

This is the peer reviewed version of the following article:

Pineda-Tenor, Daniel; Berenguer, Juan; García-Broncano, Pilar; Jiménez-Sousa, María A; Fernández-Rodríguez, Amanda; Diez, Cristina; García-Álvarez, Mónica; Carrero, Ana; Catalán, Pilar; Aldámiz-Echevarria, Teresa; Resino, Salvador. **Association of adiponectin (ADIPOQ) rs2241766 polymorphism and dyslipidemia in HIV/HCV-coinfected patients.** Eur J Clin Invest. 2014 May;44(5):453-62.

which has been published in final form at:

<https://doi.org/10.1111/eci.12250>

Title page

Type of manuscript: Article

Title: Association of adiponectin (*ADIPOQ*) rs2241766 polymorphism and dyslipidemia in HIV/HCV coinfecting patients

Short title: *ADIPOQ* polymorphism and dyslipidemia

Authors: Daniel PINEDA-TENOR ¹, Ph.D.; Juan BERENGUER ^{2,3}, M.D., Ph.D.; Pilar GARCÍA-BRONCANO ¹, Bs.C.; María A JIMÉNEZ-SOUSA ¹, Ph.D.; Amanda FERNÁNDEZ-RODRÍGUEZ ¹, Ph.D.; Cristina DIEZ ^{2,3}, M.D.; Mónica GARCÍA-ÁLVAREZ ¹, Ph.D.; Ana CARRERO ^{2,3}, M.D.; Pilar CATALÁN ⁴, Ph.D.; Teresa ALDÁMIZ-ECHEVARRIA ^{2,3}, M.D.; Salvador RESINO ^{1(*)}, Ph.D.

(*) Corresponding author.

Current affiliations: (1) Viral Infection and Immunity Unit, National Centre of Microbiology. Instituto de Salud Carlos III, Majadahonda, Madrid, Spain. (2) Infectious Diseases-HIV Unit; Hospital General Universitario "Gregorio Marañón", Madrid, Spain. (3) Instituto de Investigación Sanitaria Gregorio Marañón (IISGM), (4) Microbiology Department, Hospital General Universitario "Gregorio Marañón," Madrid, Spain.

Correspondence and requests for reprints: Salvador Resino; Centro Nacional de Microbiología, Instituto de Salud Carlos III (Campus Majadahonda); Carretera Majadahonda- Pozuelo, Km 2.2; 28220 Majadahonda (Madrid); Telf.: +34 918 223 266; Fax: +34 918 223 269; e-mail: sresino@isciii.es

ABSTRACT

Background: The adiponectin (*ADIPOQ*) rs2241766 polymorphism is related to metabolic abnormalities. The aim of this study was to evaluate the association of the *ADIPOQ* rs2241766 polymorphism with serum dyslipidemia and insulin resistance (IR) in human immunodeficiency virus (HIV)/hepatitis C virus (HCV) coinfecting patients.

Methods: We carried out a cross-sectional study on 262 patients. *ADIPOQ* rs2241766 polymorphisms were genotyped by GoldenGate® assay. Generalized Linear Models (GLM) were used to compare continuous outcome variables (total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), non-HDL-C, and homeostatic model assessment (HOMA)); and categorical outcome variables (TC≥200 mg/dL, TG≥170 mg/dL, LDL-C≥100 mg/dL, HDL-C≤35 mg/dL, non-HDL-C≥120 mg/dL, and HOMA≥3.8) according to *ADIPOQ* genotype under a dominant inheritance model.

Results: Patients with the rs2241766 GG/GT genotype had significantly lower serum TC levels (p=0.038) and percentages of TC≥200 mg/dL (p=0.022) than rs2241766 TT carriers. When adjusted GLM was performed, rs2241766 GG/GT was associated with low serum TC levels (Arithmetic mean ratio (AMR)=0.92 ((95%CI=0.85; 0.99) p=0.024) and low likelihood of TC≥200 mg/dL (Odds ratio (OR)=0.32 ((95%CI=0.11; 0.88) p=0.027). When stratifying by steatosis, no significant values were found for patients without steatosis. However, for patients with steatosis, rs2241766 GG/GT genotypes were related to low TC serum values of TC (AMR= 0.89; p= 0.027), LDL-C (AMR= 0.85; p= 0.039), and non-HDL-C (AMR= 0.86; p= 0.015). No significant associations were found between rs2241766 and HOMA values.

Conclusions: The presence of the *ADIPOQ* rs2241766 G allele (GG/GT genotype) was associated with a protective effect against dyslipidemia, primarily in HIV/HCV coinfecting patients with steatosis.

Key words: Adiponectin; chronic hepatitis C; AIDS; SNPs; metabolic disturbance; dyslipidemia.

BACKGROUND

In human immunodeficiency virus (HIV) infected patients, combined antiretroviral therapy (cART) has decreased the rates of morbidity and mortality. However, cART has been linked to metabolic disturbances, such as insulin resistance (IR) and dyslipidemia (reductions in total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), and low-density lipoprotein cholesterol (LDL-C)) [1], which facilitate the possible emergence of other previously hidden comorbidities such as metabolic syndrome, type 2 diabetes mellitus (T2DM) and cardiovascular disease [2, 3]. Moreover, hepatitis C virus (HCV) infection has been associated with steatosis and metabolic abnormalities such as IR, T2DM, and dyslipidemia in HCV monoinfected patients [4]. These metabolic disturbances have also been associated with an increased risk of cardiovascular disease [4-6].

Chronic hepatitis C (CHC) has become a major cause of morbidity and mortality in HIV/HCV coinfecting patients [7, 8]. In fact, HIV/HCV coinfection in the cART era has been associated with a significantly increased risk of cardiovascular disease among HIV-infected patients [9]. Moreover, the presence of dyslipidemia in these patients is not well established because HCV coinfection appears to reduce the lipid build-up associated with cART during HIV infection [10]. In any case, HIV/HCV coinfection appears to exacerbate metabolic disturbance and induce prothrombotic changes in patients [10].

Adipose tissue has been recognized as an active secreting organ that releases a variety of proteins, collectively named adipokines, which may be directly involved in metabolic fluxes to the amount of stored energy [11]. The most abundant adipokine is adiponectin (ADIPOQ), which is a protein that plays a critical role in lipid metabolism and glucose homeostasis [12, 13]. The activity of adiponectin in glucose and lipid metabolism confers protection against obesity and metabolic syndrome traits [14]. In CHC, ADIPOQ exhibits a hepatoprotective and antifibrogenic effect in cases of liver injury, as well as protecting against liver steatosis [15, 16]. Thus, ADIPOQ attenuates inflammatory activity [15], improves insulin sensitivity [17], counteracts ectopic fat deposition and visceral obesity [18], and has a protective effect against hepatocellular carcinoma [19].

The adiponectin gene (*ADIPOQ*) is located on chromosome 3q27, spanning a 16 kb region, and has more than 160 genetic variants [20]. Several single nucleotide polymorphisms (SNPs) of *ADIPOQ* have been associated with metabolic disturbance and cardiovascular disease [21-23]. The rs2241766 (+45T>G) mutation is one of the most studied, but contradictory results have been published about the association of the *ADIPOQ* rs2241766 polymorphism with dyslipidemia and IR in the general population [22, 23].

The aim of our study was to evaluate the association of the *ADIPOQ* rs2241766 polymorphism with the lipid profiles [TC, HDL-C, LDL-C, non-HDL-C, and triglycerides (TG)] and IR in white European HIV/HCV coinfecting patients.

PATIENTS AND METHODS

Study design

We carried out a cross-sectional study on 262 non-diabetic HIV/HCV coinfecting patients at Hospital General Universitario Gregorio Marañón (Madrid, Spain) between September 2000 and July 2009. The study was approved by the Institutional Review Board and the Research Ethic Committee of the Instituto de Salud Carlos III (ISCIII). This study was conducted in accordance with the Declaration of Helsinki and patients gave their written consent. Study reporting conforms to the STROBE statement along with references to STROBE and the broader EQUATOR guidelines [24].

All subjects included in our study were HCV treatment-naïve patients who were potential candidates for HCV therapy and, in most cases, underwent a liver biopsy. The Inclusion criteria for the study were: detectable HCV-RNA by polymerase chain reaction, negative hepatitis B surface antigen, availability of a DNA sample, no clinical evidence of hepatic decompensation, no diabetes mellitus, and successful cART or no need for cART. Patients with active opportunistic infections, active drug and/or alcohol addiction, and other concomitant diseases or conditions, such as nephropathies, autoimmune diseases, hemochromatosis, primary biliary cirrhosis, Wilson's disease, a1-antitrypsin deficiency and neoplasia, were excluded.

Of the 495 HIV/HCV coinfecting patients who met the criteria described above, 293 had a DNA sample available for genotyping. Additionally, 13 patients were excluded due to genotyping problems, and 18 had to be excluded because of no homeostatic model assessment (HOMA) or lipid data. The final study sample consisted of 262 HIV/HCV coinfecting patients, 212 of which had a liver biopsy performed. All patients were European whites.

Clinical and laboratory data

The following information was obtained from medical records when HCV therapy was started and/or liver biopsy was performed: age, gender, HIV transmission category, weight, height, nadir CD4+ T cell count, antiretroviral therapy, HCV genotype, CD4+ T-cell count, plasma HIV viral load (HIV-RNA), plasma HCV viral load (HCV-RNA), and biochemical liver panel tests. Body mass index (BMI) was calculated as the weight in kilograms divided by the square of the height in meters.

The biochemistry panel was measured using an autoanalyzer Hitachi 912 (Boehringer Mannheim, Germany), while patients were fasting. We collected data of TC, HDL-C, and TG. The LDL-C was calculated by Friedewald estimation ($LDL-C = TC - HDL-C - (TG/5)$) [25], and non-HDL-C was calculated as TC minus HDL-C [26].

The degree of IR was estimated for each patient using HOMA described by Matthews et al [27]: fasting plasma glucose (mmol/l) times fasting serum insulin (mU/l) divided by 22.5.

Liver biopsy

Liver biopsies were performed on an outpatient basis following the recommendations of the Patient Care Committee of the American Gastroenterological Association [28]. Liver fibrosis was estimated according to METAVIR score [29]. Fibrosis was scored as follows: F0, no fibrosis; F1, portal fibrosis; F2, periportal fibrosis or rare portal-portal septa; F3, fibrous septa with architectural distortion; no obvious cirrhosis (bridging fibrosis); and F4, definite cirrhosis. Liver steatosis was evaluated according to the existence of hepatocytes containing visible macrovesicular fat droplets. We considered hepatic steatosis to be clinically significant when fatty hepatocytes exceeded 10% of the hepatic parenchyma.

Genotyping of *ADIPOQ* polymorphisms

Genomic DNA was extracted from peripheral blood with Qiagen columns (QIAamp DNA Blood Midi/Maxi; Qiagen, Hilden, Germany). A SNP within the *ADIPOQ* gene (rs2241766 (+45T>G)) was genotyped at the Spanish National Genotyping Center (CeGen). Genotyping was performed by using GoldenGate® assay with VeraCode® Technology (Illumina Inc. San Diego, CA, USA).

Outcome variables

Lipid disturbance: a) serum concentration of TC, LDL-C, HDL-C, non-HDL-C, and TG; b) thresholds of lipid profile: TC \geq 200 mg/dL, LDL-C \geq 100 mg/dL, HDL-C $<$ 35 mg/dL, non-HDL-C \geq 120 mg/dL, and TG \geq 170 mg/dL [26, 30].

Insulin resistance: a) HOMA continuous values; b) HOMA \geq 3.8 [29].

Statistics

All statistical tests were performed with the Statistical Package for the Social Sciences (SPSS) 19.0 software (IBM Corp., Chicago, USA). Graphics were generated by GraphPad PRISM 6.01 (GraphPad Software INC, La Jolla, USA). All p-values were two-tailed and statistical significance was defined as $p < 0.05$. Continuous variables were expressed as median (interquartile range) and categorical variables as percentage (absolute frequency).

For the description of the study population, p-values were estimated with a nonparametric test: Mann-Whitney U test was used for continuous variables and Chi-square for categorical variables. Hardy-Weinberg equilibrium (HWE) was assessed by a Chi-square test, considering equilibrium when $p > 0.05$.

For the genetic association study, univariate and multivariate Generalized Linear Models (GLM) were used to compare the outcome variables (lipid profile and IR) according to *ADIPOQ* polymorphism. We analyzed the data according to several models, including dominant, recessive and additive. The analysis was carried out according to a dominant genetic model for G allele (GG/GT versus TT), which was the model that best fit our data according to the statistical power to detect significant associations.

On the one hand, a GLM with a gamma distribution (log-link) was used to investigate the association between *ADIPOQ* polymorphisms and continuous outcome variables. This test gives the differences between groups and the arithmetic mean ratio (AMR) in continuous outcome variables between groups. On the other hand, a GLM with binomial distribution (logit-link) was used to investigate the association of *ADIPOQ* polymorphisms with categorical outcome variables. This test gives the differences between groups and the odds ratio (OR) for categorical outcome variables. GLM tests were adjusted by the most important clinical and epidemiological characteristics. In each adjusted regression analysis, we included a SNP and the most significant covariables (backward criterion with a p-value for exit of 0.20). The covariables used were gender, age, BMI, nadir CD4+ T-cells, undetectable HIV viral load (<50 copies/mL), time on cART, cART with protease inhibitor, specific antiretroviral drugs (saquinavir, efavirenz, ritonavir, etc.), HCV genotype (GT1/4 vs. GT2/3), and HCV viral load $\geq 500,000$ IU/ml. The goodness of fit of GLM tests against other statistical tests were evaluated by Akaike information criterion (AIC) value and Bayesian information criterion (BIC) [31].

RESULTS

Characteristics of patients

Table 1 shows the main characteristics of the 262 HIV/HCV coinfecting patients participating in the study. No significant differences were found when stratifying by rs2241766 genotypes (GG/GT versus TT).

Table 1. Epidemiological and demographic characteristics of HIV/HCV coinfecting patients. Categorical variables are expressed in absolute numbers (percentage of patients). Continuous variables are expressed in median (interquartile range). P-values were estimated with a nonparametric Mann-Whitney U test for continuous variables and Chi-square for categorical variables.

Characteristics	All Patients	rs2241766		p-value
		GG/GT	TT	
N (%)	262 (100%)	81 (30.9%)	181 (69.1%)	
Male, n (%)	197 (75.2%)	61 (75.3%)	136 (75.1%)	0.976
Age, years	40.8 (IQR=6.9)	40.9 (IQR=6.6)	40.9 (IQR=7)	0.213
BMI, kg/m²	22.5 (IQR=3.7)	22.4 (IQR=3.6)	22.5 (IQR=3.9)	0.936
HIV acquired by IVDU, n (%)	224 (85.5%)	73 (90.1%)	151 (83.4%)	0.565
Prior AIDS, n (%)	76 (29%)	23 (28.4%)	53 (29.3%)	0.884
Time on cART, years	4.8 (IQR=5)	4.4 (IQR=5.5)	5.0 (IQR=4.9)	0.760
Current cART protocols, n (%)				
Any NRTIs + any PI	64 (24.2%)	20 (24.7%)	44 (24.3%)	0.947
Any NRTIs + PI + NNRTI	2 (0.8%)	0 (0%)	2 (1.1%)	0.342
Any NRTIs + any NNRTI	135 (51.1%)	36 (44.4%)	99 (54.7%)	0.125
Only NRTIs	19 (7.3%)	5 (6.2%)	14 (7.7%)	0.652
Specific antiretroviral drugs, n (%)				
Thymidine analogues + ddl	156 (59.5%)	42 (51.9%)	114 (63%)	0.090
Efavirenz	79 (30.2%)	19 (23.5%)	60 (33.1%)	0.114
Saquinavir	3 (1.1%)	2 (2.5%)	1 (0.6%)	0.178
Fosamprenavir	6 (2.3%)	2 (2.5%)	4 (2.2%)	0.897
Ritonavir	16 (6.1%)	6 (7.4%)	10 (5.5%)	0.556
HIV markers				
Nadir CD4+, cells/ μ L	206 (IQR=222.8)	210 (IQR=226.5)	203 (IQR=229)	0.909
Nadir CD4+ <200 cells/ μ L, n (%)	127 (48.5%)	39 (48.1%)	88 (48.6%)	0.944
CD4+ T, cells/ μ L	462 (IQR=332.3)	483 (IQR=300)	456 (IQR=350)	0.540
CD4+ \geq 500 cells/ μ L, n (%)	113 (43.3%)	38 (46.9%)	75 (41.7%)	0.429

HIV-RNA < 50 copies/mL, n (%)	200 (76.6%)	56 (69.1%)	144 (80%)	0.055
HCV markers, n (%)				
HCV-GT 1	143 (56.5%)	42 (55.3%)	101 (57.1%)	0.791
HCV-GT 2	5 (2%)	1 (1.3%)	4 (2.2%)	0.625
HCV-GT 3	65 (25.6%)	17 (22.4%)	48 (27%)	0.442
HCV-GT 4	41 (16.1%)	16 (21.1%)	25 (14%)	0.165
HCV-RNA ≥ 500.000 IU/ml	187 (74.8%)	57 (76%)	130 (74.3%)	0.775
Metavir, n (%)				
Liver biopsy patients	210 (80.2%)	66 (81.5%)	144 (79.6%)	0.620
Significant fibrosis (F≥2)	103 (49%)	28 (42.4%)	75 (52.1%)	0.194
Moderate or severe activity (A≥2)	108 (52.2%)	29 (43.9%)	79 (56%)	0.105
Steatosis	116 (57.4%)	36 (58.1%)	80 (57.1%)	0.903

Abbreviations: AIDS, acquired immunodeficiency syndrome; BMI, body mass index; cART, combined antiretroviral therapy; ddl, didanosine; GT, Genotype; HCV, Hepatitis C virus; HCV-RNA, HCV plasma viral load; HDL-C, High-density lipoprotein; HIV, human immunodeficiency virus; HIV-RNA, HIV plasma viral load; HOMA, homeostatic model assessment method; IQR, interquartile range; IVDU, intravenous drug users; LDL-C, low-density lipoprotein; NNRTI, no nucleoside analog reverse-transcriptase inhibitors; NRTI, nucleoside analog reverse-transcriptase inhibitors; PI, protease inhibitors.

Frequencies of the *ADIPOQ* polymorphism

Allele frequencies for the *ADIPOQ* rs2241766 polymorphism were 0.17 (G allele) and 0.83 (T allele). Genotype frequencies for the *ADIPOQ* rs2241766 polymorphism were 0.03 (GG), 0.28 (GT), and 0.69 (TT). These frequencies in our dataset were in accordance with data listed on the NCBI SNP database (http://www.ncbi.nlm.nih.gov/projects/SNP/snp_ref.cgi?rs=2241766). The rs2241766 SNP had fulfilled the minimum allele frequency (MAF) >0.05 for all samples and displayed less than 5% of missing values. Furthermore, rs2241766 was in HWE ($p= 0.940$).

Lipid profiles and the *ADIPOQ* polymorphism

Patients with rs2241766 GG/GT genotype had significantly lower serum TC levels ($p= 0.038$; **Figure 1A**) and lower percentages of TC values ≥ 200 mg/dL ($p= 0.022$; **Figure 1B**) than patients with rs2241766 TT. When adjusted GLM was performed, rs2241766 GG/GT genotype was associated with low serum TC levels (AMR= 0.92; $p= 0.024$; **Figure 2A**) and low likelihood of TC values ≥ 200 mg/dL (OR= 0.32; $p= 0.027$; **Figure 2B**), supporting the protective effect of the G allele. However, although rs2241766 GG/GT carriers seem to have reduced values of LDL-C, HDL-C, non-HDL-C, and triglycerides, no significant differences were observed (**Figure 1 & Figure 2**).

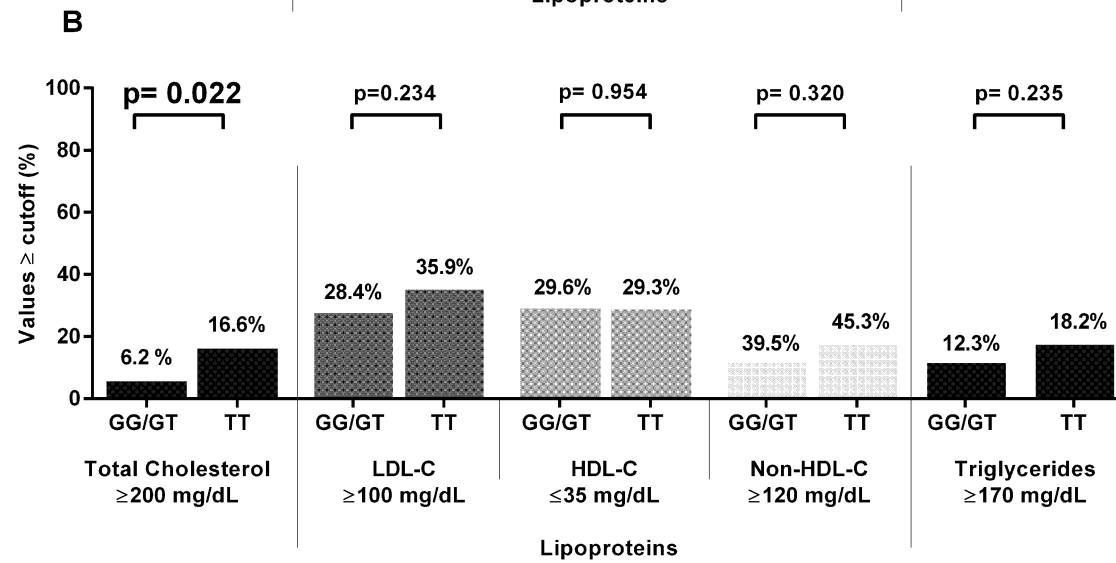
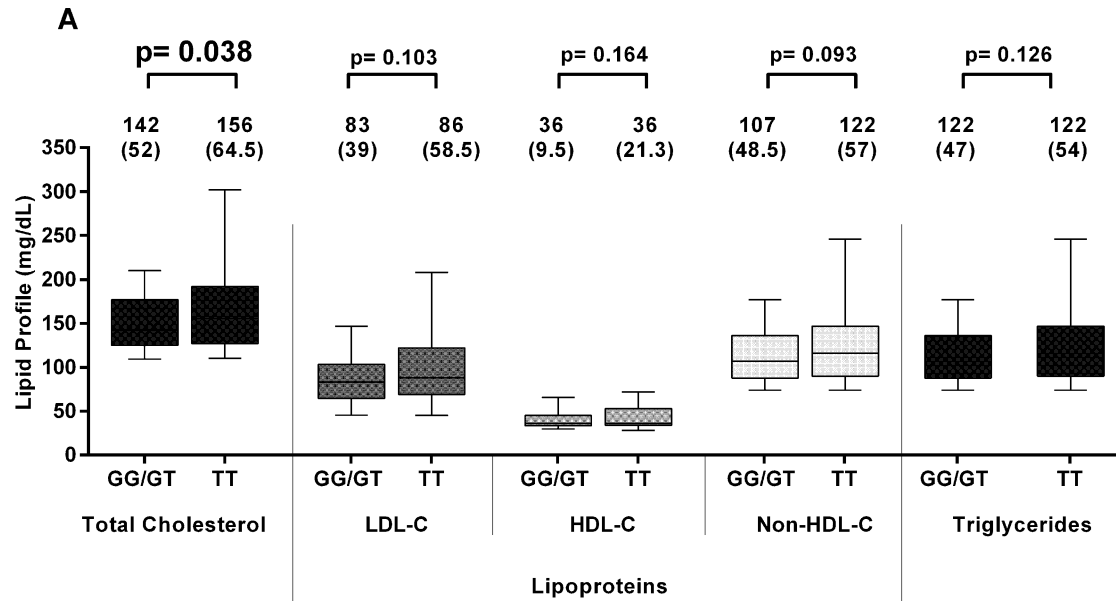


Figure 1. Distribution of lipid profile values according to *ADIPOQ* rs2241766 polymorphism in HIV/HCV coinfecting patients. The median (interquartile range) and p-values (unadjusted GLM with gamma distribution) are shown for continuous variables (**A**). The percentage of patients and p-values (unadjusted GLM with binomial distribution) are shown for thresholds (**B**).

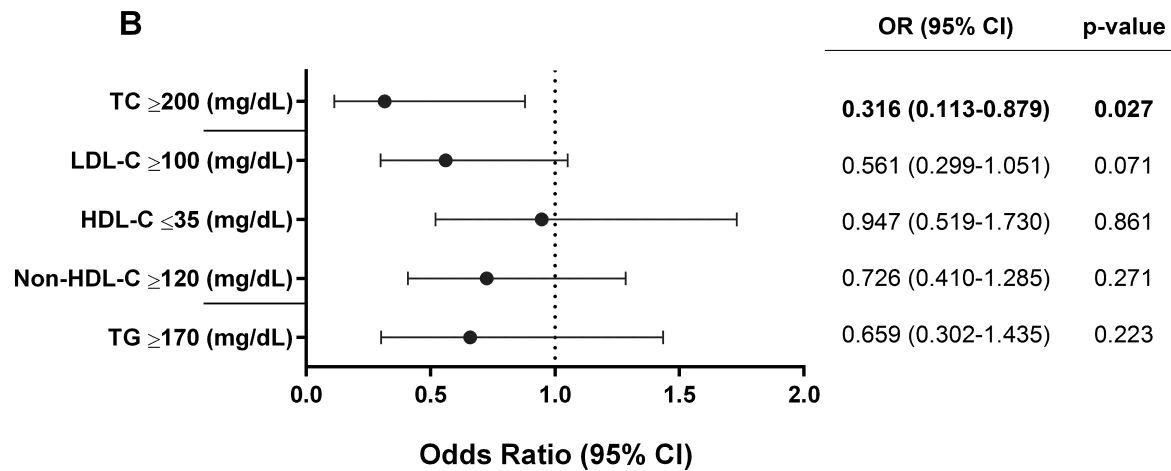
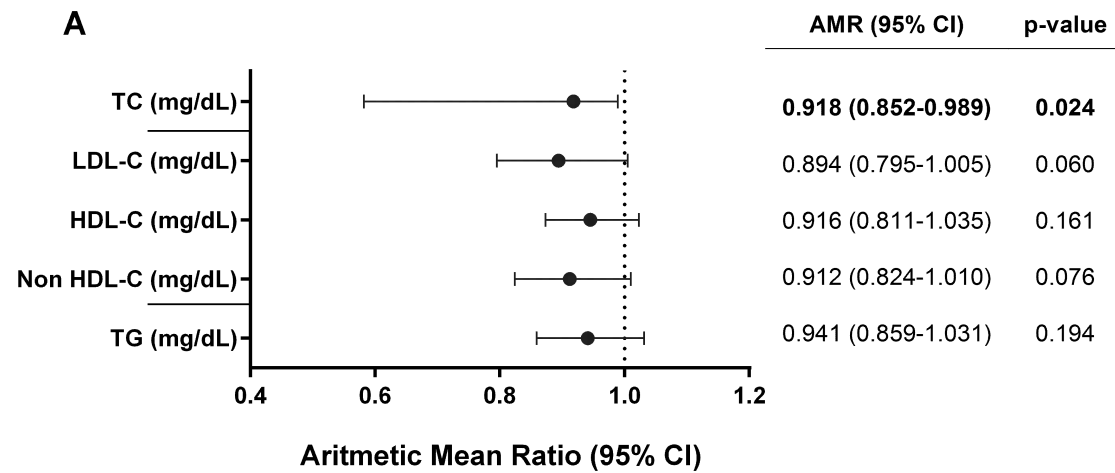


Figure 2. Association of *ADIPOQ* rs2241766 polymorphisms with serum lipid values (A) and dyslipidemia thresholds (B) in HIV/HCV coinfecting patients. Multivariate GLM were adjusted by several epidemiological and clinical factors [age, gender, BMI, nadir CD4+, HIV plasma viral load, HCV plasma viral load, time on cART and specific antiretroviral drugs].

Additionally, we analyzed the association between the *ADIPOQ* rs2241766 polymorphism and serum lipid values stratified by steatosis (**Figure 3**). No significant values were found for patients without steatosis (**Figure 3A**). However, for patients with steatosis (**Figure 3B**), rs2241766 GG/GT carriers had lower serum values of TC ($p=0.039$), LDL-C ($p=0.036$), and non-HDL-C ($p=0.021$) than rs2241766 TT carriers. When adjusted GLM was performed, the rs2241766 GG/GT genotype was related with low serum values of TC (AMR= 0.89; $p=0.027$), LDL-C (AMR= 0.85; $p=0.039$), and non-HDL-C (AMR= 0.86; $p=0.015$) (**Figure 4A**).

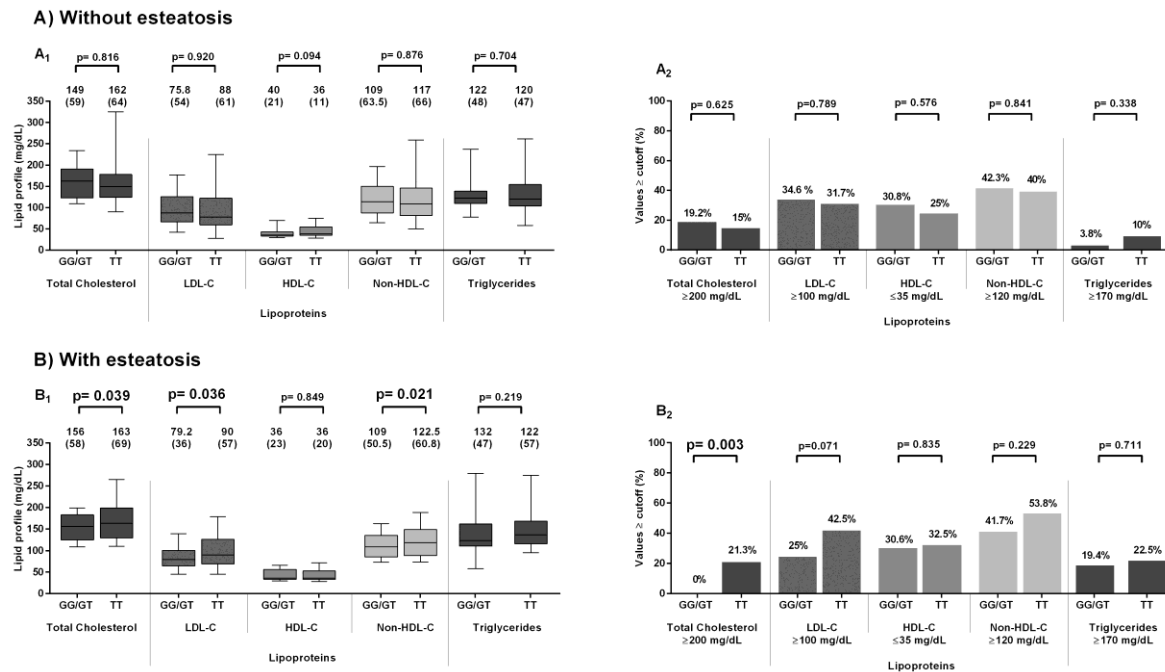


Figure 3. Distribution of lipid profile values according to *ADIPOQ* rs2241766 polymorphism in HIV/HCV coinfecting patients stratified by steatosis. The median (interquartile range) and p-values (unadjusted GLM with gamma distribution) are shown for continuous variables (**A₁**, **B₁**). The percentage of patients and p-values (unadjusted GLM with binomial distribution) are shown for thresholds (**A₂**, **B₂**).

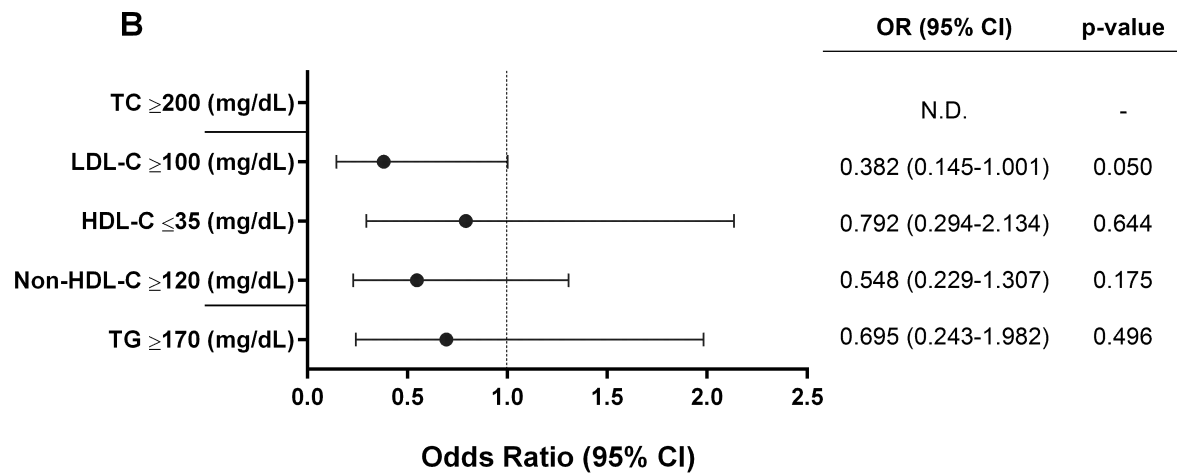
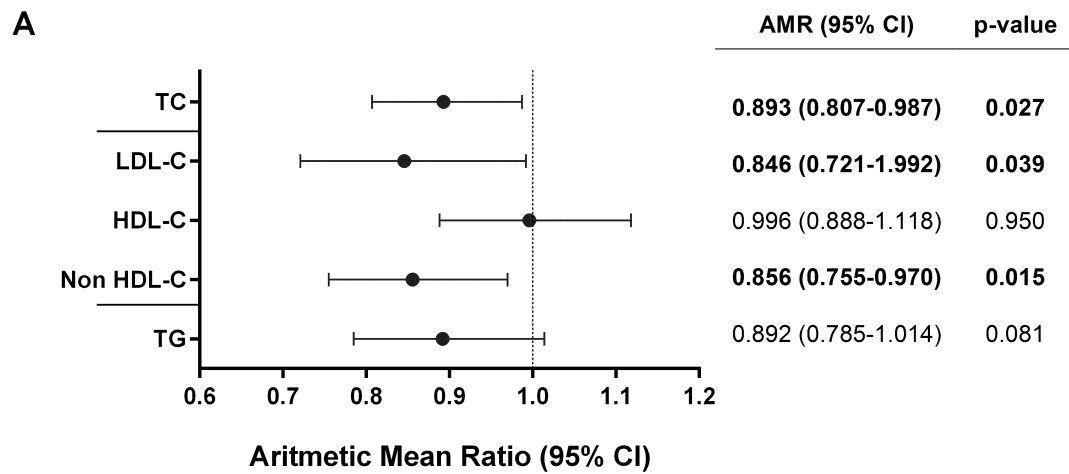


Figure 4. Association of *ADIPOQ* rs2241766 polymorphisms with serum lipid values (**A**) and dyslipidemia thresholds (**B**) by multivariate GLM tests in HIV/HCV coinfectd patients with steatosis. These tests were adjusted by several epidemiological and

clinical factors [age, gender, BMI, nadir CD4+, HIV plasma viral load, HCV plasma viral load, time on cART and specific antiretroviral drugs].

Insulin resistance and the *ADIPOQ* polymorphism

When analyzing the association of the *ADIPOQ* rs2241766 polymorphism with HOMA values and the percentage of patients with HOMA ≥ 3.8 , no significant differences were found for all patients, nor for patients stratified by significant liver fibrosis (**Supplemental Figure 1**). Additionally, when adjusted GLM was performed, no significant association was found (**Supplemental Figure 2**).

DISCUSSION

To the best of our knowledge, there is no data about the relationship of the *ADIPOQ* rs2241766 polymorphism with metabolic disturbances in HIV/HCV coinfecting patients. In this study, we found that the rs2241766 GG/GT genotype was significantly associated with lower cholesterol levels (TC, LDL-C, and non-HDL-C) mainly in patients with liver steatosis.

Adiponectin is a potent modulator of lipid metabolism and an indicator of metabolic disorders [11]. Thus, serum adiponectin levels have been found to be inversely associated with components of metabolic syndrome, such as obesity, IR and T2DM [11]. The serum adiponectin concentration has a strong genetic component, with heritability estimated at 88% [32], but the mechanism of how the rs2241766 (+45T>G) *ADIPOQ* polymorphism is related to metabolic disturbance remains to be determined. The *ADIPOQ* rs2241766 polymorphism is a synonymous mutation (GGT→GGG, Gly→Gly), and the way in which this variation influences metabolic phenotypes remains to be elucidated. The G allele at the rs2241766 polymorphism has been associated with higher serum adiponectin concentrations and higher adiponectin mRNA expression in adipose tissue [33, 34]. However, associations between the *ADIPOQ* rs2241766 polymorphism and adiponectin concentration in population studies have not always been confirmed [35-37]. In our study, we did not find any association between *ADIPOQ* genotypes and values of serum adiponectin (*data not shown*). This lack of significance might be due to the limited number of patients used in the analysis (only 92 samples were available), or also due to the possible distortive effect of direct and indirect factors related to HIV and HCV infection [38].

Dyslipidemia and IR are two complex metabolic disturbances in which genetic and environmental factors interact to produce homeostatic abnormalities [39-42]. The *ADIPOQ* rs2241766 polymorphism has been extensively studied among HIV and HCV seronegative subjects in relation to the development of T2DM, blood lipids and blood pressure, but the influence of the *ADIPOQ* rs2241766 polymorphism on metabolism disturbance is not clear [22, 23]. Thus, the rs2241766 G allele has been associated with obesity, adverse lipid profiles, T2DM, hyperglycemia and IR [43-49], while other studies proposed a similar association for the rs2241766 T allele [50, 51] or a lack of associations between rs2241766 and alterations in the T2DM status, obesity, dyslipidemia or hypertension [36, 52]. In our study, the rs2241766 GG/GT genotype had a protective effect against dyslipidemia, with lower cholesterol levels mainly in HIV/HCV coinfecting patients with steatosis. Additionally, no significant association between rs2241766 and HOMA values were found in our study.

Our findings are not consistent with a recent meta-analysis, which shows that the G allele of the *ADIPOQ* rs2241766 polymorphism is associated with a higher risk of cardiovascular disease [21]. However, other recent meta-analysis has shown that the presence of +45T>G (rs2241766) does not appear to influence the development of type 2 diabetes [22]. These discrepancies could be due to the characteristics of the study population, since HIV/HCV coinfecting patients had an intrinsic metabolic deregulation due to the HIV and HCV coinfection [38]. In addition, cART is often associated with severe metabolic disorders, such as IR and hyperlipidemia [38]. Liver steatosis in CHC has been associated with metabolic disturbance such as IR and the metabolic syndrome [4]; reductions in TC as well as HDL-C and LDL-C are observed in HCV-infected patients

compared to matched controls [4]. Thus, these observations highlight that HIV/HCV coinfecting patients form a very unique population, where perhaps our findings need not coincide with those of the general population.

Some aspects must be taken into account for a correct interpretation of our data: a) this is a cross-sectional study with a relatively low sample size, however our population provides adequate statistical power to detect the significant associations that are shown in this study (**Supplemental Table 1**); b) it is uncertain whether our results would be observed in HCV mono-infected patients; c) the patients selected for our study were patients who met a set of criteria for starting HCV treatment (for example little alcohol abuse, high CD4 cell counts, controlled HIV replication, and good treatment adherence), and it is possible that this may have introduced a selection bias; d) this study was carried out entirely in white Europeans, therefore as the frequency of these alleles differs among different ethnicities, it would be necessary to perform an independent replication of this study for different ethnic groups; e) the rs2241766 polymorphism is located in a linkage disequilibrium (LD) block spanning from the intron 1 to exon 3 of the adiponectin gene [20], and it is possible that other genetic variants in the LD with the rs2241766 polymorphism are causal variants for the associations.

In conclusion, the presence of the rs2241766 G allele (GG and GT genotypes) was associated with a protective effect against dyslipidemia, mainly in HIV/HCV coinfecting patients with steatosis. Thus, our results suggest that rs2241766 may play a significant role in metabolic disorders in HIV/HCV coinfecting patients. Further studies in larger populations will be required to determine whether the *ADIPOQ* polymorphisms play a pivotal role in energy metabolism of HIV/HCV coinfecting patients.

ACKNOWLEDGEMENTS, CONFLICT OF INTEREST, AND FUNDING

ACKNOWLEDGEMENTS

The authors wish to thank the Spanish National Genotyping Center (CeGen) for providing the SNP genotyping services (<http://www.cegen.org>).

COMPETING INTERESTS

The authors do not have any commercial or other association that might pose a conflict of interest.

Funding/Support: This work has been supported by grants given by Fondo de Investigación de Sanidad en España (FIS) [Spanish Health Funds for Research] [grant numbers PI08/0738, PI11/00245; PI08/0928, and PI11/01556], Red Española de Investigación en SIDA (RIS) [AIDS Research Network] [grant numbers RD12/0017/0024 and RD12/0017/0004] and “Fundación para la Investigación y la Prevención del Sida en España” (FIPSE) [grant number 361020/10].

DPT, PGB, MAJS, AFR, and MGA are supported by “Instituto de Salud Carlos III” [grant numbers CM12/00043, FI12/00036, CM10/00105, PI11/00245, CD12/00442, respectively].

JB is an investigator from the Programa de Intensificación de la Actividad Investigadora en el Sistema Nacional de Salud (I3SNS) .

AUTHORS' CONTRIBUTIONS:

JB and SR participated in the study concept and design. DPT and SR performed all statistical analysis, interpretation of the data and wrote the manuscript. JB, CD, AC, PC and TA participated in patient selection, collection of samples and acquisition of data. DPT, PGC, MAJS, AFR, and MGA participated in sample preparation, DNA isolation and genotyping pre-procedure, and contributed with critical revision of the manuscript. SR supervised the study.

All authors revised the manuscript from a draft by SR.

REFERENCES

1. Samaras K. Metabolic consequences and therapeutic options in highly active antiretroviral therapy in human immunodeficiency virus-1 infection. *J Antimicrob Chemother* 2008,**61**:238-245.
2. Calza L, Manfredi R, Pocaterra D, Chiodo F. Risk of premature atherosclerosis and ischemic heart disease associated with HIV infection and antiretroviral therapy. *J Infect* 2008,**57**:16-32.
3. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. *J Clin Endocrinol Metab* 2007,**92**:2506-2512.
4. Negro F, Sanyal AJ. Hepatitis C virus, steatosis and lipid abnormalities: clinical and pathogenic data. *Liver Int* 2009,**29 Suppl 2**:26-37.
5. Younossi ZM, McCullough AJ. Metabolic syndrome, non-alcoholic fatty liver disease and hepatitis C virus: impact on disease progression and treatment response. *Liver Int* 2009,**29 Suppl 2**:3-12.
6. Younossi ZM, Stepanova M, Nader F, Younossi Z, Elsheikh E. Associations of chronic hepatitis C with metabolic and cardiac outcomes. *Aliment Pharmacol Ther* 2013,**37**:647-652.
7. Lopez-Dieguez M, Montes ML, Pascual-Pareja JF, Quereda C, Von Wichmann MA, Berenguer J, *et al*. The natural history of liver cirrhosis in HIV-hepatitis C virus-coinfected patients. *AIDS* 2011,**25**:899-904.
8. Macias J, Berenguer J, Japon MA, Giron JA, Rivero A, Lopez-Cortes LF, *et al*. Fast fibrosis progression between repeated liver biopsies in patients coinfected with human immunodeficiency virus/hepatitis C virus. *Hepatology* 2009,**50**:1056-1063.
9. Bedimo R, Westfall AO, Mugavero M, Drechsler H, Khanna N, Saag M. Hepatitis C virus coinfection and the risk of cardiovascular disease among HIV-infected patients. *HIV Med* 2010,**11**:462-468.
10. Kotler DP. Hepatitis C, human immunodeficiency virus and metabolic syndrome: interactions. *Liver Int* 2009,**29 Suppl 2**:38-46.
11. Sahin-Efe A, Katsikeris F, Mantzoros CS. Advances in adipokines. *Metabolism* 2012,**61**:1659-1665.
12. Martin LJ, Woo JG, Daniels SR, Goodman E, Dolan LM. The relationships of adiponectin with insulin and lipids are strengthened with increasing adiposity. *J Clin Endocrinol Metab* 2005,**90**:4255-4259.
13. Scherer PE. Adipose tissue: from lipid storage compartment to endocrine organ. *Diabetes* 2006,**55**:1537-1545.
14. Bremer AA, Jialal I. Adipose tissue dysfunction in nascent metabolic syndrome. *J Obes* 2013,**2013**:393192.
15. Xu A, Wang Y, Keshaw H, Xu LY, Lam KS, Cooper GJ. The fat-derived hormone adiponectin alleviates alcoholic and nonalcoholic fatty liver diseases in mice. *J Clin Invest* 2003,**112**:91-100.
16. Bastard JP, Maachi M, Lagathu C, Kim MJ, Caron M, Vidal H, *et al*. Recent advances in the relationship between obesity, inflammation, and insulin resistance. *Eur Cytokine Netw* 2006,**17**:4-12.
17. Bertolani C, Marra F. Role of adipocytokines in hepatic fibrosis. *Curr Pharm Des* 2010,**16**:1929-1940.
18. Grigorescu M, Radu C, Crisan D, Grigorescu MD, Serban A, Neculoiu D, *et al*. Metabolic syndrome, insulin resistance and adiponectin level in patients with chronic hepatitis C. *J Gastrointest Liver Dis* 2008,**17**:147-154.

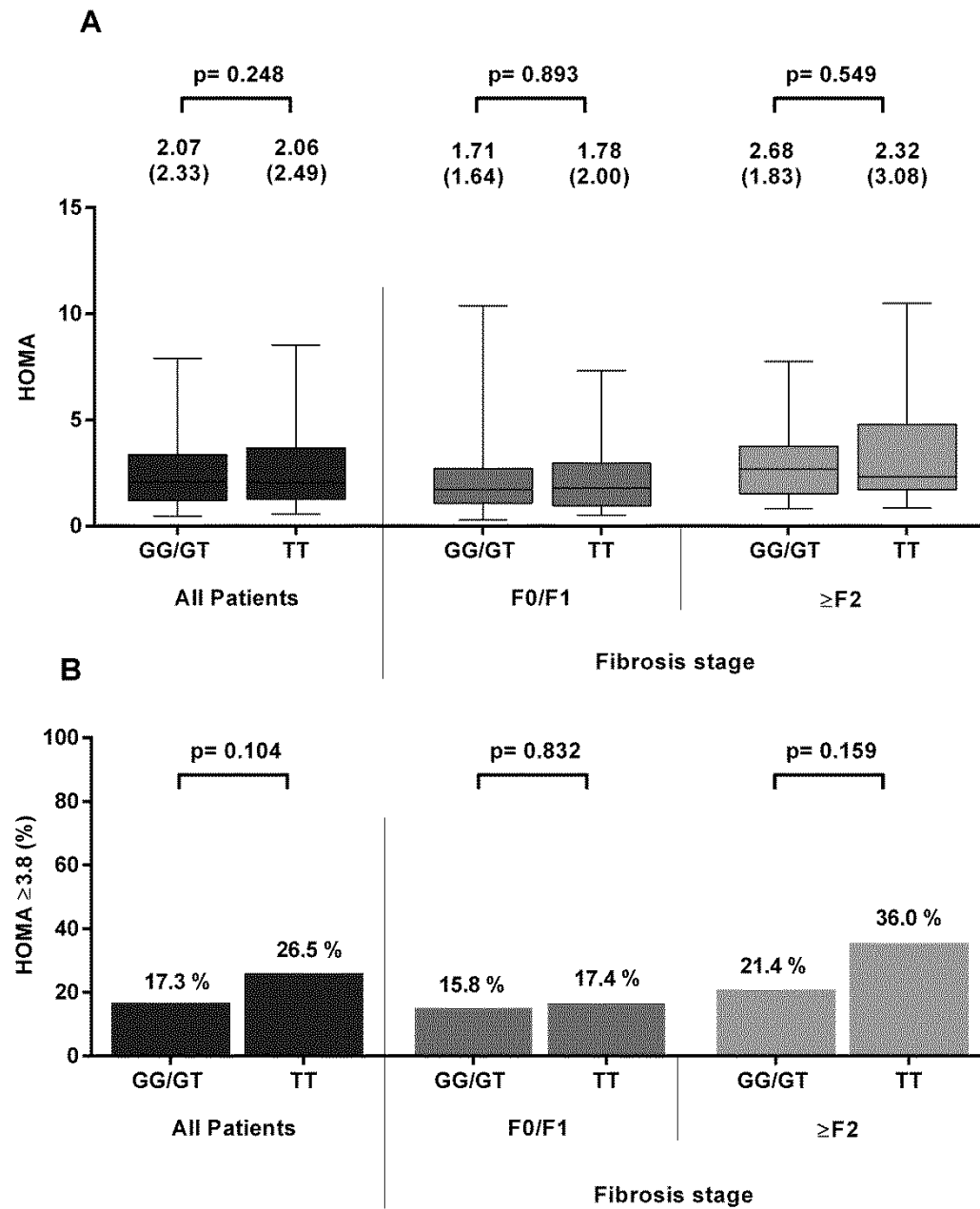
19. Fukushima N, Kuromatsu R, Arinaga-Hino T, Ando E, Takata A, Sumie S, *et al.* Adipocytokine involvement in hepatocellular carcinoma after sustained response to interferon for chronic hepatitis C. *Hepatol Res* 2010,**40**:911-922.
20. Heid IM, Wagner SA, Gohlke H, Iglseider B, Mueller JC, Cip P, *et al.* Genetic architecture of the APM1 gene and its influence on adiponectin plasma levels and parameters of the metabolic syndrome in 1,727 healthy Caucasians. *Diabetes* 2006,**55**:375-384.
21. Zhang H, Mo X, Hao Y, Gu D. Association between polymorphisms in the adiponectin gene and cardiovascular disease: a meta-analysis. *BMC Med Genet* 2012,**13**:40.
22. Han LY, Wu QH, Jiao ML, Hao YH, Liang LB, Gao LJ, *et al.* Associations between single-nucleotide polymorphisms (+45T>G, +276G>T, -11377C>G, -11391G>A) of adiponectin gene and type 2 diabetes mellitus: a systematic review and meta-analysis. *Diabetologia* 2011,**54**:2303-2314.
23. Zhao T, Zhao J. Genetic effects of adiponectin on blood lipids and blood pressure. *Clin Endocrinol (Oxf)* 2011,**74**:214-222.
24. Simeria I, Moher D, Hoey J, Schulz KF, Altman DG. A catalogue of reporting guidelines for health research. *Eur J Clin Invest* 2010,**40**:35-53.
25. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem* 1972,**18**:499-502.
26. Martinez-Hervas S, Real JT, Priego MA, Carratala A, Sniderman AD, Carmena R, *et al.* Establishing cut-off values for apolipoprotein B and non-HDL-C according to LDL-C values in a South European population. *Int J Clin Pract* 2013,**67**:81-88.
27. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985,**28**:412-419.
28. Jacobs WH, Goldberg SB. Statement on outpatient percutaneous liver biopsy. *Dig Dis Sci* 1989,**34**:322-323.
29. Eslam M, Kawaguchi T, Del Campo JA, Sata M, Khatrab MA, Romero-Gomez M. Use of HOMA-IR in hepatitis C. *J Viral Hepat* 2011,**18**:675-684.
30. Bersot TP, Pepin GM, Mahley RW. Risk determination of dyslipidemia in populations characterized by low levels of high-density lipoprotein cholesterol. *Am Heart J* 2003,**146**:1052-1059.
31. Lindsey JK, Jones B. Choosing among generalized linear models applied to medical data. *Stat Med* 1998,**17**:59-68.
32. Cesari M, Narkiewicz K, De Toni R, Aldighieri E, Williams CJ, Rossi GP. Heritability of plasma adiponectin levels and body mass index in twins. *J Clin Endocrinol Metab* 2007,**92**:3082-3088.
33. Mackevics V, Heid IM, Wagner SA, Cip P, Doppelmayr H, Lejnieks A, *et al.* The adiponectin gene is associated with adiponectin levels but not with characteristics of the insulin resistance syndrome in healthy Caucasians. *Eur J Hum Genet* 2006,**14**:349-356.

34. Berthier MT, Houde A, Cote M, Paradis AM, Mauriege P, Bergeron J, *et al.* Impact of adiponectin gene polymorphisms on plasma lipoprotein and adiponectin concentrations of viscerally obese men. *J Lipid Res* 2005,**46**:237-244.
35. Menzaghi C, Trischitta V, Doria A. Genetic influences of adiponectin on insulin resistance, type 2 diabetes, and cardiovascular disease. *Diabetes* 2007,**56**:1198-1209.
36. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Hussain T, Yakout S, *et al.* Adiponectin gene polymorphisms (T45G and G276T), adiponectin levels and risk for metabolic diseases in an Arab population. *Gene* 2012,**493**:142-147.
37. Antonopoulos AS, Tousoulis D, Antoniadis C, Miliou A, Hatzis G, Papageorgiou N, *et al.* Genetic variability on adiponectin gene affects myocardial infarction risk: The role of endothelial dysfunction. *Int J Cardiol* 2012.
38. Kotler DP. Hepatitis C, human immunodeficiency virus and metabolic syndrome: interactions. *Liver Int* 2009,**29 Suppl 2**:38-46.
39. Asselbergs FW, Guo Y, van Iperen EP, Sivapalaratnam S, Tragante V, Lanktree MB, *et al.* Large-scale gene-centric meta-analysis across 32 studies identifies multiple lipid loci. *Am J Hum Genet* 2012,**91**:823-838.
40. Morris AP, Voight BF, Teslovich TM, Ferreira T, Segre AV, Steinthorsdottir V, *et al.* Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet* 2012,**44**:981-990.
41. Manning AK, Hivert MF, Scott RA, Grimsby JL, Bouatia-Naji N, Chen H, *et al.* A genome-wide approach accounting for body mass index identifies genetic variants influencing fasting glycemic traits and insulin resistance. *Nat Genet* 2012,**44**:659-669.
42. Kraja AT, Vaidya D, Pankow JS, Goodarzi MO, Assimes TL, Kullo IJ, *et al.* A bivariate genome-wide approach to metabolic syndrome: STAMPEED consortium. *Diabetes* 2011,**60**:1329-1339.
43. Gable DR, Hurel SJ, Humphries SE. Adiponectin and its gene variants as risk factors for insulin resistance, the metabolic syndrome and cardiovascular disease. *Atherosclerosis* 2006,**188**:231-244.
44. Hara K, Boutin P, Mori Y, Tobe K, Dina C, Yasuda K, *et al.* Genetic variation in the gene encoding adiponectin is associated with an increased risk of type 2 diabetes in the Japanese population. *Diabetes* 2002,**51**:536-540.
45. Siitonen N, Pulkkinen L, Lindstrom J, Kolehmainen M, Eriksson JG, Venojärvi M, *et al.* Association of ADIPOQ gene variants with body weight, type 2 diabetes and serum adiponectin concentrations: the Finnish Diabetes Prevention Study. *BMC Med Genet* 2011,**12**:5.
46. Ukkola O, Ravussin E, Jacobson P, Sjostrom L, Bouchard C. Mutations in the adiponectin gene in lean and obese subjects from the Swedish obese subjects cohort. *Metabolism* 2003,**52**:881-884.
47. Tso AW, Sham PC, Wat NM, Xu A, Cheung BM, Rong R, *et al.* Polymorphisms of the gene encoding adiponectin and glycaemic outcome of Chinese subjects with impaired glucose tolerance: a 5-year follow-up study. *Diabetologia* 2006,**49**:1806-1815.
48. Fumeron F, Aubert R, Siddiq A, Betoulle D, Pean F, Hadjadj S, *et al.* Adiponectin gene polymorphisms and adiponectin levels are independently associated with the development of hyperglycemia during a 3-year period: the epidemiologic data on the insulin resistance syndrome prospective study. *Diabetes* 2004,**53**:1150-1157.

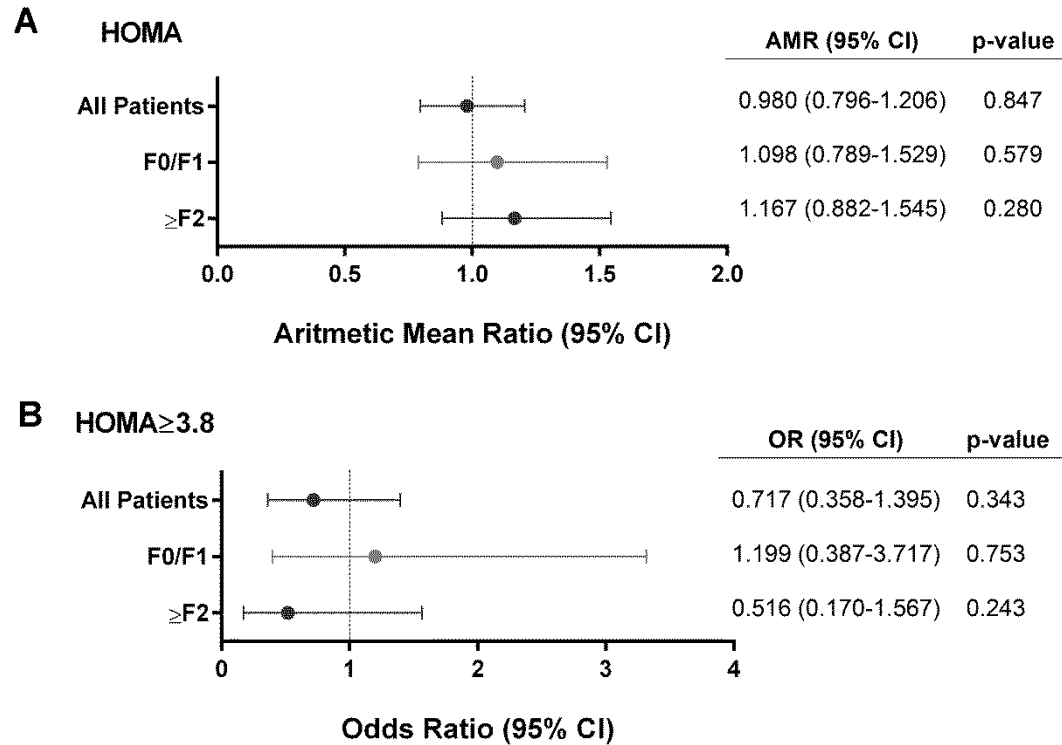
49. Zacharova J, Chiasson JL, Laakso M. The common polymorphisms (single nucleotide polymorphism [SNP] +45 and SNP +276) of the adiponectin gene predict the conversion from impaired glucose tolerance to type 2 diabetes: the STOP-NIDDM trial. *Diabetes* 2005,**54**:893-899.
50. Menzaghi C, Ercolino T, Di Paola R, Berg AH, Warram JH, Scherer PE, *et al.* A haplotype at the adiponectin locus is associated with obesity and other features of the insulin resistance syndrome. *Diabetes* 2002,**51**:2306-2312.
51. Yang WS, Tsou PL, Lee WJ, Tseng DL, Chen CL, Peng CC, *et al.* Allele-specific differential expression of a common adiponectin gene polymorphism related to obesity. *J Mol Med (Berl)* 2003,**81**:428-434.
52. Bowden DW, An SS, Palmer ND, Brown WM, Norris JM, Haffner SM, *et al.* Molecular basis of a linkage peak: exome sequencing and family-based analysis identify a rare genetic variant in the ADIPOQ gene in the IRAS Family Study. *Hum Mol Genet* 2010,**19**:4112-4120.

SUPPLEMENTAL DATA

Supplemental Figure 1. Distribution of HOMA values and insulin resistance according to *ADIPOQ* rs2241766 polymorphism and stratified by liver fibrosis in HIV/HCV coinfecting patients. The median (interquartile range) and p-values (unadjusted GLM test with gamma distribution) are shown for continuous values (**A**). The percentage of patients and p-values (unadjusted GLM with binomial distribution) are shown for HOMA ≥ 3.8 (**B**).



Supplemental Figure 2. Association of *ADIPOQ* rs2241766 polymorphism with HOMA (A) and insulin resistance (HOMA \geq 3.8) (B) by Generalized Linear Model (GLM) tests in HIV/HCV coinfecting patients. These tests were adjusted by several epidemiological and clinical factors [age, gender, BMI, nadir CD4+, HIV plasma viral load, HCV plasma viral load, time on cART and specific antiretroviral drugs].



Supplemental Table 1. Estimation of statistical power performed *a posteriori* for two independent groups (<http://www.imim.cat/ofertadeserveis/software-public/granmo/>), in those cases where the statistical significance was found. The simulation was performed accepting an alpha risk of 0.05 in a two-sided test with 81 subjects in the GG/GT group and 181 in the TT group.

	p-value	Statistical power
Non stratified		
Total Cholesterol	0.038	69%
Total Cholesterol ≥ 200 mg/dL	0.022	71%
Liver steatosis		
Total Cholesterol	0.039	67%
Total Cholesterol ≥ 200 mg/dL	0.003	99%
LDL-C	0.036	77%
Non-HDL-C	0.021	78%