. Boston: Prentice Hall; 2022. Available from: https://www.researchgate.net/profile/Cahyono-St/publication/361910508_A _QFD_Handbook_Quality_Function_Deployment_and_Six_Sigma_Second _Edition/links/62cc515200d0b4511049d481/A-QFD-Handbook-Quality -Function-Deployment-and-Six-Sigma-Second-Edition.pdf
- 35. Juran JM. Juran on quality by design: the new steps for planning quality into goods and services. New York: Free Press; 1992. Available from: https://www.google.co.ke/books/edition/Juran_on_Quality_by_Design/ KPUXbZ2Hw1EC?hl=en&gbpv=0 [cited 2023 Jun 23].
- 36. Kukkaro P, Vedithi SC, Blok DJ, van Brakel WH, Geluk A, Srikantam A, et al. Supplemental information for target product profiles: leprosy diagnostics [online repository]. London: figshare; 2023. doi: http://dx.doi.org/10.6084/ m9.figshare.25199423
- 37. IVD risk-based classification [internet]. Geneva: World Health Organization; 2014. Available from: https://extranet.who.int/prequal/vitro-diagnostics/ ivd-risk-based-classification [cited 2023 Jun 23].

- 38. Neves KVRN, Nobre ML, Machado LMG, Steinmann P, Ignotti E. Misdiagnosis of leprosy in Brazil in the period 2003–2017: spatial pattern and associated factors. Acta Trop. 2021 Mar;215:105791. doi: http://dx.doi.org/10.1016/j .actatropica.2020.105791 PMID: 33310076
- 39. Gass K. Time for a diagnostic sea-change: rethinking neglected tropical disease diagnostics to achieve elimination. PLoS Negl Trop Dis. 2020 Dec 31;14(12). doi: http://dx.doi.org/10.1371/journal.pntd.0008933 PMID: 33382694
- 40. Martinez AN, Britto CF, Nery JA, Sampaio EP, Jardim MR, Sarno EN, et al. Evaluation of real-time and conventional PCR targeting complex 85 genes for detection of Mycobacterium leprae DNA in skin biopsy samples from patients diagnosed with leprosy. J Clin Microbiol. 2006 Sep;44(9):3154-9. doi: http://dx.doi.org/10.1128/JCM.02250-05 PMID: 16954241
- 41. Jiang H, Shi Y, Chokkakula S, Zhang W, Long S, Wang Z, et al. Utility of multitarget nested PCR and ELISPOT assays for the detection of paucibacillary leprosy: a possible conclusion of clinical laboratory misdiagnosis. Front Cell Infect Microbiol. 2022 Apr 11;12. doi: http://dx.doi.org/10.3389/fcimb.2022 .814413 PMID: 35480232
- 42. Manta FSN, Barbieri RR, Moreira SJM, Santos PTS, Nery JAC, Duppre NC, et al. Quantitative PCR for leprosy diagnosis and monitoring in household contacts: a follow-up study, 2011-2018. Sci Rep. 2019 Nov 13;9(1). doi: http://dx.doi.org/10.1038/s41598-019-52640-5 PMID: 31723144
- 43. Chaitanya VS, Cuello L, Das M, Sudharsan A, Ganesan P, Kanmani K, et al. Analysis of a novel multiplex polymerase chain reaction assay as a sensitive tool for the diagnosis of indeterminate and tuberculoid forms of leprosy. Int J Mycobacteriol. 2017 Jan-Mar;6(1):1–8. doi: http://dx.doi.org/10.4103/2212 -5531.201885 PMID: 28317797
- 44. Barth-Jaeggi T, Steinmann P, Mieras L, van Brakel W, Richardus JH, Tiwari A, et al.; LPEP study group. Leprosy post-exposure prophylaxis (LPEP) programme: study protocol for evaluating the feasibility and impact on case detection rates of contact tracing and single dose rifampicin. BMJ Open. 2016 Nov 17;6(11):e013633. doi: http://dx.doi.org/10.1136/bmjopen-2016 -013633 PMID: 27856484
- 45. Moet FJ, Pahan D, Oskam L, Richardus JH; COLEP Study Group. Effectiveness of single dose rifampicin in preventing leprosy in close contacts of patients with newly diagnosed leprosy: cluster randomised controlled trial. BMJ. 2008 Apr 5;336(7647):761-4. doi: http://dx.doi.org/10.1136/bmj.39500 .885752.BE PMID: 18332051
- Ortuno-Gutierrez N, Younoussa A, Randrianantoandro A, Braet S, Cauchoix B, Ramboarina S, et al. Protocol, rationale and design of PEOPLE (post exposure prophylaxis for leprosy in the Comoros and Madagascar): a cluster randomized trial on effectiveness of different modalities of implementation of post-exposure prophylaxis of leprosy contacts. BMC Infect Dis. 2019 Dec 5;19(1):1033. doi: http://dx.doi.org/10.1186/s12879-019-4649-0 PMID:
- 47. Younoussa A, Samidine SN, Bergeman AT, Piubello A, Attoumani N, Grillone SH, et al. Protocol, rationale and design of BE-PEOPLE (bedaquiline enhanced exposure prophylaxis for leprosy in the Comoros): a cluster randomized trial on effectiveness of rifampicin and bedaquiline as post-exposure prophylaxis of leprosy contacts. BMC Infect Dis. 2023 May 9;23(1):310. doi: http://dx.doi.org/10.1186/s12879-023-08290-0 PMID: 37161571