

This is the peer reviewed version of the following article:

Prospective Multicenter Study of Carbapenemase-Producing Enterobacteriaceae from 83 Hospitals in Spain Reveals High In Vitro Susceptibility to Colistin and Meropenem.

Jesús Oteo, Adriana Ortega, Rosa Bartolomé, Germán Bou, Carmen Conejo, Marta Fernández-Martínez, Juan José González-López, Laura Martínez-García, Luis Martínez-Martínez, María Merino, Elisenda Miró, Marta Mora, Ferran Navarro, Antonio Oliver, Álvaro Pascual, Jesús Rodríguez-Baño, Guillermo Ruiz-Carrascoso, Patricia Ruiz-Garbajosa, Laura Zamorano, Verónica Bautista, María Pérez-Vázquez, and José Camposa,

Antimicrob Agents Chemother. 2015;59(6):3406-12.

which has been published in final form at <a href="https://doi.org/10.1128/AAC.00086-15">https://doi.org/10.1128/AAC.00086-15</a>

# Prospective multicenter study of carbapenemase producing

# Enterobacteriaceae from 83 hospitals in Spain: High in vitro

# 3 **susceptibility to colistin and meropenem**

4

1

2

- 5 Jesús Oteo<sup>1,#</sup>, Adriana Ortega<sup>1</sup>, Rosa Bartolomé<sup>2</sup>, Germán Bou<sup>3</sup>, Carmen Conejo<sup>4</sup>,
- 6 Marta Fernández-Martínez<sup>5</sup>, Juan José González-López<sup>2</sup>, Laura Martínez-García<sup>6</sup>, Luis
- 7 Martínez-Martínez<sup>5,7</sup>, María Merino<sup>3</sup>, Elisenda Miró<sup>8</sup>, Marta Mora<sup>9</sup>, Ferran Navarro<sup>8</sup>,
- 8 Antonio Oliver<sup>10</sup>, Álvaro Pascual<sup>4,11</sup>, Jesús Rodríguez-Baño<sup>11,12</sup>, Guillermo Ruiz-
- 9 Carrascoso<sup>13</sup>, Patricia Ruiz-Garbajosa<sup>6</sup>, Laura Zamorano<sup>10</sup>, Verónica Bautista<sup>1</sup>, María
- 10 Pérez-Vázquez<sup>1</sup>, José Campos<sup>1,14</sup>, on behalf of GEIH-GEMARA (SEIMC) and REIPI\*

- 12 <sup>1</sup>Laboratorio de Antibióticos, Bacteriología, Centro Nacional de Microbiología, Instituto
- de Salud Carlos III, Majadahonda, Madrid, Spain.
- <sup>2</sup>Servei de Microbiologia, Hospital Vall d'Hebrón, Universitat Autònoma de Barcelona,
- 15 Barcelona, Spain.
- <sup>3</sup>Servicio de Microbiología-INIBIC, Complejo Hospitalario Universitario A Coruña, A
- 17 Coruña, Spain.
- <sup>4</sup>Departamento de Microbiología, Universidad de Sevilla, Seville, Spain.
- 19 <sup>5</sup>Servicio de Microbiología, Hospital Universitario Marqués de Valdecilla-IDIVAL,
- 20 Santander, Spain.
- 21 <sup>6</sup>Servicio de Microbiología, Hospital Universitario Ramón y Cajal e Instituto Ramón y
- 22 Cajal de Investigación Sanitaria (IRYCIS), Madrid, Spain.
- <sup>7</sup>Departamento de Biología Molecular, Universidad de Cantabria, Santander, Spain.
- <sup>8</sup>Servei de Microbiología, Hospital de la Santa Creu i Sant Pau. Institut d'Investigació
- 25 Biomèdica Sant Pau, Barcelona, Spain.
- <sup>9</sup>Unidad de Microbiología Clínica y Enfermedades Infecciosas, Hospital Universitario La
- 27 Paz-IdiPAZ, Madrid, Spain.
- 28 <sup>10</sup>Servicio de Microbiología, Hospital Son Espases, Instituto de Investigación Sanitaria
- 29 de Palma (IdISPa), Palma de Mallorca, Spain.

- 30 <sup>11</sup>Unidad Clínica de Enfermedades Infecciosas, Microbiología y Medicina Preventiva,
- 31 Hospitales Universitarios Virgen Macarena y Virgen del Rocío, Seville, Spain.
- 32 <sup>12</sup>Departamento de Medicina. Universidad de Sevilla, Seville, Spain.
- 33 <sup>13</sup>Servicio de Microbiología, Hospital Universitario La Paz-IdiPAZ, Madrid, Spain.
- 34 <sup>14</sup>Consejo Superior de Investigaciones Científicas, Madrid, Spain.

35

- 36 \*Other participants from GEIH-GEMARA (SEIMC) and REIPI are listed in the
- 37 Acknowledgements section.
- 38 Abbreviations: GEIH-GEMARA, Grupo de Estudio de Infección Hospitalaria-Grupo
- 39 de Estudio de Mecanismos de Acción y Resistencia a Antimicrobianos; REIPI, Red
- 40 Española de Investigación en Patología Infecciosa; SEIMC, Sociedad Española de
- 41 Enfermedades Infecciosas y Microbiología Clínica.
- 42 **Keywords:** carbapenem resistance, population structure, geographic distribution,
- 43 prevalence.
- 44 **Running head:** Carbapenemase producing Enterobacteriaceae in Spain, 2013.

- **\*Corresponding author:**
- 47 Jesús Oteo, Centro Nacional de Microbiología, Instituto de Salud Carlos III, Carretera
- 48 Pozuelo a Majadahonda, 28220 Majadahonda, Madrid, Spain.
- 49 Phone: ++34 918 22 3650. Fax: ++34 915097966. E-mail: jesus.oteo@isciii.es

### Abstract

50

51 The aim of this study was to determine the impact of the carbapenemase-52 producing Enterobacteriaceae (CPE) in Spain in 2013 by describing their prevalence, 53 dissemination and geographic distribution of CPE clones, their population structure and 54 antibiotic susceptibility. 55 From February 2013 to May 2013, 83 hospitals (about 40,000 hospital beds) 56 prospectively collected non-duplicate Enterobacteriaceae using the screening cut-off 57 recommended by EUCAST. Carbapenemase characterisation was performed by 58 phenotypic methods and confirmed by PCR and sequencing. MLST types were 59 determined for Klebsiella pneumoniae and Escherichia coli. A total of 702 Enterobacteriaceae isolates met the inclusion criteria; 379 (54%) were 60 61 CPE. OXA-48 (71.5%) and VIM-1 (25.3%) were the most frequent carbapenemases, 62 and K. pneumoniae (74.4%), Enterobacter cloacae (10.3%), and E. coli (8.4%) were the 63 species most affected. Susceptibility to colistin, amikacin and meropenem was 95.5%, 64 81.3%, and 74.7%, respectively. The most prevalent STs were ST11 and ST405 in K. 65 pneumoniae, and ST131 in E. coli. Forty-five (54.1%) of the hospitals had at least one 66 CPE case. In K. pneumoniae, ST11/OXA-48, ST15/OXA-48, ST405/OXA-48, and ST11/VIM-1 were detected in two or more Spanish provinces. ST11 carried four 67 68 carbapenemases (VIM-1, OXA-48, KPC-2, and OXA-245), but ST405 carried OXA-48 69 only. 70 A wide interregional spread of CPE in Spain was observed mainly due to a few 71 successful clones of OXA-48-producing K. pneumoniae (e.g. ST11 and ST405). 72 Dissemination of OXA-48-producing E. coli is a new finding of public health concern. 73 According to in vitro susceptibilities, most of the CPE (94.5%) had three or more 74 options of antibiotic treatment.

#### Introduction

Carbapenemase-producing Enterobacteriaceae (CPE), mainly *Klebsiella pneumoniae*, are an emerging threat to public and individual health worldwide. These microorganisms are often resistant to almost all available antibiotics (1,2), so there are few alternative treatment options. The most common carbapenemases are KPC (class A); VIM, IMP, and NDM (class B); and the OXA-48 types (class D). However, the extent to which healthcare systems have been affected, and the predominant carbapenemase types, differ substantially from country to country (3).

A multicenter study performed in Spain in 2009 revealed 43 (0.04%) cases of CPE, which were mostly VIM-1 and IMP-22 (4). After that, we reported a rapid increase in the number of cases of CPE, mainly OXA-48-producing *K. pneumoniae*, in this country from 2010–2012 (5-7).

Because previous studies (5,6) were based on voluntary reports without taking into account key important issues, in this manuscript we present data on the impact of CPE as obtained from a prospective, multicenter and population-based study. We show carbapenemase-production in this country is widely and irregularly distributed; however susceptibility rates to meropenem and colistin were still high so far.

## Material and methods

Study design and bacterial isolates

A prospective multicentre study was designed to identify Enterobacteriaceae isolates with decreased susceptibility to carbapenems. Isolates were collected from clinical infections and carriers between February and May, 2013. Eighty-three Spanish hospitals from 33 out of the 50 Spanish provinces participated in the study; these 33 provinces belonged to 15 of the 17 Spanish Autonomous Communities. Eight of the hospitals acted as coordinating centers. The estimated catchment population was

approximately one-half of the Spanish population, and consisted of approximately 21.7 million individuals and 40,100 hospital beds. The participating hospitals registered the total number of infections caused by Enterobacteriaceae during the study period so that the values for CPE prevalence could be estimated; presence of infections was established according to previously defined criteria (8).

EUCAST screening cut-off values were used to identify CPE (9). The inclusion criteria were all Enterobacteriaceae isolates presenting either MICs >0.125 mg/L to meropenem and/or ertapenem and/or >1 mg/L to imipenem, or disk inhibition zones obtained using the disk diffusion method <25 mm to meropenem and/or ertapenem and/or ertapenem and/or <23 mm to imipenem. Only one isolate per patient and species was considered for further analysis. Isolates from the genera *Proteus*, *Providencia*, and *Morganella* that had reduced susceptibility to imipenem, but were susceptible to ertapenem and meropenem, were not included in the analysis; in addition, *Enterobacter* isolates that had reduced susceptibility to ertapenem, but were susceptible to imipenem and meropenem, were also excluded.

### Bacterial identification and drug-susceptibility testing

The initial assays on the isolates were performed at each participating hospital, using standard microbiological methods. Each hospital also submitted their isolates to one of the eight coordinating centres, where carbapenemase production was confirmed using phenotypic and genotypic methods. Finally, all study isolates were submitted to the antibiotic laboratory of the Spanish National Centre of Microbiology, which acted as a central reference laboratory. All isolates meeting the phenotype inclusion criteria (9) were classified using the algorithm for phenotypic carbapenemase detection recommended by EUCAST (9). A modified Hodge test using a meropenem disk with

cloxacillin (600  $\mu g$ ) was performed on all isolates. In addition, inhibition of carbapenemase activity was carried out by comparing the inhibition zones obtained from meropenem disks, with or without EDTA (10  $\mu L$  0.5 M solution), phenyl-boronic acid (400  $\mu g$ ), and cloxacillin (600  $\mu g$ ) in all isolates. Carba NP method was used as confirmatory test of carbapenemase activity when unclear phenotypic results or discrepancies between phenotypic and genotypic results, were observed (10).

Antibiotic susceptibility testing was performed by disk-difussion and microdilution susceptibility methods according to EUCAST guidelines (11,12) in addition susceptibility to ertapenem, imipenem, meropenem and colistin were carried out by gradient test (bioMérieux, Marcy-l'Étoile, France).

Extended spectrum  $\beta$ -lactamase (ESBL) production in OXA-48 and class B carbapenemase producers was suspected if activity of cefotaxime or aztreonam, respectively, was recovered in presence of clavulanic acid. In the case of KPC-producers molecular characterization of ESBL genes were carried out in all isolates.

## Characterisation of resistance mechanisms

The presence of genes encoding carbapenemases ( $bla_{OXA-48}$ ,  $bla_{KPC}$ ,  $bla_{VIM}$ ,  $bla_{IMP}$ , and  $bla_{NDM}$ ) (5) and ESBLs ( $bla_{TEM}$ ,  $bla_{SHV}$ , and  $bla_{CTX-M}$ ) (13) was determined using PCR and DNA sequencing assays.

## Molecular epidemiology

MLST was performed for all carbapenemase-producing *K. pneumoniae* using the Institut Pasteur scheme (http://www.pasteur.fr/recherche/genopole/PF8/mlst/Kpneumoniae.html; accessed January 2015). Carbapenemase-producing *E. coli* were typed by MLST using the

University of Warwick (Warwick Medical School, Coventry, UK) scheme (http://mlst.warwick.ac.uk; accessed January 2015). The phylogenetic relationships among the different sequence types (STs) found in this study were established according to the eBURST program version 3 (http://eburst.mlst.net).

A simple diversity index (SD) previously described by Grundmann et al. (14) was applied to analyze the population diversity as follows: number of STs/total number of isolates x 100.

We considered that two or more isolates of *K. pneumoniae* or *Escherichia coli* (including clinical cases and carriers) were epidemiologically related if they belonged to the same species, had the same MLST type, and produced the same carbapenemase type. For *E. cloacae*, this epidemiological association was established when the genetic linkage was >85% using PFGE after total chromosomal DNA digestion with *Xba*I (13).

## Statistical analysis

Differences in the prevalence values for resistance mechanisms between the different groups were assessed using Fisher's exact test. Associations were determined by calculating ORs with 95% CIs. The null hypothesis was rejected for values of P < 0.05. Statistical analysis was performed using GraphPad Prism, version 3.02, software (GraphPad Software, Inc., San Diego, CA, USA).

### **Results and Discussion**

172 Bacterial isolates and carbapenemases types

A total of 702 Enterobacteriaceae isolates that were collected from an equal number of patients met the phenotypic criteria for inclusion; being *K. pneumoniae* (53.3%), *E. cloacae* (15.8%), and *E. coli* (13.8%) the most common species (Table 1).

Of these 702 isolates, 379 (54%) were confirmed as CPE and were distributed as 176 177 follows: 282 (74.4%) were K. pneumoniae, 39 (10.3%) E. cloacae, 32 (8.4%) E. coli, 11 178 (2.9%) Klebsiella oxytoca, 7 (1.8%) Citrobacter freundii, 4 (1.1%) Serratia 179 marcescens, 2 (0.5%) Enterobacter aerogenes, 1 (0.3%) Morganella morganii, and 1 180 (0.3%) isolate *Enterobacter* sp. (Table 1). The percentages of CPE isolates significantly 181 varied between species as follows: 75.4% (282 of 374) of the K. pneumoniae, 35.1% 182 (39 of 111) of the E. cloacae, and 33% (32 of 97) of the E. coli isolates; the percentage 183 obtained for K. pneumoniae was significantly higher than those for E. cloacae and E. 184 coli (P < 0.0001). In a recent French study, 1485 Enterobacteriaceae isolates non 185 susceptible to carbapenems according EUCAST criteria were detected (42.2% and 186 35.2% were *Enterobacter* spp. and *K. pneumoniae*, respectively); of them 340 (22.9%) 187 isolates were carbapenemase producers (65.9% and 9.7% were K. pneumoniae and 188 Enterobacter spp., respectively) (15). 189 Of the 379 CPE isolates detected, 300 (79.2%) were isolated from clinical 190 samples, and were mostly from urine (158, 52.7%), wound (59, 19.7%), respiratory (37, 191 12.3%), and blood (28, 9.3%) samples. The remaining 79 CPE (20.2%) isolates were 192 from screening samples (81% from rectal or perianal samples). In total, 193 CPE cases 193 (50.9%) were isolated from males, and 239 (63.1%) were from patients >65 years of 194 age. 195 The carbapenemase types identified were: 271 (71.5%) OXA-48 group (258 196 OXA-48 and 13 OXA-245), 96 (25.3%) VIM (95 VIM-1 and one VIM-2), 8 (2.1%) 197 KPC-2, and 6 (1.6%) IMP (4 IMP-13 and two IMP-22). Each of two K. oxytoca isolates 198 produced both KPC-2 and VIM-1, these two isolates came from two different hospitals 199 from the Madrid area, and were isolated in March-2013 and May-2013. Isolates from 200 carriers had the following carbapenemase types: 69.6% OXA-48 group, 30.3% VIM

group, and 1.3% KPC. *K. pneumoniae* was more prevalent among the OXA-48 isolates (86.7%) than among the VIM isolates (44.8%) (P < 0.0001). Comparison of OXA-48-group- and VIM-group-producing isolates is showed in Table 2.

OXA-48 prevalence is also increasing in other European countries as France, Germany, Belgium, and the United Kingdom, where increasing numbers of outbreaks have been described (1,16-20). However, OXA-48 is rarely identified in North America (21). Compared with OXA-48 and VIM, KPC was identified very infrequently in this study. However, KPC-producing Enterobacteriaceae have already caused sporadic hospital outbreaks in Spain (3,22). KPC enzymes are endemic in other European countries, such as Greece and Italy (1,3,23), and it produces nosocomial outbreaks in North America (1). The number of carbapenemase-producing *E. coli* identified in this study was much higher than in previous studies (4,5). This remarkable finding is of serious concern because *E. coli* may facilitate the community spread of carbapenemases.

ESBL production was detected in 267 of the 379 CPE (70.4%); 227 (85%) produced CTX-M-15, 16 (6%) SHV-12, 15 (5.6%) CTX-M-9, 9 (3.4%) SHV-134, 1 (0.4%) produced CTX-M-14, and 1 (0.4%) produced CTX-M-1. Two CPE isolates had two different types of ESBLs. Carbapenemase-producing *K. pneumoniae* more frequently co-produced ESBLs (235 of 282 isolates, 83.3%) than did *E. cloacae* (18 of 38 isolates, 47.4%) or *E. coli* (11 of 32, 34.4%) (*P* < 0.0001). Co-production of ESBLs occurred in the 90.6% and 28% of OXA-48-producing *K. pneumonia* and *E. coli*, respectively (*P* < 0.0001), mostly CTX-M-15. Potron *et al.* (24) found that co-production of OXA-48 and CTX-M-15 occurred in 41.7% of OXA-48-producing *E. coli* isolates from 10 different European and African countries. Five OXA-48- and SHV-12-producing *K. pneumoniae* belonged to ST147 were isolated in Asturias (northern

Spain). Associations between OXA-48 and ESBLs of the SHV-type have previously been infrequently reported (24).

Antibiotic susceptibility testing.

Overall the antibiotics showing the highest percentages of susceptibility were colistin (95.5%), amikacin (81.3%), meropenem (74.7%), tigecycline (71%), and imipenem (67.6%) (Table 3). However, antibiotic susceptibility significantly varied between the OXA-48-producing and VIM-producing isolates (Table 3), the VIM-producing isolates usually being more resistant.

A total of 182 CPE (48%) were susceptible to colistin, amikacin, tigecycline, and carbapenems (imipenem or meropenem); of them, 107 were also susceptible to fosfomycin. According to previous clinical experience (2), use of a carbapenem (meropenem or imipenem) for the treatment of a CPE with a MIC of ≤8 mg/L, in combination with another active agent, seems reasonable; following this criteria, we identified 21 highly resistant CPE isolates (5.5%; 12 OXA-48 isolates, 8 VIM-1 and one KPC-2) presenting only one or two options to choose from for clinical purposes, mainly colistin and carbapenems (n=5) or colistin and tigecycline (n=5).

Geographic distribution and prevalence of infections due to CPE in Spain.

At least one case of CPE was identified at 45 (54.1%) of the 83 participating hospitals (Figure 1). These hospitals were located in 18 of the 33 (54.5%) participating provinces. In twenty one (46.7%) of the 45 hospitals with CPE isolates, potential outbreaks of epidemiological related CPE were detected affecting 209 (55.1%) of the 379 CPE isolates included in this study.

Data about the total numbers of infections caused by Enterobacteriaceae during the study period were provided by 75 (90.4%) of the participating hospitals. A total of 120,808 single infections by patient and specie were identified. The estimated overall prevalence of infection by carbapenemase-producing *K. pneumoniae* was 1.7% (231/13842; range: 0–11.6%), and 23 (30.7%) hospitals had a prevalence >1%. For *Enterobacter* spp. and *E. coli*, these figures were 0.5% (28/5085; range: 0–6.4%) and 0.03% (28/91553; range: 0-0.4%), respectively. The prevalence of carbapenemase-production in *K. pneumoniae* and *E. coli* in a multicenter study performed in Spain in 2009 was 0.2% and 0.001%, respectively (4).

Although the number of OXA-48 producing isolates was greater than the number of VIM-producing isolates (271 *vs.* 96, Table 1), the VIM-producing isolates were more widely geographically distributed. VIM-producing isolates were detected in 16 (40.5%) provinces and OXA-48-producing isolates were detected in 10 (30.3%). This finding could be related to the earlier detection of VIM in Spain that occurred in 2005 (25), compared with OXA-48 in 2009 (26).

Population structure of carbapenemase-producing K. pneumoniae and E. coli isolates causing clinical infections.

Using MLST, 24 different sequence-types (ST) were identified among the 221 carbapenemase-producing *K. pneumoniae* implicated in clinical infections (SD: 10.6; mean: 9.2 isolates per ST; range: 1–66). The most prevalent STs were ST11 (66 isolates, 29.9%), ST405 (65, 29.4%), ST15 (28, 12.7%) and ST326 (15, 6.8%). These four STs were found in 78.8% of the carbapenemase-producing *K. pneumoniae* isolates. ST405 and ST11 clinical isolates were isolated in 18 different each one. ST11 carried four different types of carbapenemases (VIM-1, OXA-48, KPC-2, and OXA-245), but

ST405 carried only OXA-48. Analysis with eBURST provided an overview of STs of the Spanish carbapenemase producing *K. pneumoniae* isolates from this study compared with all *K. pneumoniae* isolates from the MLST database. The major ST found are represented in Figure 2. Excluding the isolates from this study, ST11 was the second most frequent ST in the MLST database (103 of 2405, 4.28%) and ST405 was uncommon (4 of 2405, 0.17%). In this eBurst analysis, ST405 was shown as a singleton non-related with other classical STs (Figure 2). These results suggest that *K. pneumoniae* belonging to ST11 and ST405 strongly contribute to the dissemination of OXA-48-producing Enterobacteriaceae in Spain. ST11 has been identified as an international high-risk clone associated ESBL- and carbapenemase production (27,28), but ST405 has recently been associated with OXA-48 production in Spain, Belgium, and France (5,6,17,18).

Among the 27 carbapenemase-producing *E. coli* implicated in clinical infections, 16 different STs were identified (SD: 59.3; mean: 1.7 isolates per ST; range: 1–7). ST131 (seven isolates, 26%) and ST156 (three isolates, 11.1%) were the most prevalent STs identified. These findings suggest that, compared with *K. pneumoniae*, carbapenemase-producing *E. coli* isolates may have a more diverse and polyclonal population structure, as previously demonstrated for other resistance mechanisms like ESBL (29). The acquisition of carbapenemases by the globally distributed *E. coli* ST131 that has been detected in this and other studies (30, 31) is a finding of serious concern. In a recent study of OXA-48-producing *K. pneumoniae* and *E. coli* in several European and north-African countries, the most common STs identified were ST101 and ST38, respectively (24).

Potential interregional spread of CPE strains.

One finding with epidemiological implications was that some of the more prevalent *K. pneumoniae* clones carrying carbapenemases were identified in more than one Spanish province suggesting that potential interregional spread of these clones may be occurred; however, the possibility of OXA-48 encoding plasmids acquisition by two isolates of a same prevalent ST from different regions cannot be excluded. ST405/OXA-48 was detected in six different Spanish provinces, ST15/OXA-48 in four, and ST11/OXA-48 and ST11/VIM-1 in three each one. This interregional spread clearly indicates that further progress has occurred since the "independent hospitals outbreak" stage described by Canton et al. in 2010 (1).

In addition, these carbapenemase genes are plasmid encoded and therefore their spread is probably due to both the clonal dissemination of a few specific strains and the transmission of epidemic auto-transferred plasmids carrying them (1,6,22).

## **Conclusions**

We found that there was a wide geographic distribution of CPE species, and a clear increasing trend in the number of infections caused to CPE in Spain. Two successful clones of *K. pneumoniae* (ST11 and ST405) carried mainly OXA-48, while ST15 more often carried VIM. Although still infrequent, detection of a polyclonal dissemination of OXA-48-producing *E. coli* has serious implications for public health.

According to *in vitro* susceptibilities, most of the CPE (94.5%) had three or more options of antibiotic treatment.

The spread of CPE in Spain affected at least 68% of all provinces with a potential interregional spread of CPE strains. This finding also suggests that the public health situation posed by CPE has worsened in the last few years in Spain.

## 326 Acknowledgements

327 Members of GEIH-GEMARA (SEIMC) and REIPI participating in this study: Ángel 328 Zaballos (Centro Nacional de Microbiología, Majadahonda, Madrid); Rafael Cantón (Hospital 329 Universitario Ramón y Cajal, Madrid); Ana María Fleites and Carlos Rodríguez-Lucas 330 (Hospital Universitario Central de Asturias); Mª Isabel Sánchez-Romero (Hospital Universitario 331 Puerta de Hierro, Majadahonda, Madrid); Luisa García-Picazo (Hospital El Escorial, Madrid); 332 Esteban Aznar and Carolina Campelo (Laboratorio BRSalud, San Sebastián de los Reyes, 333 Madrid); Alejandro González-Praetorius and Sonia Solís (Hospital Universitario de 334 Guadalajara); Salvador Giner and Miguel Salavert (Hospital Universitari i Politècnic La Fe, 335 Valencia); Juan Manuel Hernández (Hospital Carlos Haya, Málaga); Josep Vilaró Pujals and 336 Anna Vilamala Bastarras (H. General de Vic, Barcelona); Mª Ángeles Orellana (Hospital 12 de 337 Octubre); Emilia Cercenado (Hospital General Universitario Gregorio Marañón, Madrid); 338 Mateu Espasa and Dionisia Fontanals (Corporació Sanitària Parc Taulí, Sabadell, Barcelona); 339 Mª Victoria García-López (Hospital Clínico de Málaga); José Luis Hernández-Almaraz 340 (Hospital de Cruces, Barakaldo, Vizcaya); Carmina Martí-Sala (Hospital General de Granollers, 341 Barcelona); Adelina Gimeno (Hospital Universitario de Alicante); Teresa Alarcón and Laura 342 Llorca (Hospital Universitario de la Princesa, Madrid); Concepción Segura (Laboratori de 343 Referència de Catalunya, Barcelona); Raquel Clivillé-Abad (Sant Joan Despí Moisès Broggi, 344 CLI, Barcelona); Montse Motjé and Delia Garcia i Parés (Hospital Universitario de Girona Dr. 345 Josep Trueta, Girona); Pedro de la Iglesia and Beatriz Iglesias (Hospital San Agustín de Avilés, 346 Asturias); Juanjo Castón and María Dolores Romero (Hospital de Ciudad Real); José Antonio 347 Rodríguez-Polo (Hospital Virgen de la Salud, Toledo); Gloria Trujillo and Montserrat Morta 348 (Hospital San Joan de Deu de Manresa, Barcelona); Alberto Gil Setas and Carmen Ezpeleta 349 (Complejo Hospitalario de Navarra); Mª Dolores Miguel-Martínez (Hospital de Cabueñes, 350 Gijón); Antonio Sánchez-Porto and Javier Casas (Hospital del SAS de la Línea, Cádiz); David 351 Molina (Hospital Universitario de Getafe, Madrid); Eugenio Garduño (Complejo Hospitalario 352 Universitario de Badajoz); Juan Carlos Alados (Hospital del SAS de Jerez de la Frontera, 353 Cádiz); Pepa Pérez-Jové (Mutua de Terrassa, Consorcio Sanitario de Terrassa, Hospital 354 Fundación Sant Joan de Déu de Martorell); Goretti Sauca (Hospital de Mataró, Barcelona); 355 Carmen Gallés (Corporació de Salut del Maresme i La Selva, Barcelona); Fátima Galán and 356 Francisca Guerrero (Hospital Puerta del Mar, Cádiz); Mª Fe Brezmes (Complejo Asistencial de 357 Zamora); Ma Pilar Ortega (Complejo Asistencial de Burgos); Francisco Javier Castillo and 358 Cristina Seral (Hospital Clínico Universitario Lozano Blesa, Zaragoza); Alberto Delgado-359 Iribarren (Hospital Universitario Fundación Alcorcón, Madrid); Alberto Yagüe (Hospital "La 360 Plana", Villarreal, Castellón); Carmen Aspiroz (Hospital Royo Villanova, Zaragoza); Ma Isabel 361 Fernández-Natal (Complejo Asistencial Universitario de León); Isabel Wilhelmi and Pilar

362 Reyes (Hospital Universitario Severo Ochoa, Leganés, Madrid); Mª Dolores Pérez-Ramírez 363 (Hospital Universitario Virgen de las Nieves, Granada); Inocente Cuesta (Complejo 364 Hospitalario de Jaén); Mar Olga Pérez Moreno (Hospital de Tortosa Verge de la Cinta, Tortosa, 365 Tarragona); Amparo García (Hospital General de Igualada, CLI, Barcelona); Frederic Ballester 366 and Isabel Pujol (Laboratori de Referència Sud, Hospital Universitari Sant Joan, Reus, 367 Tarragona); Montserrat Sierra (Hospital de Barcelona-SCIAS); Araceli González-Cuevas 368 (Hospital General del Parc Sanitari Sant Joan de Deu, Sant Boi de Llobregat, Barcelona); Pilar 369 López García (Hospital General Universitario de Elche, Alicante); Lluís Carbó Saladrigas 370 (Hospital Mateu Orfila, Mahón, Menorca); Jesús Martínez-López (Complejo Hospitalario de 371 Pontevedra); Lucía Martínez-Lamas and Jorge Julio Cabrera (Complejo Hospitalario 372 Universitario de Vigo, Pontevedra); Susana García de Cruz (Complejo Hospitalario de Soria); 373 Carmen Raya (Hospital del Bierzo, Ponferrada); Ana Belén Campo and Inés de Benito 374 (Hospital Sierrallana, Torrelavega, Cantabria); Andrés Canut (Hospital Universitario de Álava); 375 Pilar Berdonces (Hospital de Galdakao); María Concepción Lecaroz Agara (Hospital 376 Universitario de Álava-Txagorritxu); Susana Hernando Real (Hospital General de Segovia); 377 Belén Hernández (Hospital Universitario Niño Jesús, Madrid); Mª Teresa Ledo and Firdaous El 378 Knaichi (Hospital Universitario de Torrejón); Carlos García Tejero (Hospital Virgen del Puerto, 379 Plasencia, Cáceres); Jose Manuel Azcona (Hospital San Pedro, Logroño, La Rioja); Isabel 380 Ferrer (Hospital Universitario Miguel Servet de Zaragoza); Marta Lamata (Fundación Hospital 381 de Calahorra, La Rioja); Carmen Pazos (Hospital San Pedro de Alcántara de Cáceres); Mª Pilar 382 Chocarro (Hospital Obispo Polanco, Teruel).

383

- We thank the Genomics Unit of the Centro Nacional de Microbiología for support with
- 385 DNA sequencing.
- 386 The preliminary results of this study were presented, in part, at the 24th European
- 387 Congress of Clinical Microbiology and Infectious Diseases annual meeting on 10-13
- 388 May 2014 in Barcelona, Spain (Abstract eP-953).

389

390

### Funding

- 391 This work was supported by a grant from the Fondo de Investigación Sanitaria (grant
- 392 number PI12/01242); the Antibiotic Resistance Surveillance Programme of the Spanish
- 393 Centro Nacional de Microbiología, Instituto de Salud Carlos III, Ministerio de

394 Economía y Competitividad; the Plan Nacional de I+D+I 2008-2011; and the Instituto 395 de Salud Carlos III, Subdirección General de Redes y Centros de Investigación 396 Cooperativa, Ministerio de Economía y Competitividad, Spanish Network for Research 397 in Infectious Diseases (REIPI RD12/0015) – co-financed by European Development 398 Regional Fund "A Way to Achieve Europe" ERDF. 399 400 **Conflict of interest** 401 L. M-M. was speaker for Merck, Pfizer, Janssen-Cilag, and Astra-Zeneca and received 402 research support from Merck, Wyeth, and Janssen-Cilag and Astra-Zeneca.

J. R-B. was speaker for Merck, AstraZeneca, Astellas, Novartis and Pfizer, served as 403

scientific advisor for Merck, AstraZeneca, Roche and Achaogen and received research

405 grants from Novartis.

406 None of these poses a conflict of interest with this work.

407

- 408 **References.**
- 409 1. Cantón R, Akóva M, Carmeli Y, Giske CG, Glupczynski Y, Gniadkowski M,
- 410 Livermore DM, Miriagou V, Naas T, Rossolini GM, Samuelsen Ø, Seifert H,
- Woodford N, Nordmann P; European Network on Carbapenemases. 2012. Rapid
- 412 evolution and spread of carbapenemases among Enterobacteriaceae in Europe. Clin
- 413 Microbiol Infect **18**: 413-431.
- 2. Tzouvelekis LS, Markogiannakis A, Piperaki E, Souli M, Daikos GL. 2014.
- 415 Treating infections caused by carbapenemase-producing Enterobacteriaceae. Clin
- 416 Microbiol Infect **20**: 862-872.
- 417 3. Glasner C, Albiger B, Buist G, Tambić Andrasević A, Canton R, Carmeli Y,
- 418 Friedrich AW, Giske CG, Glupczynski Y, Gniadkowski M, Livermore DM,
- 419 Nordmann P, Poirel L, Rossolini GM, Seifert H, Vatopoulos A, Walsh T,
- 420 Woodford N, Donker T, Monnet DL, Grundmann H; European Survey on
- 421 Carbapenemase-Producing Enterobacteriaceae (EuSCAPE) Working Group.
- 422 2013. Carbapenemase-producing Enterobacteriaceae in Europe: a survey among
- national experts from 39 countries, February 2013. Euro Surveill **18(28)**: pii: 20525.
- 424 4. Miró E, Agüero J, Larrosa MN, Fernández A, Conejo MC, Bou G, González-
- 425 López JJ, Lara N, Martínez-Martínez L, Oliver A, Aracil B, Oteo J, Pascual A,
- 426 Rodríguez-Baño J, Zamorano L, Navarro F. 2013. Prevalence and molecular
- 427 epidemiology of acquired AmpC β-lactamases and carbapenemases in
- 428 Enterobacteriaceae isolates from 35 hospitals in Spain. Eur J Clin Microbiol Infect Dis
- 429 **32**: 253-259.
- 5. Oteo J, Saez D, Bautista V, Fernández-Romero S, Hernández-Molina JM, Pérez-
- Vázquez M, Aracil B, Campos J; Spanish Collaborating Group for the Antibiotic
- 432 **Resistance Surveillance Program**. 2013. Carbapenemase-producing
- enterobacteriaceae in Spain in 2012. Antimicrob Agents Chemother **57**: 6344-6347.
- 6. Oteo J, Hernández JM, Espasa M, Fleites A, Sáez D, Bautista V, Pérez-Vázquez
- 435 M, Fernández-García MD, Delgado-Iribarren A, Sánchez-Romero I, García-
- 436 Picazo L, Miguel MD, Solís S, Aznar E, Trujillo G, Mediavilla C, Fontanals D,
- 437 **Rojo S, Vindel A, Campos J,** et al. 2013. Emergence of OXA-48-producing Klebsiella
- 438 pneumoniae and the novel carbapenemases OXA-244 and OXA-245 in Spain. J
- 439 Antimicrob Chemother **68**: 317-321.
- 440 7. Paño-Pardo JR, Ruiz-Carrascoso G, Navarro-San Francisco C, Gómez-Gil R,

- 441 Mora-Rillo M, Romero-Gómez MP, Fernández-Romero N, García-Rodríguez J,
- 442 **Pérez-Blanco V, Moreno-Ramos F, Mingorance J**. 2013. Infections caused by OXA-
- 443 48-producing Klebsiella pneumoniae in a tertiary hospital in Spain in the setting of a
- prolonged, hospital-wide outbreak. J Antimicrob Chemother 68: 89-96.
- 8. Horan TC, Andrus M, Dudeck MA. 2008. CDC/NHSN surveillance definition of
- health care–associated infection and criteria for specific types of infections in the cute
- care setting. Am J Infect Control **36**: 309-332.
- 9. EUCAST guidelines for detection of resistance mechanisms and specific resistances
- of clinical and/or epidemiological importance. Version 1.0, December 2013. Available
- 450 in:
- 451 http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\_files/Resistance\_mechanis
- 452 <u>ms/EUCAST\_detection\_of\_resistance\_mechanisms\_v1.0\_20131211.pdf</u> (July 2014,
- 453 data last accessed)
- 454 10. Nordmann P, Poirel L, Dortet L. 2012. Rapid detection of carbapenemase-
- producing Enterobacteriaceae. Emerg Infect Dis **18**: 1503-1507.
- 456 11. Matuschek E, Brown DF, Kahlmeter G. 2014. Development of the EUCAST disk
- 457 diffusion antimicrobial susceptibility testing method and its implementation in routine
- 458 microbiology laboratories. Clin Microbiol Infect **20**: O255-O266.
- 459 12. Clinical laboratory testing and in vitro diagnostic test systems Susceptibility
- 460 testing of infectious agents and evaluation of performance of antimicrobial
- susceptibility test devices Part 1: Reference method for testing the in vitro activity of
- antimicrobial agents against rapidly growing aerobic bacteria involved in infectious
- 463 diseases. ISO 20776-1, EUCAST 2006.
- 13. Oteo J, Navarro C, Cercenado E, Delgado-Iribarren A, Wilhelmi I, Orden B,
- García C, Miguelañez S, Pérez-Vázquez M, García-Cobos S, Aracil B, Bautista V,
- 466 Campos J. 2006. Spread of Escherichia coli strains with high-level cefotaxime and
- 467 ceftazidime resistance between the community, long-term care facilities, and hospital
- institutions. J Clin Microbiol 44: 2359-2366.
- 14. **Gastmeier P, Schwab F, Bärwolff S, Rüden H, Grundmann H**. 2006. Correlation
- 470 between the genetic diversity of nosocomial pathogens and their survival time in
- intensive care units. J Hosp Infect **62**: 181-186.
- 472 15. Dortet L, Cuzon G, Nordmann P. 2014. Dissemination of carbapenemase-
- producing Enterobacteriaceae in France, 2012. J Antimicrob Chemother **69**: 623-627.
- 16. Robert J, Pantel A, Mérens A, Lavigne JP, Nicolas-Chanoine MH; ONERBA's

- 475 Carbapenem Resistance Study Group. 2014. Incidence rates of carbapenemase-
- 476 producing Enterobacteriaceae clinical isolates in France: a prospective nationwide
- 477 study in 2011-12. J Antimicrob Chemother **69**: 2706-2712.
- 478 17. Glupczynski Y, Huang TD, Bouchahrouf W, Rezende de Castro R, Bauraing C,
- 479 Gérard M, Verbruggen AM, Deplano A, Denis O, Bogaerts P. 2012. Rapid
- 480 emergence and spread of OXA-48-producing carbapenem-resistant *Enterobacteriaceae*
- isolates in Belgian hospitals. Int J Antimicrob Agents 39:168-172.
- 482 18. Liapis E, Pantel A, Robert J, Nicolas-Chanoine MH, Cavalié L, van der
- 483 Mee-Marquet N, de Champs C, Aissa N, Eloy C, Blanc V, Guyeux C, Hocquet D,
- 484 Lavigne JP, Bertrand X; ONERBA. 2014. Molecular epidemiology of OXA-48-
- 485 producing *Klebsiella pneumoniae* in France. Clin Microbiol Infect **20**: O1121-O1123.
- 486 19. **Pfeifer Y, Schlatterer K, Engelmann E, et al.** 2012. Emergence of OXA-48-type
- 487 carbapenemase-producing *Enterobacteriaceae* in German hospitals. Antimicrob Agents
- 488 Chemother **56**: 2125-2128.
- 489 20. Thomas CP, Moore LS, Elamin N, Doumith M, Zhang J, Maharjan S, Warner
- 490 M, Perry C, Turton JF, Johnstone C, Jepson A, Duncan ND, Holmes AH,
- 491 Livermore DM, Woodford N. 2013. Early (2008-2010) hospital outbreak of *Klebsiella*
- 492 pneumoniae producing OXA-48 carbapenemase in the UK. Int J Antimicrob Agents 42:
- 493 531-536.
- 494 21. Lascols C, Peirano G, Hackel M, Laupland KB, Pitout JD. 2013. Surveillance
- 495 and molecular epidemiology of Klebsiella pneumoniae isolates that produce
- 496 carbapenemases: first report of OXA-48-like enzymes in North America. Antimicrob
- 497 Agents Chemother **57**:130-136.
- 498 22. Ruiz-Garbajosa P, Curiao T, Tato M, Gijón D, Pintado V, Valverde A,
- 499 Baquero F, Morosini MI, Coque TM, Cantón R. 2013. Multiclonal dispersal of KPC
- genes following the emergence of non-ST258 KPC-producing Klebsiella pneumoniae
- clones in Madrid, Spain. J Antimicrob Chemother **68**: 2487-2492.
- 502 23. Giani T, Pini B, Arena F, Conte V, Bracco S, Migliavacca R; AMCLI-CRE
- 503 Survey Participants, Pantosti A, Pagani L, Luzzaro F, Rossolini GM. 2013.
- 504 Epidemic diffusion of KPC carbapenemase-producing *Klebsiella pneumoniae* in Italy:
- results of the first countrywide survey, 15 May to 30 June 2011. Euro Surveill **18(22)**.
- 506 pii: 20489.

- 507 24. Potron A, Poirel L, Rondinaud E, Nordmann P. 2013. Intercontinental spread of
- 508 OXA-48 beta-lactamase-producing *Enterobacteriaceae* over a 11-year period, 2001 to
- 509 2011. Euro Surveill **18(31)**. pii: 20549.
- 510 25. Tórtola MT, Lavilla S, Miró E, González JJ, Larrosa N, Sabaté M, Navarro F,
- 511 Prats G. 2005. First detection of a carbapenem-hydrolyzing metalloenzyme in two
- enterobacteriaceae isolates in Spain. Antimicrob Agents Chemother **49**: 3492-3494.
- 513 26. Pitart C, Solé M, Roca I, Fàbrega A, Vila J, Marco F. 2011. First outbreak of a
- 514 plasmid-mediated carbapenem-hydrolyzing OXA-48 beta-lactamase in Klebsiella
- 515 pneumoniae in Spain. Antimicrob Agents Chemother **55**: 4398-4401.
- 516 27. Woodford N, Turton JF, Livermore DM. 2011. Multiresistant Gram-negative
- 517 bacteria: the role of high-risk clones in the dissemination of antibiotic resistance. FEMS
- 518 Microbiol Rev **35**: 736-755.
- 519 28. Andrade LN, Vitali L, Gaspar GG, Bellissimo-Rodrigues F, Martinez R, Darini
- 520 **AL.** 2014. Expansion and evolution of a virulent, extensively drug-resistant (polymyxin
- 521 B-resistant), QnrS1-, CTX-M-2-, and KPC-2-producing Klebsiella pneumoniae ST11
- international high-risk clone. J Clin Microbiol **52**: 2530-2535.
- 523 29. Oteo J, Diestra K, Juan C, Bautista V, Novais A, Pérez-Vázquez M, Moyá B,
- 524 Miró E, Coque TM, Oliver A, Cantón R, Navarro F, Campos J; Spanish Network
- in Infectious Pathology Project (REIPI). 2009. Extended-spectrum beta-lactamase-
- 526 producing Escherichia coli in Spain belong to a large variety of multilocus sequence
- 527 typing types, including ST10 complex/A, ST23 complex/A and ST131/B2. Int J
- 528 Antimicrob Agents **34**: 173-176.
- 529 30. Morris D, McGarry E, Cotter M, Passet V, Lynch M, Ludden C, Hannan MM,
- 530 Brisse S, Cormican M. 2012. Detection of OXA-48 carbapenemase in the pandemic
- clone Escherichia coli O25b:H4-ST131 in the course of investigation of an outbreak of
- 532 OXA-48-producing Klebsiella pneumoniae. Antimicrob Agents Chemother 56: 4030-
- 533 4031.

- 31. Peirano G, Schreckenberger PC, Pitout JD. 2011. Characteristics of NDM-1-
- producing Escherichia coli isolates that belong to the successful and virulent clone
- 536 ST131. Antimicrob Agents Chemother 55: 2986-2968.

## TABLE 1. Distribution of carbapenemase-producing Enterobacteriaceae.

**CBP**<sup>a</sup> CBP KPC Species All OXA-48 VIM **IMP** group (%) isolates positive group (%) negative group (%) group (%) (%)(%) (%) K. pneumoniae 374 (53.3) 92 (28.5) 282 235 (86.7) 43 (44.8) 3 (37.5) 1(16.7) (74.4)E. cloacae 111 (15.8) 72 (22.3) 39 (10.3) 5 (1.8) 29 (30.2) 1 (12.5) 4 (66.6) E. coli 97 (13.8) 65 (20.1) 32 (8.4) 25 (9.2) 7 (7.3) 16 (2.3) 11 (2.9) 1 (0.4) 10\* (10.4) 2\*(25) 0 K. oxytoca 5 (1.5) C. freundii 7 (2.2) 2 (0.7) 0 14(2) 7 (1.8) 3 (3.1) 2 (25) S. marcescens 13 (1.8) 9 (2.8) 4 (1.1) 3 (1.1) 1(1) 0 0 0 E. aerogenes 34 (4.8) 32 (9.9) 2 (0.5) 0 2 (2.1) 0 M. morgannii 21 (3) 20 (6.2) 1 (0.3) 0 1(1) 0 0 0 0 1(16.7) Enterobacter 5 (0.7) 4 (1.2) 1 (0.3) 0 spp. Other 0 0 0 17 (2.4) 17 (5.3) 0 0 Total 702 (100) 323 (100) 379 (100) 271 (100) 96 (100) 8 (100) 6 (100)

540

542

543

538

<sup>&</sup>lt;sup>a</sup>CBP, carbapenemases

<sup>\*</sup>Two isolates producing both KPC and VIM carbapenemases.

TABLE 2. Comparison of OXA-48-group- and VIM-group-producing isolates.

Variable	OXA-48-group (%)	VIM-group (%)	Odds ratio	95% CI	P value	
Age > 65 years	187 (69)	32 (47.4)	2.47	1.54-3.97	0.0002	
K. pneumoniae	235 (86.7)	43 (44.8)	8.05	4.72-13.72	< 0.0001	
ST11	79 (33.6)	4 (9.3)	4.94	1.70-14.31	< 0.0001	
ST405	73 (31.1)	0	39.35	3.39- 648.40	< 0.0001	
ST15	22 (9.4)	15 (34.9)	0.19	0.089-0.41	< 0.0001	
ST326	22 (9.4)	1 (2.3)	4.34	0.57-33.08	0.22	
E. coli	25 (9.2)	7 (7.2)	1.31	0.55-3.13	0.67	
E. cloacae	5 (1.8)	29 (29.9)	0.044	0.016-0.12	< 0.000	
K. oxytoca	1 (0.4)	10 (10.3)	0.032	0.004-0.25	< 0.000	

551 TABLE 3. Susceptibility to antibiotics in carbapenemase-producing Enterobacteriaceae 552 isolates.

	Susceptibility (%)				
Antibiotic	Total of	OXA-48-group- producing isolates	VIM-group- producing	P value	
Allubouc	isolates				
	(n=379)	(n=270)	isolates (n=97)		
Colistin	95.5	95.2	95.9	1	
Amikacin	81.3	84.8	73.2	0.014	
Meropenem	74.7	80	63.9	0.002	
Tigecycline	71	72.6	67	0.30	
Imipenem	67.6	74.8	49.5	< 0.0001	
Fosfomycin	48	44.8	57.7	0.03	
Chloramphenicol	39.6	46.7	23.7	< 0.0001	
Gentamicin	33.2	37.4	22.7	0.008	
Aztreonam	20.1	12.2	40.2	0.0001	
Tobramycin	16.4	20.7	5.2	0.0002	
Trimethoprim/sulfamethoxazole	16.1	13.7	18.6	0.25	
Ciprofloxacin	12.7	9.3	23.7	0.0007	
Ceftazidime	9.5	13.3	0	< 0.0001	
Cefotaxime	7.7	10.8	0	0.0001	
Ertapenem	7.1	4.1	16.5	0.0002	

Figure legends FIGURE 1. Geographic distribution of carbapenemase types detected during a prospective multicentre study in Spain (February–May 2013). FIGURE 2. Population snapshot of sequence types (STs) of Klebsiella pneumoniae isolates from the multilocus sequence typing (MLST) database and STs carbapenemase-producing *K. pneumoniae* from this study considered altogether. Footnote: The most important ST complexes found in this study (ST11, ST405, ST15, and ST 326) are emphasised. Some STs with single isolates from the MLST database have been excluded from the image for easy viewing.