

This is the peer reviewed version of the following article:

Bar, C., Chatterjee, S., Pires, I. F., Rodrigues, P., Sluijter, J. P. G., Boon, R. A., . . . Thum, T. (2020). Non-coding RNAs: update on mechanisms and therapeutic targets from the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart. *Cardiovascular Research*, 116(11), 1805-1819.
doi:10.1093/cvr/cvaa195

which has been published in final form at: <https://doi.org/10.1093/cvr/cvaa195>

Non-coding RNAs - Update on mechanisms and therapeutic targets from the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart

AUTHOR LIST

Christian Bär^{1,2}, Shambhabi Chatterjee^{1,2}, Inês Falcão Pires³, Patrícia Rodrigues³, Joost P.G. Sluijter⁴, Reinier A. Boon^{5,6,7}, Rosa M. Nevado^{8,9}, Vicente Andrés^{8,9}, Marida Sansonetti^{1,2,10,11}, Leon de Windt^{10,11}, Michele Ciccarelli¹², Nazha Hamdani^{13,14}, Stephane Heymans¹⁵, Raquel Figuinha Videira^{3,10,11}, Carlo G. Tocchetti¹⁶, Mauro Giacca^{17,18,19}, Serena Zacchigna^{17,19}, Stefan Engelhardt^{20,21}, Stefanie Dimmeler²², Rosalinda Madonna^{23,24} and Thomas Thum^{1,2}

AFFILIATIONS

¹ Institute for Molecular and Translational Therapeutic Strategies (IMTTS), Hannover Medical School, Hannover, Germany.

² REBIRTH Center for Translational Regenerative Medicine, Hannover Medical School, Hannover, Germany.

³ Cardiovascular Research and Development Center, Faculty of Medicine of the University of Porto, Porto, Portugal.

⁴ Experimental Cardiology Laboratory, UMC Utrecht Regenerative Medicine Center, University Medical Center Utrecht, University Utrecht, Utrecht, The Netherlands.

⁵ Amsterdam UMC, Vrije Universiteit Amsterdam, Department of Physiology, Amsterdam Cardiovascular Sciences, Amsterdam, Netherlands.

⁶ Institute for Cardiovascular Regeneration, Centre for Molecular Medicine, Goethe University, Frankfurt am Main, Germany.

⁷ Partner site Rhein/Main, German Center for Cardiovascular Research (DZHK), Frankfurt am Main, Germany.

⁸ Centro Nacional de Investigaciones Cardiovasculares (CNIC), Madrid, Spain.

⁹ Centro de Investigación Biomédica en Red de Enfermedades Cardiovasculares (CIBERCV), Spain.

¹⁰ Department of Molecular Genetics, Faculty of Science and Engineering; Maastricht University, Maastricht, The Netherlands.

¹¹ CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences; Maastricht University, Maastricht, The Netherlands.

¹² Department of Medicine, Surgery and Dentistry; University of Salerno, Italy.

¹³ Department of Molecular and Experimental Cardiology, Ruhr University Bochum, Bochum, Germany.

- ¹⁴ Department of Cardiology, St. Josef-Hospital, Ruhr University Bochum, Bochum, Germany.
- ¹⁵ Department of Cardiology, Maastricht University Medical Centre, Center for Heart Failure Research, Cardiovascular Research Institute Maastricht (CARIM), University Hospital Maastricht, the Netherlands.
- ¹⁶ Department of Translational Medical Sciences and Interdepartmental Center of Clinical and Translational Research (CIRCET), Federico II University, Naples, Italy.
- ¹⁷ International Centre for Genetic Engineering and Biotechnology (ICGEB), Trieste, Italy.
- ¹⁸ School of Cardiovascular Medicine & Sciences, King's College London, London, UK.
- ¹⁹ Department of Medicine, Surgery and Health Sciences, University of Trieste, Italy.
- ²⁰ Institute of Pharmacology and Toxicology, Technische Universität München, Biedersteiner Str. 29, Munich 80802, Germany.
- ²¹ DZHK (German Center for Cardiovascular Research), Partner Site Munich Heart Alliance, Biedersteiner Str. 29, Munich 80802, Germany.
- ²² Institute for Cardiovascular Regeneration, Goethe University, Germany; German Center for Cardiovascular Research (DZHK), Frankfurt, Germany; Cardio-Pulmonary Institute (CPI), Frankfurt, Germany.
- ²³ Institute of Cardiology, University of Pisa, Pisa, Italy.
- ²⁴ Department of Internal Medicine, University of Texas Medical School in Houston, TX, USA.

Correspondence:

Thomas Thum
Hannover Medical School
Institute of Molecular and Translational Therapeutic Strategies
Carl-Neuberg-Str. 1
D-30625 Hannover
Phone: +49 511 532 5272
Fax: +49 511 532 5274
e-mail: Thum.Thomas@mh-hannover.de

Keywords: cardiovascular disease, non-coding RNA, microRNA, remodelling, regeneration, aging, non-coding RNA therapy

Abstract

Vast parts of mammalian genomes are actively transcribed, predominantly giving rise to non-coding RNA transcripts including microRNAs, long non-coding RNA and circular RNAs amongst others. Contrary to previous opinions that most of these RNA are non-functional molecules, they are now recognised as critical regulators of many physiological and pathological processes including those of the cardiovascular system. The discovery of functional non-coding RNAs has opened up new research avenues aiming at understanding non-coding RNA-related disease mechanisms as well as exploiting them as novel therapeutics in cardiovascular therapy. In this review we give an update on the current progress in non-coding RNA research, particularly focussing on cardiovascular physiological and disease processes, which are subject of current investigation at the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart. This includes a range of topics such as extracellular vesicle-mediated communication, neurohormonal regulation, inflammation, cardiac remodelling, cardio-oncology as well as cardiac development and regeneration, collectively highlighting the wide-spread involvement and importance of non-coding RNAs in the cardiovascular system.

Introduction

The completion of the human genome project in 2003 had spurred great hope to identify underlying mechanisms and to find cures for pandemic diseases such as cancer and cardiovascular disease (CVD). Unfortunately, even 20 years later, CVDs are still on the rise accompanied by lack of effective therapeutic options. This is partially because most of the drug development has been focussed on protein-coding genes. Considering that the human phenotype is not only dictated by protein-coding genes, but also by genes which give rise to non-coding RNA (ncRNA) transcripts ¹, recently more studies have been directed towards detailed investigation of such ncRNA molecules for CVD diagnostic and treatment options ^{2,3}. The class of ncRNA molecules which make up to 98% of the human transcriptome comprises a wide variety of transcripts with numerous functions. MicroRNAs (miRs), long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs) represent the major classes of ncRNAs involved in cardiovascular development and pathology (Table 1). The short miRs (~20nt) mainly interact with the 3' untranslated region of the target mRNA, thereby suppressing protein translation through RNA-interference mechanisms ⁴. The lncRNAs comprise a large group of extremely diverse molecules which are defined as ncRNA >200nt in length and are known to regulate gene expression both at the nuclear and cytoplasmic level ⁵. They can influence gene expression and cellular function of all cardiac cell types playing pivotal roles in CVDs ⁶. The more recently discovered circRNAs are covalently closed RNA rings formed through alternative back-splicing of protein-coding exons. Their function may range from host gene regulation to scaffold and molecular sponge function ⁷. CircRNAs have also been implicated in the regulation of several cellular and pathological functions in the heart ⁸. While the biogenesis and the principle functional mechanisms of the different types of ncRNA have been reviewed in detail recently ^{7,9,10}, in here, we will mainly highlight crucial roles of ncRNAs in cardiac development and disease progression. The importance of ncRNA in this field is reflected by the significant and increasing research efforts by members of the Working groups on *Myocardial Function* and *Cellular Biology of the Heart* of the European Society of Cardiology. As a follow up of the 2019 meeting in Naples we will here provide an overview of the state-of-the-art of cardiovascular ncRNA research of both ESC-Working groups focusing on different aspects of heart development, cardiac disease as well as regeneration (Figure 1). Importantly, since ncRNAs have made the first steps from basic science into clinical application we will provide an outlook on the therapeutic perspective of ncRNAs.

Table 1: The major types of ncRNA and their modes of action.

ncRNA type	Mechanisms of action	Ref
miRNA	<ul style="list-style-type: none"> • silences gene by RNA interference • suppresses protein translation by destabilizing target mRNA 	11,12
lncRNA	<ul style="list-style-type: none"> • regulates chromatin modification by remodelling complexes • sponges miRNAs and/or proteins • acts as protein decoy • provides scaffold function for protein complexes • transcriptional enhancer for target genes 	6,13
circRNA	<ul style="list-style-type: none"> • regulates host gene expression • sponges miRNAs and/or proteins • acts as protein decoy • provides scaffold function for protein complexes • encodes for micro-, mini-peptides 	7,8

1. Circular RNA: New kid on the block with specific challenges

CircRNAs comprise more recently described ncRNAs which differ from linear RNAs as they are covalently closed, do not possess strand polarities and are generated in a process termed as back-splicing in which a downstream sequence is spliced to an upstream one ¹⁴. Although we are just beginning to unravel the molecular mechanisms of circRNAs, several studies have addressed the expression in the cardiovascular system and have assigned functions to some circRNAs. In the vascular system, cZNF292 was first identified among the >7.000 circRNAs in endothelial cells (ECs) to control endothelial EC and angiogenic sprouting *in vitro* ¹⁵. Various subsequent studies documented that several circRNAs affect vascular cell functions *in vitro* (for a comprehensive overview see ^{8,16}). A prominent example is the circular form of the non-coding RNA ANRIL, circANRIL, which is induced by coronary artery disease and regulates smooth muscle cells (SMCs), specifically cell death and proliferation, by interfering with ribosomal RNA maturation ¹⁷. This effect is mediated by binding to the pre-ribosomal assembly factor PES1. More recently, a circular transcript lipoprotein receptor 6, circLrp6, was identified as a crucial regulator of vascular SMCs by sponging of and counterbalancing miR-145 ¹⁸.

Further circRNAs were identified to control cardiac functions (for a comprehensive review see ¹⁴). Interesting examples include circFoxo3a, which aggravated doxorubicin-induced cardiomyopathy. CircFoxo3a primarily affected senescence, possibly by interacting with the senescence inhibitory protein and the transcription factors E2F1 and HIF1 α ¹⁹. The highly abundant circSlc8a1 contributes to pressure overload-induced hypertrophy by sponging miR-133 ²⁰. A recent study further elegantly demonstrates that circFndc3c modulates cardiac repair after myocardial infarction. Interestingly, the authors report a circRNA-protein interaction with the RNA binding protein “fused in sarcoma” (FUS), which affects VEGF-A expression and subsequent cardiomyocyte (CM)-EC cross-talks ²¹.

As mentioned above, circRNA research is still in its infancy and comes with specific challenges for this somewhat peculiar type of RNA. While overexpression and RNA silencing mediated deletion of circRNAs demonstrated functional roles in the cardiovascular system, the field still suffers from the lack of definitive genetic evidence confirming the proposed functions *in vivo* ^{8,16}. This limitation is predominantly due to the fact that it is challenging to specifically interfere with the circularization without affecting the expression or the splicing of the host gene, which in most cases has also important functions as coding RNA. The diagnostic use and therapeutic targeting of circRNA is also more challenging as compared to miRs. The diagnostic potential is limited due to the fact that circRNAs are expressed at low levels and often come in various spliced isoforms, which makes their specific detection in the blood very challenging. The therapeutic application of circRNAs requires the overexpression of transcripts, which can be done using viral vectors containing flanking regions facilitating circularization ⁸. While this strategy was successfully used by many investigators, it is often neglected that the circularization is quite ineffective, leading to the co-expression of linear parts of the gene, which may have effects as well. Alternatively, recombinant circles can be generated, but their up-take is limited without delivery vehicles such as liposomes or nanoparticles. It is unclear if the major achievements in siRNA delivery strategies by linking them to molecules that target the siRNA to specific cell types (e.g. GalNAc for liver cells ²²) can be used as well for circRNAs. Inhibition of circRNA expression is also hampered by the fact that silencing strategies to specifically

target the circRNA are limited to the backsplice site, thereby reducing the flexibility of RNA sequences that can be used. Finally, the molecular mechanism of action is not always very compelling. Many studies reported that circRNAs act via sponging of miRs. However, given that miRs are expressed in much higher copy numbers, this mechanism of action depends on relatively high expression of the circRNA and/or the existence of many microRNA binding sites in the circRNA. Since a cross-talk between lowly expressed circRNAs with highly expressed miRs has been experimentally validated in various cases, one may need to find an alternative explanation how circRNA may interfere with the processing or localization of miRNAs or consider alternative interactions in RNA networks.

2. Non-coding RNAs in extracellular vesicle-mediated communication in the heart

In the past decade, extracellular vesicles (EVs) have come to light as novel elements of cell-to-cell communication in the cardiovascular system, not only constitutively released from many cardiac cell types, including CMs, fibroblasts (FBs), ECs, inflammatory cells, and resident stem cells but also detected in most body fluids ²³. This EV-mediated way of intercellular communication is critical in physiological and pathological cardiovascular circumstances by allowing the exchange of biological information and therefore, the coordination of cell/organ and maintenance of homeostasis ²⁴⁻²⁶.

Incorporation of miRNAs in vesicles is not random as they are selectively exported to EVs at constant ratios, varying under specific pathophysiological conditions ²⁷. Following delivery, miRNA-enriched EVs can exert functional roles in recipient cells, orchestrating their entire gene programs and affecting their phenotype. Recent developments in the field of intercellular cross-talk, demonstrate that EVs enriched in specific miRNAs could be key players during different cardiac disorders ^{26,28}.

The release of EVs, including exosomes, is a common way of communication between different cardiac cells such as CMs and ECs ²⁹. Halkein et al. demonstrated that during peripartum cardiomyopathy, miR-146a-enriched EVs released by ECs are taken up by CMs and, by interfering with their physiological metabolism, affect contractility and lead to CM hypertrophy ²⁹. Reciprocally, increased levels of miR-143 and miR-222 in EVs released by ischemic CMs exert a pro-angiogenic effect on recipient ECs ³⁰. Moreover, CMs from diabetic rat hearts were shown to release EVs enriched in miR-320 which, once delivered to cardiac ECs, compromised their proliferation, migration and tube formation capacity, leading to impaired angiogenesis ³¹. In a recent study, CM-derived EVs were reported to increase cardiac angiogenesis and CM survival as a result of EV-mediated miR-21-5p transfer ³², and CM autophagy through the uptake of miR-30a ³³.

Hergenreider et al. demonstrated that ECs can also use EVs to transfer miR-143/145 to SMC cells and reduce atherosclerotic lesion formation ³⁴. Under atherosclerotic conditions, enrichment of miR-155 in endothelial EVs were responsible for modulating the phenotype of recipient monocytes and/or macrophages *in vivo* and *in vitro* ³⁵.

Extremely relevant is the cross-talk among ECs; indeed these cells seem to be particularly enriched of miR-214 that once released, it targets other recipient ECs, thereby stimulating angiogenesis ³⁶. In accordance with this, Balkom et al. proved miR-214 to be released from ECs

contributes to EV-mediated angiogenesis and migration in neighbouring recipient cells ³⁷. Among the different cardiac cells, also cardiac FB-derived EVs have been shown to play a relevant role in many cardiovascular diseases. In mice subjected to cardiac pressure overload, miR-21-3p (miR-21*) is upregulated and transferred from cardiac FBs to CMs through EVs, inducing hypertrophy in the recipient cells ³⁸. EV-mediated cross-talk was also reported between the major cardiac cell-types and cardiosphere-derived cells ³⁹, mesenchymal stem cells ^{40,41}, and pluripotent stem cells (induced and embryonic) ^{42,43}.

Although less intensively studied to date, lncRNAs and circRNAs were also identified in cardiac EV-mediated cell-to-cell communication ⁴⁴. For example, RNA-enriched EVs secreted by hypoxic CMs were demonstrated to drive cardiac fibrosis ⁴⁵. Nevertheless, further investigations are necessary to understand the molecular mechanisms which elicit and are driven by EV-mediated intercellular communication.

3. Reciprocal regulation of non-coding RNAs and the neurohormonal system

The neurohormonal system is a pivotal contributor to organ homeostasis, as well as responsible for the adaptive mechanisms observed in chronic conditions such as HF ⁴⁶. In turn, ncRNAs modulate cellular phenotypes through the regulation of gene expression at the transcriptional and translational level ⁶. Therefore, it may be plausible that these systems are tightly interconnected with reciprocal influence in terms of receptors expression and signal transduction. These connections are particularly evident in the pathophysiological condition such as HF and hypertension, where the dysregulation of the neurohormonal system is involved in the development and progression of the disease.

In HF, the beta-adrenergic receptor stimulation can regulate miRNA expression in the animal model, and thus, mediate the effects on cardiac remodelling. For example, miRNA-214 is upregulated following chronic isoproterenol stimulation in rat and promotes cardiac FB proliferation as well collagen production and fibrosis by regulating the target gene *Mfn2* and its downstream ERK1/2 signalling pathway ⁴⁷. Moreover, miRNAs can directly interfere with the adrenergic signalling by modulating receptor expression and as well as components of its intracellular pathway ⁴⁸. For example, miR-133 can directly target the 3' UTR of the β 1-adrenergic receptor (β 1AR) and its downstream effectors, thereby limiting the cAMP production and its deleterious effects in the presence of adrenergic overdrive ⁴⁹.

This picture, however, is more complicated when considering that multiple microRNAs can impinge the expression of molecules belonging to the neurohormonal signalling. MiR-155 was found to target the 3'UTRs of the angiotensin II type I receptor (AGTR1) ⁵⁰. MiR-125a/b downregulates the expression of endothelin 1 in vascular ECs ⁵¹. MiR-766 downregulates the expression of the aldosterone synthase gene, *CYP11B2* ⁵². Beneficial seems to be miR-425 which reduces the expression of atrial natriuretic peptide (NPPA) ⁵³, while miR-100 negatively regulates expression of the natriuretic peptide receptor 3 (NPR3), the clearance receptor for natriuretic peptides, in cardiac derived cells ⁵⁴.

In summary, miRs interfere with the expression of both hormones and their cognate receptors and, vice versa, hormones seem to control the expression of miRs, highlighting that the regulation of blood pressure level depends upon the dynamic interactions between those factors.

It is likely that lncRNAs and circRNAs play important roles in neurohormonal regulation, however, this topic requires further research.

4. Non-coding RNAs in inflammatory responses

Metabolic syndrome represents a cluster of cardiovascular risk factors, including hypertension, insulin resistance, hyperlipidemia, and obesity that are associated with increased risk of HF. These comorbidities are characterized by chronic inflammation⁵⁵. Inflammation is not only critical for the development and progression of HF, but the inflammatory response is also important for adverse remodelling processes following myocardial infarction.

Inflammation and oxidative stress are major sources of both endogenous, e.g., sterile inflammation⁵⁶ and exogenous challenges that promote HF phenotypes. There is a physiological interaction that links inflammatory and oxidative stress processes, to the activation of downstream networks that promote the physiological characteristics of various human pathologies, including aging, carcinogenesis, neurodegenerative disorders and HF associated with various causes and phenotypes⁵⁷⁻⁵⁹.

In view of the manifold ncRNA mechanisms in the regulation of cardiovascular inflammation (recently reviewed by others⁶⁰), we will focus here on the implication of ncRNAs as immune regulators of the susceptibility to myocarditis upon cardiac viral infection. Human and experimental miR expression studies reveal a strong association between miR dysregulation and human myocarditis and suggest novel miRNA therapeutic targets⁶¹. Indeed, inhibition of miR-155⁶²⁻⁶⁴, -21 and -146b⁶⁵ by systemically delivered anti-miRs reduces cardiac inflammation and damage in CVB3- or auto-immune myocarditis in mice. Cardiac overexpression of miR-590-3p also prevents cardiac injury and dysfunction by inhibiting p50 expression, suppressing NF- κ B activity and blocking IL-6/TNF- α expression⁶⁶. MiRs may also modulate the virulence of cardiotrophic viruses⁶⁷. The miR-221/222 cluster in CMs regulates both virulence and inflammatory pathways in the heart⁶⁸. Systemic inhibition of miR-221/-222 in mice increases cardiac viral load, prolongs the viremic state, and aggravates cardiac inflammation and injury. Mechanistically, miR-221/-222 targets the expression of proteins that orchestrate viral replication and inflammation, including ETS1/2, IRF2, BCL2L11, TOX, BMF, and CXCL12. Similarly, miRNA-155 inhibits PU.1 and SOCS1 in the heart and as such de-represses the production of pro-inflammatory cytokines, enhancing T-cell and monocyte activation^{68,69}. Together, these results support the concept that a single miR or miR clusters may orchestrate immune activation and modulate myocarditis.

Very limited knowledge exists on the contribution of lncRNAs to viral myocarditis. A very recent report revealed that the loss of lncRNA AK085865 in macrophages promotes the polarization to M1 phenotype at the detriment of M2 macrophage levels, which increases the susceptibility of mice to coxsackievirus B3-induced viral myocarditis⁷⁰. In addition to viral myocarditis, a number of lncRNAs emerged as mediators of cardiac inflammation of other origin. The three best-studied lncRNAs are MALAT1 (metastasis-associated lung adenocarcinoma transcript, ANRIL (antisense RNA in the INK4 locus) and HOTAIR (HOX transcript antisense RNA). MALAT1 regulates T-cell and macrophage activation^{71,72}. Knockdown of MALAT1 in a rat model of systemic inflammation protected against cardiac dysfunction in part by decreasing cardiac NF- κ B protein levels, and also by decreased

circulating TNF- α and IL-6⁷³. In diabetic mice, the absence of MALAT-1 decreased inflammatory cytokines in the heart⁷⁴. However, MALAT-1 does not affect cardiac inflammation in pressure overloaded hearts, suggesting that the pro-inflammatory property of MALAT1 is dependent on the stimulus⁷⁵. LncRNA ANRIL is expressed from the ANRIL locus, a major hotspot for disease associated mutations (including coronary artery disease)⁷⁶. *In vitro*, ANRIL, as for MALAT-1 and ageing-related lncRNA HOTAIR⁷⁷, is a pro-inflammatory lncRNA. It is induced by TNF- α via the NF- κ B pathway⁷⁸. ANRIL itself increases IL-6, the cell adhesion molecules ICAM-1 (intercellular adhesion molecule-1) and VCAM-1 (vascular cell adhesion molecule-1)⁷⁹. ANRIL, in particular circular ANRIL, induces apoptosis of human ECs^{79,80}, all together suggesting a pro-inflammatory role for ANRIL mainly in the vasculature.

5. Non-coding RNAs in the course of left ventricular remodelling and reverse remodelling

Cardiac remodelling is a complex process that introduced molecular, cellular and interstitial changes leading to changes in size, mass, geometry and function of the heart in response to pathological stimuli (e.g. high blood pressure, aortic stenosis, MI)⁸¹. The literature focusing on the role of ncRNAs in pathological LV remodelling as well as their therapeutic use capable of preventing involved adverse processes (hypertrophy and fibrosis) is vast and has been adequately reviewed previously^{2,6,82–84}. However, limited research has been published describing which ncRNAs correlate to the extent of reverse remodelling (RR).

RR refers to any alteration in cardiac disease or HF that can be chronically reversed by a given therapeutic approach (pharmacological or surgical)⁸⁵. Importantly, it represents a surrogate parameter for patient prognosis^{86,87}. For instance, severe aortic valve stenosis triggers cardiac remodelling characterized by left ventricular concentric hypertrophy associated with diastolic dysfunction, while aortic valve replacement elicits RR by reducing hypertrophy and improving function. Other examples of RR include the expected recovery of cardiac function and structure after acute MI or in HF patients after cardiac resynchronisation therapy (CRT) or left ventricle assist devices (LVAD) implantation. Incomplete RR is associated with poor prognosis, thus, identification of biomarkers of RR progression or altered signalling pathways to reverse deleterious remodelling stands as a promising target. A recent study by Shah identified a cluster of miRs that provided higher discrimination than a clinical model to predict RR⁸⁸.

It is tempting to assume that ncRNAs dysregulated in cardiac remodelling are most likely the ones that will normalize during RR. In this context, a study carried out in a rat model of heterotopic transplantation following abdominal aortic constriction, developed to mimic cardiac remodelling and RR, respectively, showed that the expression levels of 7 miRs (miR-347, -483, -326, -212, -130b, -29a and -23a) were significantly altered in hypertrophic samples but normalized in unloaded heart samples⁸⁹. However, and most importantly, these authors indicated a subset of miRs whose expression only changed in unloaded hearts, e.g. miR-125a, miR-143, miR-382 and let7 family, suggesting its exclusive role during RR⁸⁹ (Figure 2). In LVAD patients, Akat et al. reported that relative miRNA abundance changes in myocardial tissue could not be detected within the pool of circulating miRNAs but that the former could still serve as excellent biomarkers of heart muscle injury⁹⁰. Nevertheless, miR-208a, miR-208b

and miR-499 in the circulation closely follow its myocardial expression, which is consistent with its cardiac production⁹⁰. Interestingly, these same miRs were later shown to serve as novel biomarkers for monitoring and forecasting postoperative myocardial injury and recovery after cardiac surgery in children⁹¹. MiR-132 is upregulated during adverse cardiac remodelling and directly contributes to the pathological processes. Importantly, pharmacological blockade of miR-132 to normalize its levels promotes functional recovery and RR in small and large HF animal models^{92,93}.

In CRT patients, the beneficial effects of CRT on RR of HF patients were associated with modulation of circulating miR patterns implicated in cardiac hypertrophy, fibrosis, and apoptosis⁹⁴ and vary significantly in responders versus non-responders (patients that do not undergo RR) as depicted in Table 2. In addition, baseline plasma miR-30d levels were shown to be associated with a beneficial RR and to induced CM growth and protection against apoptosis in vitro⁹⁵.

In RR induced by aortic valve replacement, regression of hypertrophy 1 year after the intervention correlated to the profile of myocardial gene expression at the time of surgery (e.g. anti-hypertrophic miR-133a, β -myosin heavy chain, myosin light chain-2 and other genes) in conjunction with patients' clinical background (age, BMI, diabetes mellitus and male gender). These constitute crucial determinants of RR, which were stronger predictors of cardiac mass reduction than postoperative improvement of valve haemodynamics⁹⁶.

In idiopathic dilated cardiomyopathy patients, anti-hypertrophic miR-1, miR-199, the profibrotic miR-21-5p, the cardioprotective miR-494-3p, the anti-proliferative miR-591 and miR-208a miRs were differentially expressed according to the degree of RR and predicted the time-dependent RR in response to β -blocker treatment⁹⁷.

Matkovich has shown that cardiac miR signature is an exquisitely discriminating biomarker of the severely failing heart and of the extent of RR after cardiac unloading, greatly enhancing the predictive ability of mRNA profiles to categorise the clinical status of heart failure⁹⁸. Nevertheless, a subsequent comprehensive deep-sequencing analysis highlighted that lncRNAs had the most dynamic expression changes in response to haemodynamic unloading assessed in myocardial biopsies from ischemic and non-ischemic patients before and after LVAD implantation. This study revealed that the expression profiles of lncRNAs, but not mRNAs or miRNAs, can discriminate failing hearts of different aetiologies and are markedly altered in response to LVAD support⁹⁹. A particularly striking finding here is the high abundance of mRNAs and lncRNAs of mitochondrial origin: 13 mitochondrial mRNAs and 9 mitochondrial lncRNAs alone account for 37% and 71% of the total cardiac mRNA and lncRNA read counts, respectively⁹⁹.

Table 2: Changes in miRNAs during several types of cardiac reverse remodelling and its relationship with the extent of myocardial recovery.

Type of reverse remodelling	Changes in plasma or myocardial miRNAs levels	Implications for the extent of myocardial recovery and related signalling pathways	Ref
Left Ventricular Assist Device (LVAD)	↓ plasma levels of miRs-23a and miR-195 after LVAD	Associated with smaller CM size and a favourable ventricular RR	100
	↑ myocardial levels of miR-338-3p, miR-142-5p and -3p, miR-216a-5p, miR-223-3p, miR-27a-5p, and miR-378g	Correlation with off-pump cardiac index values. Predicted targets of these miRs were involved in focal adhesion/integrin pathway and in actin cytoskeleton regulation	101

	↓ miR-29b-3p and miR-374b-5p	Correlation with pulmonary vascular resistance values	102
	↑ myocardial levels of miR-137	Correlates with down-regulation of α -1-antichymotrypsin mRNA tissue levels in RR	103
	↑ plasma levels of miR-155	Upregulated with long-term LVAD support	104
	↑ plasma levels of miR-483-3p and ↓ miR-1202	Potential capacity to monitor and predict response to LVAD therapy. Low levels of miR-1202 are associated with better response to LVAD	105
	↑ plasma levels of miR-210 at baseline	Increases in parallel with increment in NT-proBNP levels Associated with higher mortality after 3.5 year follow-up	106
Cardiac Resynchronization Therapy (CRT)	↑ plasma levels of miR-30d at baseline	Associated with a beneficial RR and to induced CM growth and protection against apoptosis in vitro	94
	↑ plasma levels of miR-26b-5p, miR-145-5p, miR-92a-3p, miR-30e-5p and miR-29a-3p	List of miRs that distinguish responders vs non-responders patients after 1 year of CRT implantation	107
Aortic Valve Replacement (AVR)	↑ levels of miR-21 both in plasma and myocardial tissue	Correlation with mean transvalvular gradient and LV fibrosis	108
	↓ plasma levels of miR-1	Associated with LV hypertrophy (LVH) and correlated with levels of soluble heart-type fatty acid-binding protein-3 (FABP3)	109
	↑ myocardial levels of miR-29, miR-21	Independent predictors of reverse remodelling and systolic function recovery	110
	↓ plasma levels of miR-206	Correlated negatively with the left ventricular ejection fraction	111
	↑ plasma levels of miR-133a	Plasma levels reflect its myocardial expression; Positive predictor of the hypertrophy reversibility after surgery	112
	↓ plasma levels of miR-378 at baseline	Predicts LVH independent of the pressure gradient	113
Drug Therapy	↓ myocardial levels of miR-208a-3p, miR-208b-3p, miR-21-5p, miR-591 and miR-199a-5p and ↑ miR-1-3p 5 months after B-blocker treatment	Differentially expressed accordingly to the degree of RR and predicted the time-dependent RR in response to β -blocker treatment	97
Myocardial Infarction remodelling	↑ plasma levels of miR-1254	Predicted changes in LV volumes and LVEF at 6 months after STEMI	114
	↑ plasma levels of miR-208b and miR-34a	The increase levels were strongly associated with increased risk of mortality or HF within 6 months after acute myocardial infarction	115
	↑ plasma levels of miR-1 and miR-29b	Increase levels correlate with infarct volume accessed by MRI and miR-29b was also associated with left ventricular end-diastolic volumes over time	116
	↑ plasma levels of miR-30a-5p was elevated on admission day	Increased levels at the time of admission are associated development of LV dysfunction and HF symptoms 6 months after acute myocardial infarction	117

6. Non-coding RNAs in right ventricular remodelling

Maintenance of normal heart haemodynamics is dependent on right ventricle (RV) function, which is compromised in common diseases such as pulmonary hypertension (PH), congenital diseases (e.g. Tetralogy of Fallot-TOF) and cardiomyopathies (e.g. arrhythmogenic right ventricle cardiomyopathy -ARVC). Despite its importance, the molecular mechanisms underlying RV remodelling in response to stress still remain understudied.

Historically associated with low pressure pulmonary circulation, RV holds unique features compared to the left ventricle (LV), including thin walls, a crescentic shape, greater compliance to volume-overload and less ability to adapt to higher pressures¹¹⁸. Disturbances in RV homeostasis, such as increased afterload, can lead to RV maladaptive remodelling characterized by increased hypertrophy, fibrosis, increased oxygen consumption, a metabolic switch and alteration of the RV molecular signature including non-coding RNAs (ncRNAs), ultimately leading to RV failure (RVF)¹¹⁸.

A variety of miRs have been associated with PH-induced RVF¹¹⁹. MiR-223 was found downregulated in the RV of two rat models of PH, where it targets insulin-like growth factor (IGF) receptor 1 (IGF-IR)/IGF downstream signalling. De-repression of IGF-IR following ablation of miR-223 prevented maladaptive remodelling and improved RV function¹²⁰. Another study reported miR-126 as an angiogenic miRNA dysregulated during PH-induced RVF, as its levels drastically decrease in end-stage RVF possibly due to alterations in the angiogenic vascular endothelial growth factor (VEGF)/VEGF receptor-2 (VEGFR2)/ mitogen-activated protein kinase (MAPK) pathway¹²¹. *In vivo* administration of miR-126 mimics to a RVF rat model increased expression levels of the target gene sprouty related EVH1 domain containing 1 (SPRED1), a known negative regulator of the VEGF pathway, and consequently improved RV function¹²¹. MiR expression is also dysregulated in TOF and ARVC pathologies, both intrinsically involved in RVF^{122,123}.

More recently, lncRNAs have been linked to RV remodelling and dysfunction. The lncRNA H19 is downregulated in a rat model of PH-induced RV hypertrophy. The observed protective effect of melatonin treatment in this model, leading to a decrease in RV hypertrophy, were mediated through restored H19 levels and direct suppression of miR-200a by H19¹²⁴. A profiling study to identify the lncRNA signature in human RVF samples reported 78 lncRNAs to be differentially expressed when compared to healthy RVs¹²⁵. However, to date, no direct association has been reported regarding the functional role of other lncRNAs in RV function. Thus, the knowledge of ncRNAs on RV remodelling and failure is still limited. Despite promising, studies directly focused on RV circRNAs and lncRNAs remain scarce and more data needs to be generated in order better understand RV remodelling and ultimately develop better therapies.

7. Non-coding RNAs in cardio-oncology

The role of ncRNAs in cardio-oncology is still debated¹²⁶. Nevertheless, miRNA-mediated modulation of anthracycline (e.g. doxorubicin)-induced cardiomyopathy has been hypothesized in recent papers.

It has been suggested that the cardiac-specific miR-208a is upregulated in an experimental model of acute cardiomyopathy induced by anthracyclines¹²⁷. The inhibition of miR-208a was able to counteract the deleterious action of anthracyclines on cardiac function and apoptosis.

Such inhibition can also de-repress miR-208a-target Gata4 with enhanced expression of the anti-apoptotic gene Bcl2¹²⁷. miR-532-3p is another miRNA that was enhanced in CMs administered with anthracyclines¹²⁸, increasing CM susceptibility to doxorubicin by promoting mitochondrial fission¹²⁶. Interestingly, inhibiting miR-532-3p in tumor cells did not impact anthracyclines-mediated apoptosis. Hence, miR-532-3p blockade improves rescues CM death and mitochondrial fission caused by anthracyclines, with no changes in their anticancer activity¹²⁸.

Importantly, beside CMs, FBs, SMCs, and cardiac progenitor cells, ECs may also play a major role in doxorubicin-induced cardiomyopathy¹²⁹. Acute administration of anthracyclines in murine models reduced microvessel density and VEGF-A expression, while it enhanced miR-320a¹³⁰. Inhibiting of miR320a ameliorated heart function, reduced apoptosis, and enhanced microvessel density in mice administered with anthracyclines, while overexpression of miR-320a produced opposite results. Conversely, overexpression of the miR-320a target VEGF-A prevented detrimental effects of miR-320a in cardiac dysfunction induced by anthracyclines, confirming that VEGF is a downstream target molecule^{126,130}. Moreover, miR-212/132 overexpression was shown to prevent cardiac atrophy in a chronic mouse model of doxorubicin-induced cardiotoxicity¹³¹.

Particularly, circulating miRs are intensively investigated as potential blood/plasma-based biomarkers in cardio-oncology¹³². For example, circulating miR-1 levels could be enhanced upon anthracyclines administration in breast cancer patients who later developed cardiotoxicity¹³³. In a pediatric study, children with different tumors exhibited higher levels of circulating miR-29b and miR-499 post-chemotherapy¹³⁴. Furthermore, miR-29b and -499 levels were higher in subjects who showed higher hsTnT levels. Oatmen and colleagues performed miRNA profiling in childhood oncology patients and validated several miRNAs (miR-486-3p, -103-3p, -142-3p, and -92a-3p) as potential biomarkers for both acute and chronic anthracycline-induced LV dysfunction. However, larger patient cohort studies are needed for further validations assessment of the prognostic potential of serum miRNAs¹³⁵.

Also lncRNAs seem to play a role in anthracyclines cardiomyopathy. LncRNA myosin heavy chain-associated RNA transcripts (Mhrt) was found to be downregulated in doxorubicin-treated hearts¹³⁶. Its overexpression in CMs is able to inhibit doxorubicin-induced apoptosis, while its inhibition exacerbates doxorubicin effects. The expression of lncRNA cardiac hypertrophy-related factor (Chrf) was higher in mice hearts after doxorubicin treatment. Chrf inhibition counters apoptosis and transforming growth factor-1 expression induced by anthracyclines. Chrf induction was not observed upon treatment with the angiotensin II receptor blocker valsartan, while adenovirus-mediated overexpression of Chrf reverts the positive effect of valsartan against cardiomyopathy from anthracyclines in rodents¹³⁷.

Recently, also circRNAs are emerging as mediators of cardiotoxicity. The mitochondrial fission and apoptosis-related circRNA (Mfacr) is upregulated in a model of anoxia/reoxygenation. Mfacr sponges miR-632-3p which leads to de-repression of Mtp18. Consequently, inhibition of Mfacr increases and decreases miR-632-3p and Mtp18, respectively, resulting in reduced mitochondrial fission and suppressed CM apoptosis¹³⁸. Interestingly, the RNA-binding protein Quaking was demonstrated to regulate a number of circRNAs derived from cardiac-specific loci (e.g. Ttn, Fhod3 and Strn3), thereby modulating the susceptibility of CMs to doxorubicin¹³⁹.

8. Non-coding RNAs in cardiac development and cell specification

The essential role mature miRs play in cardiac development was first revealed by cardiac specific removal of the miR processing ribonuclease Dicer, which prenatally resulted in defective heart morphogenesis and embryonic lethality¹⁴⁰.

Since then, many miRs have been identified that contribute to morphogenesis, e.g. miR-1/miR-133¹⁴¹, to CM proliferation via the miR-15 family^{142–144}, and thereby mediating congenital heart disease or regeneration¹⁴⁵. Muscle-specific myomiRs are involved in essential steps for cardiac development, and therefore a fine-tuned regulation of these myomiRs, which includes miR-1, -133a, -208a, -208b, and -499, is essential for the proper embryological development of the heart¹⁴⁶. Interestingly, recent cellular reprogramming and trans-differentiation insights demonstrated that a combined presence of transcription factors Gata4, Mef2c, and Tbx5 (GMT) were able to directly differentiate cardiac fibroblasts into induced CMs (iCMs)¹⁴⁷. Subsequently, the same was established by introducing several muscle-specific miRs at the same time, including miR-1, -133, -208, and -499¹⁴⁸, of which miR-133 is suggested to be responsible for furthering mature the transdifferentiated iCMs¹⁴⁹. Interestingly, the efficiency of *in vivo* reprogramming with these approaches via e.g. viral constructs is even more successful than *in vitro*, leading to more mature CM¹⁵⁰.

In addition to miRs, also many lncRNAs were reported to have a cardiac-specific origin during organ development but also in cardiovascular disease^{151–154} but, due to their complex regulation, further studies are still needed. The first described key regulator lncRNA in cardiac development was Braveheart (Bvht)¹⁵⁵. Bvht was detected in cardiac mesoderm and CMs, and involved to move cells from nascent to cardiac mesoderm and further towards full CM differentiation. Another lncRNA example was named TERMINATOR and is essential for maintaining of pluripotency in stem cells and the subsequent early mesodermal differentiation and survival. Interestingly, mesodermal specification followed by cardiac chamber formation is mediated via ALIEN, which is mainly expressed in cardiovascular progenitor cells¹⁵⁶. In addition, the CM regeneration-related lncRNA (CRRL) was identified to be involved in cardiomyocyte proliferation and regeneration, as indicated by enhanced positivity of Ki-67/pH3/EdU in cardiomyocytes¹⁵⁷.

More recently, transcriptional regulation of circRNAs was reported during rodent and human heart development and disease¹⁵⁸. The myocardium displays a general increased expression over-time of circRNAs, especially during the first weeks of the second trimester of human fetal development¹⁵⁹. Interestingly, very recently, circRNAs from the titin gene were dysregulated upon the removal of the splicing regulator RNA binding motif protein 20 (RBM20)¹⁶⁰. Nevertheless, the functional consequences of cardiac circRNA dysregulation still awaits further investigation.

9. Regulation of cardiomyocyte proliferation and cardiac regeneration by non-coding RNA

As any aspect of cardiac biology, CM proliferation is also under the control of the ncRNA network. Multiple miR and a few lncRNAs identified to date are known to either stimulate or inhibit CM proliferation.

Starting from a systematic screening of a library of approximately 1000 human miRs¹⁴³, several miRs have been reported to stimulate CM proliferation in mice, rats, pigs and human cells, as well as to induce cardiac regeneration after MI^{100,143,161,162}. Several of the pro-proliferative miRs belong to a few miR families, including members of the miR-302/367 cluster and the miR-290 family in mice. These miRNAs share a similar seed sequence and are highly expressed during the early stages of development. In particular, they are involved in the specification and maintenance of pluripotency of embryonic stem (ES) cells¹⁶³, in which the miR-290 cluster alone accounts for 70% of the entire miR content¹⁶⁴.

Other miR families are known to be crucial in the regulation of cell proliferation in other cell types also induce CM replication. Expression of the miR-17-92 cluster¹⁶⁵ is activated in several human tumours, hence the name OncomiR1^{166,167}. Transgenic overexpression of this miR cluster in CMs¹⁶⁸ or cardiac delivery of two members of the family, miR-19a and miR-19b¹⁶⁹, induce CM proliferation in both pre- and post-natal hearts, and stimulate cardiac regeneration after MI.

Withdrawal of CMs from the cell cycle also depends on increased levels of the miR-15 family¹⁴⁴ as well as of miR-29a¹⁷⁰. Inhibition of these miRs, which target various components of the cell cycle and DNA damage response machinery, leads to improved cardiac repair after MI¹⁷¹. A common characteristic of most of the pro-proliferative miRs is the activation of the Yap transcriptional co-factor, originally discovered as the final positive effector of the otherwise inhibitory Hippo pathway in *Drosophila*^{172,173}. Indeed, a high throughput screening performed on human induced pluripotent stem cell (hiPSC)-derived CMs revealed that most of the miRs that increase CM proliferation converge on the Hippo pathway¹⁷⁴.

Not surprisingly, CM proliferation is also under the control of various lncRNAs¹⁷⁵. In several instances, these inhibit CM proliferation by acting as sponges for pro-proliferative miRNAs. This is the case of CAREL (a sponge for miR-296¹⁷⁶), CRRL (for miR-199a-3p¹⁵⁷) and AZIN2-sc (for miR-214 [50]). In other cases, the sponge effect is for inhibitory miRNAs (such NR_045363 for miR-216a¹⁷⁷), with the lncRNA then exerting a positive effect on CM proliferation. Other lncRNAs that increase CM proliferation are ECRAR, which binds to and promotes phosphorylation of ERK1/2¹⁷⁸ and Sirt1 antisense lncRNA, which stabilizes the Sirt1 mRNA¹⁷⁹. The CPR lncRNA instead blocks CM proliferation by recruiting DNMT3A to the promoter region of the MCM3 gene¹⁸⁰. While most of these lncRNAs or their inhibitors exert a positive effect after MI in rodents, the entity of this effect appears more modest compared to that exerted by miRs, likely due to pleiotropic functions of the latter molecules. It is possible that more effective lncRNAs regulating proliferation of CMs or cardiac progenitor cells will be identified in the class of enhancer-associated lncRNAs¹⁷⁵.

10. Non-coding RNA in cardiovascular aging

Aging is the main risk factor for cardiovascular disease. All cells and organs relevant for the cardiovascular system are affected by aging. Not surprisingly ncRNAs seem to also play important roles in cardiac aging, and this knowledge is derived mainly from mouse studies.

Analysis of young and old mouse hearts revealed differential expression of 65 miRNAs¹⁸¹, including changes in the expression of three microRNA clusters —miR-17-92, -106a-363, and -106b-25— that potentially target the Cdc42-SRF signalling pathway¹⁸². A longitudinal study in mice from birth to 19 months of age found that miR-22 was prominently upregulated during cardiac aging and appeared to contribute at least partly to accelerated cardiac FB senescence and migratory activity during aging¹⁸³. The most notable examples in ECs are miR-34 and miR-92. MiR-34 was first identified as a P53-responsive SIRT1-targeting miR¹⁸⁴. SIRT1 is known to counteract aging and to induce EC function¹⁸⁵. Later it was shown that miR-34 contributes to endothelial aging by inhibiting SIRT1¹⁸⁶. Several other targets for miR-34 have been identified. PNUTS, the most notable aging-regulated miR-34 target, was shown to be even more important for miR-34-induced cardiac aging than SIRT1, although the role of PNUTS in endothelial aging is currently unknown¹⁸⁷. The involvement of miR-92 in vascular aging was demonstrated in 2017 by two groups^{188,189}. Mechanistically, endothelial miR-92 levels rise with aging, resulting in loss of the antioxidant transcriptional network orchestrated by NRF2. Moreover, expression and functional studies of the miR-17-92 cluster also support a role in CM aging through the upregulation of connective tissue growth factor (CTGF) and thrombospondin-1 (TSP-1)¹⁹⁰. miR-17, another miR-17-92 cluster member, is a senescence-related miRNA that inhibits mouse cardiac FB senescence by targeting Par4¹⁹¹.

In recent years, several lncRNAs have been linked to cardiovascular aging, the most prominent examples being Meg3, H19, MIAT and HOTAIR. Meg3 was identified as an aging-induced lncRNA in ECs that inhibits angiogenesis¹⁹². Inhibition of Meg3 in aged mice restored angiogenic function. Importantly, Meg3 is also highly expressed in cardiac FBs, and silencing Meg3 in CFs prevents the induction of MMP-2, thus leading to decreased cardiac fibrosis and improved diastolic performance in a mouse model of pressure overload-induced HF. LncRNA H19 that is repressed during aging, was found to regulate EC and smooth muscle cell function. In ECs, the decline of H19 contributes to senescence and pro-inflammatory signaling¹⁹³, whereas in SMCs, H19 induction is causally related to aneurysm formation¹⁹⁴. MIAT is named myocardial infarction associated transcript because it was first discovered to be genetically associated with (cardio)vascular disease. MIAT is repressed in senescent FBs¹⁹⁵ and controls microvascular function¹⁹⁶. HOTAIR, on the other hand, is induced in senescent FBs. Several mechanisms for HOTAIR function have been proposed, depending on the cell type and disease context. For example, silencing HOTAIR inhibits senescence by regulating protein ubiquitination⁷⁷.

Little is known about the involvement of circRNAs in cardiac aging. One example is the circRNA circ-Foxo3 which is upregulated in heart samples of aged patients and mice. Mechanistic studies suggest circ-Foxo3-dependent induction of cell senescence, possibly via cytoplasmic retention of multiple anti-senescence and anti-stress factors (ID-1, E2F1, HIF1a, and FAK)¹⁹⁷.

Outlook and therapeutic perspective of non-coding RNA

As shown in the overview there are many new aspects in the mechanistic view of how ncRNAs are able to control cellular functions of cardiovascular cells. We here focused on actual topics

actively investigated within working groups of the ESC but also beyond ranging from cellular communication to regeneration to aging aspects. For instance, the discovery that CM proliferation is under the control of the microRNA network prompted the development of therapeutic strategies that take advantage of these molecules. Blocking the activity of endogenous, inhibitory miRNAs is now possible through the delivery of either antisense oligonucleotides containing locked nucleic acid (LNA)-modified nucleotides¹⁹⁸, often with a GapmeR design¹⁹⁹, or Adeno-Associated Virus (AAV) vectors expressing antisense sequences. LNAs against miR-15¹⁷¹ or miR-34a²⁰⁰ and AAV vectors expressing anti-let-7 sequences^{201,202} have all been shown to induce cardiac regeneration after MI in rodents.

Another approach is to boost the minimal regenerative capacity of the heart by the delivery of miRNA mimics, independent from their endogenous, normal expression in CMs. Delivery of both miR-199a and miR-590 in mice resulted in remarkable formation of new cardiac mass after MI, with the consequent restoration of cardiac function¹⁴³. Analogous findings were reported for AAV vectors expressing miR-294¹⁶² and miR-19a/19b¹⁶⁹.

The long-term expression of miRNAs inducing proliferation, as it occurs upon AAV-mediated gene transfer or in transgenic animals, however, can be detrimental, particularly because CM replication is accompanied by their de-differentiation. In addition, the pri-miRNA gene cloned inside AAV vectors results in the production of both miRNA strands, with possible unwanted side effects. Consistent with these concerns, infarcted pigs treated with an AAV-miR-199a vector showed remarkable regeneration at one month after treatment, however developed fatal arrhythmias at later times²⁰³. Transgenic mice overexpressing the miR-302/367 cluster also developed cardiac dysfunction due to CM de-differentiation and hyperproliferation¹⁶¹.

These problems may be overcome by the transient delivery of synthetic miRNA mimics. The intracardiac injection of miR-199a-3p, miR-590-3p²⁰⁴ or miR-19a/19b mimics¹⁶⁹, using different lipids, or of cholesterol-modified miR-302b/c mimics using a hydrogel¹⁰⁰, all led to the persistence of the miRNAs for several days after administration and stimulated cardiac repair. A regenerative effect after MI was also achieved by the daily intravenous administration of lipid formulations delivering miR302b/c¹⁶¹, miR-19a/19b¹⁶⁹ and miR-708²⁰⁵ mimics.

While proven very effective in pre-clinical models already, several obstacles such as targeted delivery, off-target effects and hepatic/renal toxicity remain to be overcome for broad clinical application of ncRNA therapeutics. For instance, the use of viral vectors based on naturally occurring serotypes of adeno associated viruses is strongly limited by the presence of neutralizing antibodies in up to 70% of the population²⁰⁶. Another example is the use of synthetic miR mimics or antisense-oligonucleotides. While systemic or local administration of such ncRNA therapeutic may be beneficial in the heart or a specific cardiac cell-type, it may cause deterioration in other organs such as kidney or liver or in non-target cardiac cell types, respectively. Thus, further efforts are urgently needed allowing for precise spatiotemporal delivery of ncRNA therapeutics.

However, a promising translational example is the clinical development of anti-miR-132 molecules in patients with heart failure. Demonstrating strong efficacy of miR-132 inhibition to treat post-MI heart failure in pigs⁹³, a clinical study was launched to test the safety of CDR132L, a synthetic miR-132 blocker, directly in heart failure patients (www.clinicaltrial.gov; NCT04045405).

Collectively, basic, translational and clinical ncRNA research conducted by members of the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart helped to pave the way for future ncRNA-based therapies. Therapeutics targeting ncRNAs have now entered the clinical setting for the treatment of cardiovascular disease. We strive to continue leading the ncRNA therapeutic field and we are following new developments with great passion and excitement.

Funding

TT and CB were supported by Deutsche Forschungsgemeinschaft, DFG (TRR267) and the ERANet CVD (JTC2016 project EXPERT to TT and JTC2018 project INNOVATION to CB). CGT was supported by a “Federico II University/Ricerca di Ateneo” grant. IFP is supported by Fundo Europeu de Desenvolvimento Regional (FEDER) through Compete 2020 – Programa Operacional Competitividade E Internacionalização (POCI), the project NETDIAMOND (POCI-01-0145-FEDER-016385), supported by European Structural And Investment Funds, Lisbon’s regional operational program 2020. PR is funded by FCT (SFRH/BD/96026/2013). RM was supported by a Cardio-Oncology grants from Incyte s.r.l. and funds from Ministero dell’Istruzione, Università e Ricerca Scientifica (549901_2020_Madonna: Ateneo). NH was supported by Deutsche Forschungsgemeinschaft, DFG (HA7512/2-1). J.P.G.S. was supported by the Project EVICARE (No. 725229) of the European Research Council (ERC). VA’s laboratory received support from the Instituto de Salud Carlos III (ISCIII) (grant AC16/00091, as member of the ERA-CVD JCT2016 EXPERT Network, European Union's Horizon 2020 Framework Programme), with co-funding from the European Regional Development Fund (ERDF/FEDER, “Una manera de hacer Europa”). RMN is supported by the Ministerio de Educación, Cultura y Deporte (predoctoral contract FPU16/05027). The CNIC is supported by the Spanish Ministerio de Ciencia e Innovación (MCI), the ISCIII, and the Pro CNIC Foundation, and is a Severo Ochoa Center of Excellence (SEV-2015-0505).

Conflicts of interest

CB and TT have filed and/or granted patents in the field of ncRNA therapeutics. TT is a founder of and holds shares in Cardior Pharmaceuticals GmbH.

References

1. Djebali S, Davis CA, Merkel A, Dobin A, Lassmann T, Mortazavi A, Tanzer A, Lagarde J, Lin W, Schlesinger F, Xue C, Marinov GK, Khatun J, Williams BA, Zaleski C, Rozowsky J, Röder M, Kokocinski F, Abdelhamid RF, Alioto T, Antoshechkin I, Baer MT, Bar NS, Batut P, Bell K, Bell I, Chakraborty S, Chen X, Chrast J, Curado J, et al. Landscape of transcription in human cells. *Nature* 2012;**489**:101–108.
2. Lu D, Thum T. RNA-based diagnostic and therapeutic strategies for cardiovascular disease. *Nat Rev Cardiol* 2019;**16**:661–674.
3. Huang C-K, Kafert-Kasting S, Thum T. Preclinical and Clinical Development of Noncoding RNA Therapeutics for Cardiovascular Disease. *Circ Res* Lippincott Williams & Wilkins Hagerstown, MD; 2020;**126**:663–678.
4. O'Brien J, Hayder H, Zayed Y, Peng C. Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulation. *Front Endocrinol (Lausanne)* Frontiers; 2018;**9**:402.
5. Yao R-W, Wang Y, Chen L-L. Cellular functions of long noncoding RNAs. *Nat Cell Biol* Nature Publishing Group; 2019;**21**:542–551.
6. Bär C, Chatterjee S, Thum T. Long Noncoding RNAs in Cardiovascular Pathology, Diagnosis, and Therapy. *Circulation* 2016;**134**:1484–1499.
7. Kristensen LS, Andersen MS, Stagsted LVW, Ebbesen KK, Hansen TB, Kjems J. The biogenesis, biology and characterization of circular RNAs. *Nat. Rev. Genet.* Nature Publishing Group; 2019. p. 675–691.
8. Santer L, Bär C, Thum T. Circular RNAs: A Novel Class of Functional RNA Molecules with a Therapeutic Perspective. *Mol Ther* 2019;**27**:1350–1363.
9. Beermann J, Piccoli M-T, Viereck J, Thum T. Non-coding RNAs in Development and Disease: Background, Mechanisms, and Therapeutic Approaches. *Physiol Rev* 2016;**96**:1297–1325.
10. Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. *Nat. Rev. Genet.* Nature Publishing Group; 2016. p. 47–62.
11. Zhou S, Jin J, Wang J, Zhang Z, Freedman JH, Zheng Y, Cai L. miRNAs in cardiovascular diseases: potential biomarkers, therapeutic targets and challenges. *Acta Pharmacol Sin* Nature Publishing Group; 2018;**39**:1073–1084.
12. Colpaert RMW, Calore M. MicroRNAs in Cardiac Diseases. *Cells* Multidisciplinary Digital Publishing Institute; 2019;**8**:737.
13. Poller W, Dimmeler S, Heymans S, Zeller T, Haas J, Karakas M, Leistner D-M, Jakob P, Nakagawa S, Blankenberg S, Engelhardt S, Thum T, Weber C, Meder B, Hajjar R, Landmesser U. Non-coding RNAs in cardiovascular diseases: diagnostic and therapeutic perspectives. *Eur Heart J* Oxford Academic; 2018;**39**:2704–2716.
14. Lim TB, Lavenniah A, Foo RSY. Circles in the heart and cardiovascular system. *Cardiovasc Res* NLM (Medline); 2020;**116**:269–278.
15. Boeckel J-N, Jaé N, Heumüller AW, Chen W, Boon RA, Stellos K, Zeiher AM, John

- D, Uchida S, Dimmeler S. Identification and Characterization of Hypoxia-Regulated Endothelial Circular RNA. *Circ Res* 2015;**117**:884–890.
16. Jaé N, Heumüller AW, Fouani Y, Dimmeler S. Long non-coding RNAs in vascular biology and disease. *Vascul Pharmacol* Elsevier Inc.; 2019;**114**:13–22.
 17. Holdt LM, Stahringer A, Sass K, Pichler G, Kulak NA, Wilfert W, Kohlmaier A, Herbst A, Northoff BH, Nicolaou A, Gäbel G, Beutner F, Scholz M, Thiery J, Musunuru K, Krohn K, Mann M, Teupser D. Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nat Commun* Nature Publishing Group; 2016;**7**:12429.
 18. Hall IF, Climent M, Quintavalle M, Farina FM, Schorn T, Zani S, Carullo P, Kunderfranco P, Civilini E, Condorelli G, Elia L. Circ_Lrp6, a Circular RNA Enriched in Vascular Smooth Muscle Cells, Acts as a Sponge Regulating miRNA-145 Function. *Circ Res* NLM (Medline); 2019;**124**:498–510.
 19. Du WW, Yang W, Chen Y, Wu Z-K, Foster FS, Yang Z, Li X, Yang BB. Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses. *Eur Heart J* 2017;**38**:1402–1412.
 20. Lim TB, Aliwarga E, Luu TDA, Li YP, Ng SL, Annadoray L, Sian S, Ackers-Johnson MA, Foo RS-Y. Targeting the highly abundant circular RNA circSlc8a1 in cardiomyocytes attenuates pressure overload induced hypertrophy. *Cardiovasc Res* 2019;**115**:1998–2007.
 21. Garikipati VNS, Verma SK, Cheng Z, Liang D, Truongcao MM, Cimini M, Yue Y, Huang G, Wang C, Benedict C, Tang Y, Mallareddy V, Ibetti J, Grisanti L, Schumacher SM, Gao E, Rajan S, Wilusz JE, Goukassian D, Houser SR, Koch WJ, Kishore R. Circular RNA CircFndc3b modulates cardiac repair after myocardial infarction via FUS/VEGF-A axis. *Nat Commun* Nature Publishing Group; 2019;**10**.
 22. Haussecker D. Current issues of RNAi therapeutics delivery and development. *J Control Release* Elsevier; 2014;**195**:49–54.
 23. Niel G Van, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nat. Rev. Mol. Cell Biol.* Nature Publishing Group; 2018. p. 213–228.
 24. Bang C, Antoniadou C, Antonopoulos AS, Eriksson U, Franssen C, Hamdani N, Lehmann L, Moessinger C, Mongillo M, Muhl L, Speer T, Thum T. Intercellular communication lessons in heart failure. *Eur J Heart Fail* John Wiley and Sons Ltd; 2015;**17**:1091–1103.
 25. Bär C, Thum T, Gonzalo-Calvo D de. Circulating miRNAs as mediators in cell-to-cell communication. *Epigenomics* 2019;**11**:111–113.
 26. Ottaviani L, Costa Martins PA da. Non-coding RNAs in cardiac hypertrophy. *J. Physiol.* Blackwell Publishing Ltd; 2017. p. 4037–4050.
 27. Zhang J, Li S, Li L, Li M, Guo C, Yao J, Mi S. Exosome and exosomal microRNA: Trafficking, sorting, and function. *Genomics, Proteomics Bioinforma.* Beijing Genomics Institute; 2015. p. 17–24.
 28. Iaconetti C, Sorrentino S, Rosa S De, Indolfi C. Exosomal miRNAs in Heart Disease.

Physiology (Bethesda) American Physiological Society; 2016;**31**:16–24.

29. Halkein J, Tabruyn SP, Ricke-Hoch M, Haghikia A, Nguyen NQN, Scherr M, Castermans K, Malvaux L, Lambert V, Thiry M, Sliwa K, Noel A, Martial JA, Hilfiker-Kleiner D, Struman I. MicroRNA-146a is a therapeutic target and biomarker for peripartum cardiomyopathy. *J Clin Invest* 2013;**123**:2143–2154.
30. Ribeiro-Rodrigues TM, Laundos TL, Pereira-Carvalho R, Batista-Almeida D, Pereira R, Coelho-Santos V, Silva AP, Fernandes R, Zuzarte M, Enguita FJ, Costa MC, Pinto-do-Ó P, Pinto MT, Gouveia P, Ferreira L, Mason JC, Pereira P, Kwak BR, Nascimento DS, Girão H. Exosomes secreted by cardiomyocytes subjected to ischaemia promote cardiac angiogenesis. *Cardiovasc Res* 2017;**113**:1338–1350.
31. Wang X, Huang W, Liu G, Cai W, Millard RW, Wang Y, Chang J, Peng T, Fan G-C. Cardiomyocytes mediate anti-angiogenesis in type 2 diabetic rats through the exosomal transfer of miR-320 into endothelial cells. *J Mol Cell Cardiol* Academic Press; 2014;**74**:139–150.
32. Qiao L, Hu S, Liu S, Zhang H, Ma H, Huang K, Li Z, Su T, Vandergriff A, Tang J, Allen T, Dinh P-U, Cores J, Yin Q, Li Y, Cheng K. microRNA-21-5p dysregulation in exosomes derived from heart failure patients impairs regenerative potential. *J Clin Invest* American Society for Clinical Investigation; 2019;**129**:2237–2250.
33. Yang Y, Li Y, Chen X, Cheng X, Liao Y, Yu X. Exosomal transfer of miR-30a between cardiomyocytes regulates autophagy after hypoxia. *J Mol Med (Berl)* Springer Verlag; 2016;**94**:711–724.
34. Hergenreider E, Heydt S, Tréguer K, Boettger T, Horrevoets AJG, Zeiher AM, Scheffer MP, Frangakis AS, Yin X, Mayr M, Braun T, Urbich C, Boon RA, Dimmeler S. Atheroprotective communication between endothelial cells and smooth muscle cells through miRNAs. *Nat Cell Biol* 2012;**14**:249–256.
35. He S, Wu C, Xiao J, Li D, Sun Z, Li M. Endothelial extracellular vesicles modulate the macrophage phenotype: Potential implications in atherosclerosis. *Scand J Immunol* Blackwell Publishing Ltd; 2018;**87**:e12648.
36. Ottaviani L, Sansonetti M, Costa Martins PA da. Myocardial cell-to-cell communication via microRNAs. *Non-coding RNA Res.* KeAi Communications Co.; 2018. p. 144–153.
37. Balkom BWM va., Jong OG d., Smits M, Brummelman J, Ouden K den, Bree PM d., Eijndhoven MAJ va., Pegtel DM, Stoorvogel W, Würdinger T, Verhaar MC. Endothelial cells require miR-214 to secrete exosomes that suppress senescence and induce angiogenesis in human and mouse endothelial cells. *Blood* American Society of Hematology; 2013;**121**:3997–4006.
38. Bang C, Batkai S, Dangwal S, Gupta SK, Foinquinos A, Holzmann A, Just A, Remke J, Zimmer K, Zeug A, Ponimaskin E, Schmiedl A, Yin X, Mayr M, Halder R, Fischer A, Engelhardt S, Wei Y, Schober A, Fiedler J, Thum T. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J Clin Invest* 2014;**124**:2136–2146.
39. Ibrahim AGE, Cheng K, Marbán E. Exosomes as critical agents of cardiac regeneration triggered by cell therapy. *Stem Cell Reports* Cell Press; 2014;**2**:606–619.

40. Feng Y, Huang W, Wani M, Yu X, Ashraf M. Ischemic preconditioning potentiates the protective effect of stem cells through secretion of exosomes by targeting Mecp2 via miR-22. *PLoS One* 2014;**9**:e88685.
41. Yu B, Kim HW, Gong M, Wang J, Millard RW, Wang Y, Ashraf M, Xu M. Exosomes secreted from GATA-4 overexpressing mesenchymal stem cells serve as a reservoir of anti-apoptotic microRNAs for cardioprotection. *Int J Cardiol* Elsevier Ireland Ltd; 2015;**182**:349–360.
42. Wang Y, Zhang L, Li Y, Chen L, Wang X, Guo W, Zhang X, Qin G, He S, Zimmerman A, Liu Y, Kim I, Weintraub NL, Tang Y. Exosomes/microvesicles from induced pluripotent stem cells deliver cardioprotective miRNAs and prevent cardiomyocyte apoptosis in the ischemic myocardium. *Int J Cardiol* Elsevier Ireland Ltd; 2015;**192**:61–69.
43. Khan M, Nickoloff E, Abramova T, Johnson J, Verma SK, Krishnamurthy P, Mackie AR, Vaughan E, Garikipati VNS, Benedict C, Ramirez V, Lambers E, Ito A, Gao E, Misener S, Luongo T, Elrod J, Qin G, Houser SR, Koch WJ, Kishore R. Embryonic stem cell-derived exosomes promote endogenous repair mechanisms and enhance cardiac function following myocardial infarction. *Circ Res* Lippincott Williams and Wilkins; 2015;**117**:52–64.
44. Huang P, Wang L, Li Q, Tian X, Xu J, Xu J, Xiong Y, Chen G, Qian H, Jin C, Yu Y, Cheng K, Qian L, Yang Y. Atorvastatin enhances the therapeutic efficacy of mesenchymal stem cells-derived exosomes in acute myocardial infarction via up-regulating long non-coding RNA H19. *Cardiovasc Res* 2020;**116**:353–367.
45. Kenneweg F, Bang C, Xiao K, Boulanger CM, Loyer X, Mazlan S, Schroen B, Hermans-Beijnsberger S, Foinquinos A, Hirt MN, Eschenhagen T, Funcke S, Stojanovic S, Genschel C, Schimmel K, Just A, Pfanne A, Scherf K, Dehmel S, Raemon-Buettner SM, Fiedler J, Thum T. Long Noncoding RNA-Enriched Vesicles Secreted by Hypoxic Cardiomyocytes Drive Cardiac Fibrosis. *Mol Ther - Nucleic Acids* Cell Press; 2019;**18**:363–374.
46. Komajda M, Pousset F, Isnard R, Lechat P. The role of the neurohormonal system in heart failure. *Heart*. BMJ Publishing Group Ltd; 1998. p. 17.
47. Sun M, Yu H, Zhang Y, Li Z, Gao W. MicroRNA-214 Mediates Isoproterenol-induced Proliferation and Collagen Synthesis in Cardiac Fibroblasts. *Sci Rep* Nature Publishing Group; 2015;**5**:1–10.
48. Chen YT, Wang J, Tong KS, Wong LL, Liew OW, Richards AM. The association of heart failure-related microRNAs with neurohormonal signaling. *Biochim Biophys Acta - Mol Basis Dis* Elsevier B.V.; 2017;**1863**:2031–2040.
49. Castaldi A, Zaglia T, Mauro V Di, Carullo P, Viggiani G, Borile G, Stefano B Di, Schiattarella GG, Gualazzi MG, Elia L, Stirparo GG, Colorito ML, Pironti G, Kunderfranco P, Esposito G, Bang ML, Mongillo M, Condorelli G, Catalucci D. MicroRNA-133 modulates the β 1-adrenergic receptor transduction cascade. *Circ Res* Lippincott Williams and Wilkins; 2014;**115**:273–283.
50. Zheng L, Xu CC, Chen WD, Shen WL, Ruan CC, Zhu LM, Zhu DL, Gao PJ. MicroRNA-155 regulates angiotensin II type 1 receptor expression and phenotypic differentiation in vascular adventitial fibroblasts. *Biochem Biophys Res Commun*

2010;**400**:483–488.

51. Li D, Yang P, Xiong Q, Song X, Yang X, Liu L, Yuan W, Rui YC. MicroRNA-125a/b-5p inhibits endothelin-1 expression in vascular endothelial cells. *J Hypertens* 2010;**28**:1646–1654.
52. Maharjan S, Mopidevi B, Kaw MK, Puri N, Kumar A. Human aldosterone synthase gene polymorphism promotes miRNA binding and regulates gene expression. *Physiol Genomics* American Physiological Society; 2014;**46**:860–865.
53. Arora P, Wu C, Khan AM, Bloch DB, Davis-Dusenbery BN, Ghorbani A, Spagnoli E, Martinez A, Ryan A, Tainsh LT, Kim S, Rong J, Huan T, Freedman JE, Levy D, Miller KK, Hata A, Monte F Del, Vandenwijngaert S, Swinnen M, Janssens S, Holmes TM, Buys ES, Bloch KD, Newton-Cheh C, Wang TJ. Atrial natriuretic peptide is negatively regulated by microRNA-425. *J Clin Invest* 2013;**123**:3378–3382.
54. Wong LL, Wee ASY, Lim JY, Ng JYX, Chong JPC, Liew OW, Lilyanna S, Martinez EC, Ackers-Johnson MA, Vardy LA, Armugam A, Jeyaseelan K, Ng TP, Lam CSP, Foo RSY, Richards AM, Chen Y-T. Natriuretic peptide receptor 3 (NPR3) is regulated by microRNA-100. *J Mol Cell Cardiol* Academic Press; 2015;**82**:13–21.
55. P P, AA V, SD A, H B, JG C, AJ C, V F, JR G-J, VP H, EA J, M J, C L, P N, JT P, B P, JP R, GM R, LM R, F R, FH R, P van der M. 2016 ESC Guidelines for the Diagnosis and Treatment of Acute and Chronic Heart Failure: The Task Force for the Diagnosis and Treatment of Acute and Chronic Heart Failure of the European Society of Cardiology (ESC). Developed With the Special Contribution of the Heart Failure Association (HFA) of the ESC. *Eur J Heart Fail Eur J Heart Fail*; 2016;**18**.
56. Nakamura K, Shichita T. Cellular and molecular mechanisms of sterile inflammation in ischaemic stroke. *J Biochem* 2019;**165**:459–464.
57. J A-C, P K, D G, E E, JA R-P, N H, CM W, RJ S, WA L, JM F. S-glutathionylation of Cryptic Cysteines Enhances Titin Elasticity by Blocking Protein Folding. *Cell Cell*; 2014;**156**.
58. Kolijn D, Pabel S, Tian Y, Lódi M, Herwig M, Carrizzo A, Zhazykbayeva S, Kovács Á, Fülöp GÁ, Falcão-Pires I, Reusch PH, Linthout S Van, Papp Z, Heerebeek L van, Vecchione C, Maier LS, Ciccarelli M, Tschöpe C, Mügge A, Bagi Z, Sossalla S, Hamdani N. Empagliflozin improves endothelial and cardiomyocyte function in human heart failure with preserved ejection fraction via reduced pro-inflammatory-oxidative pathways and protein kinase G α oxidation. *Cardiovasc Res* 2020;
59. Marchant DJ, Boyd JH, Lin DC, Granville DJ, Garmaroudi FS, McManus BM. Inflammation in myocardial diseases. *Circ. Res.* 2012. p. 126–144.
60. Martens CR, Bansal SS, Accornero F. Cardiovascular inflammation: RNA takes the lead. *J Mol Cell Cardiol* Academic Press; 2019;**129**:247–256.
61. Heymans S, Eriksson U, Lehtonen J, Cooper LT. The Quest for New Approaches in Myocarditis and Inflammatory Cardiomyopathy. *J. Am. Coll. Cardiol.* Elsevier USA; 2016. p. 2348–2364.
62. Corsten MF, Papageorgiou A, Verhesen W, Carai P, Lindow M, Obad S, Summer G, Coort SLM, Hazebroek M, Leeuwen R Van, Gijbels MJJ, Wijnands E, Biessen EAL,

- Winther MPJ De, Stassen FRM, Carmeliet P, Kauppinen S, Schroen B, Heymans S. MicroRNA Profiling Identifies MicroRNA-155 as an Adverse Mediator of Cardiac Injury and Dysfunction during Acute Viral Myocarditis. *Circ Res* 2012;**111**:415–425.
63. Yan L, Hu F, Yan X, Wei Y, Ma W, Wang Y, Lu S, Wang Z. Inhibition of microRNA-155 ameliorates experimental autoimmune myocarditis by modulating Th17/Treg immune response. *J Mol Med (Berl)* Springer Verlag; 2016;**94**:1063–1079.
 64. Zhang Y, Zhang M, Li X, Tang Z, Wang X, Zhong M, Suo Q, Zhang Y, Lv K. Silencing MicroRNA-155 Attenuates Cardiac Injury and Dysfunction in Viral Myocarditis via Promotion of M2 Phenotype Polarization of Macrophages. *Sci Rep* Nature Publishing Group; 2016;**6**:22613.
 65. Liu YL, Wu W, Xue Y, Gao M, Yan Y, Kong Q, Pang Y, Yang F. MicroRNA-21 and -146b are involved in the pathogenesis of murine viral myocarditis by regulating TH-17 differentiation. *Arch Virol* 2013;**158**:1953–1963.
 66. Zhao S, Yang G, Liu P-N, Deng Y-Y, Zhao Z, Sun T, Zhuo X-Z, Liu J-H, Tian Y, Zhou J, Yuan Z, Wu Y. miR-590-3p Is a Novel MicroRNA in Myocarditis by Targeting Nuclear Factor Kappa-B in vivo. *Cardiology* S. Karger AG; 2015;**132**:182–188.
 67. I.C. N, F.M. F, H.I. N, M.A. B, G. V-P, I.R. P, A.M.G. S, J.M. R, T. DB, C. C, J. L-V, J. K, E. C-N, L.R.P. F, Navarro IC, Ferreira FM, Nakaya HI, Baron MA, Vilar-Pereira G, Pereira IR, Silva AMG, Real JM, Brito T De, Chevillard C, Lannes-Vieira J, Kalil J, Cunha-Neto E, Ferreira LRP. MicroRNA transcriptome profiling in heart of trypanosoma cruzi-infected mice: Parasitological and cardiological Outcomes. *PLoS Negl Trop Dis* 2015;**9**:e0003828.
 68. Corsten MF, Heggermont W, Papageorgiou A-P, Deckx S, Tijmsa A, Verhesen W, Leeuwen R van, Carai P, Thibaut H-J, Custers K, Summer G, Hazebroek M, Verheyen F, Neyts J, Schroen B, Heymans S. The microRNA-221/-222 cluster balances the antiviral and inflammatory response in viral myocarditis. *Eur Heart J* 2015;**36**:2909–2919.
 69. Heymans S, Corsten MF, Verhesen W, Carai P, Leeuwen REW van, Custers K, Peters T, Hazebroek M, Stöger L, Wijnands E, Janssen BJ, Creemers EE, Pinto YM, Grimm D, Schürmann N, Vigorito E, Thum T, Stassen F, Yin X, Mayr M, Windt LJ de, Lutgens E, Wouters K, Winther MPJ de, Zacchigna S, Giacca M, Bilsen M van, Papageorgiou A-P, Schroen B. Macrophage microRNA-155 promotes cardiac hypertrophy and failure. *Circulation* 2013;**128**:1420–1432.
 70. Y Z, X L, X K, M Z, D W, Y L, K L. Long Non-Coding RNA AK085865 Ablation Confers Susceptibility to Viral Myocarditis by Regulating Macrophage Polarization. *J Cell Mol Med J Cell Mol Med*; 2020;**24**.
 71. Cui H, Banerjee S, Guo S, Xie N, Ge J, Jiang D, Zörnig M, Thannickal VJ, Liu G. Long noncoding RNA Malat1 regulates differential activation of macrophages and response to lung injury. *JCI insight* NLM (Medline); 2019;**4**.
 72. Masoumi F, Ghorbani S, Talebi F, Branton WG, Rajaei S, Power C, Noorbakhsh F. Malat1 long noncoding RNA regulates inflammation and leukocyte differentiation in experimental autoimmune encephalomyelitis. *J Neuroimmunol* Elsevier B.V.; 2019;**328**:50–59.

73. Chen H, Wang X, Yan X, Cheng X, He X, Zheng W. LncRNA MALAT1 regulates sepsis-induced cardiac inflammation and dysfunction via interaction with miR-125b and p38 MAPK/NFκB. *Int Immunopharmacol Elsevier B.V.*; 2018;**55**:69–76.
74. Gordon AD, Biswas S, Feng B, Chakrabarti S. MALAT1: A regulator of inflammatory cytokines in diabetic complications. *Endocrinol Diabetes Metab Wiley*; 2018;**1**:e00010.
75. Peters T, Hermans-Beijnsberger S, Beqqali A, Bitsch N, Nakagawa S, Prasanth K V, Windt LJ de, Oort RJ van, Heymans S, Schroen B. Long Non-Coding RNA Malat-1 Is Dispensable during Pressure Overload-Induced Cardiac Remodeling and Failure in Mice. *PLoS One* 2016;**11**:e0150236.
76. Pasmant E, Sabbagh A, Vidaud M, Bieche I. Pasmant E, Sabbagh A, Vidaud M, et al. ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS[J]. *Faseb Journal Official Publication of the Federation of American Societies for Experimental Biology*, 2011, 25(2)444. *FASEB J FASEB*; 2011;**25**:444–448.
77. Yoon J-H, Abdelmohsen K, Kim J, Yang X, Martindale JL, Tominaga-Yamanaka K, White EJ, Orjalo A V., Rinn JL, Kreft SG, Wilson GM, Gorospe M. Scaffold function of long non-coding RNA HOTAIR in protein ubiquitination. *Nat Commun Nature Publishing Group*; 2013;**4**:2939.
78. Zhou X, Han X, Wittfeldt A, Sun J, Liu C, Wang X, Gan L-M, Cao H, Liang Z. Long non-coding RNA ANRIL regulates inflammatory responses as a novel component of NF-κB pathway. *RNA Biol Taylor and Francis Inc.*; 2016;**13**:98–108.
79. Guo F, Tang C, Li Y, Liu Y, Lv P, Wang W, Mu Y. The interplay of LncRNA ANRIL and miR-181b on the inflammation-relevant coronary artery disease through mediating NF-κB signalling pathway. *J Cell Mol Med Blackwell Publishing Inc.*; 2018;**22**:5062–5075.
80. Song C-L, Wang J-P, Xue X, Liu N, Zhang X-H, Zhao Z, Liu J-G, Zhang C-P, Piao Z-H, Liu Y, Yang Y-B. Effect of Circular ANRIL on the Inflammatory Response of Vascular Endothelial Cells in a Rat Model of Coronary Atherosclerosis. *Cell Physiol Biochem S. Karger AG*; 2017;**42**:1202–1212.
81. Cohn JN, Ferrari R, Sharpe N. Cardiac remodeling-concepts and clinical implications: A consensus paper from an International Forum on Cardiac Remodeling. *J Am Coll Cardiol* 2000;**35**:569–582.
82. Chatterjee S, Bär C, Thum T. Linc- ing the Noncoding Genome to Heart Function: Beating Hypertrophy. *Trends Mol Med* 2017;**23**:577–579.
83. Leite-Moreira AM, Lourenço AP, Falcão-Pires I, Leite-Moreira AF. Pivotal role of microRNAs in cardiac physiology and heart failure. *Drug Discov. Today*. 2013. p. 1243–1249.
84. Elia L, Contu R, Quintavalle M, Varrone F, Chimenti C, Russo MA, Cimino V, Marinis L De, Frustaci A, Catalucci D, Condorelli G. Reciprocal regulation of microRNA-1 and insulin-like growth factor-1 signal transduction cascade in cardiac and skeletal muscle in physiological and pathological conditions. *Circulation NIH Public Access*; 2009;**120**:2377–2385.

85. Rodrigues PG, Leite-Moreira AF, Falcão-Pires I. Myocardial reverse remodeling: how far can we rewind? *Am J Physiol Heart Circ Physiol* American Physiological Society; 2016;**310**:H1402-22.
86. Hoshikawa E, Matsumura Y, Kubo T, Okawa M, Yamasaki N, Kitaoka H, Furuno T, Takata J, Doi YL. Effect of Left Ventricular Reverse Remodeling on Long-Term Prognosis After Therapy With Angiotensin-Converting Enzyme Inhibitors or Angiotensin II Receptor Blockers and β Blockers in Patients With Idiopathic Dilated Cardiomyopathy. *Am J Cardiol Excerpta Medica*; 2011;**107**:1065–1070.
87. T R, J S, P K, P N, J C, R P, D W. Combination of Left Ventricular Reverse Remodeling and Brain Natriuretic Peptide Level at One Year After Cardiac Resynchronization Therapy Predicts Long-Term Clinical Outcome. *PLoS One* PLoS One; 2019;**14**.
88. Shah R, Ziegler O, Yeri A, Liu X, Murthy V, Rabideau D, Xiao CY, Hanspers K, Belcher A, Tackett M, Rosenzweig A, Pico AR, Januzzi JL, Das S. MicroRNAs associated with reverse left ventricular remodeling in humans identify pathways of heart failure progression. *Circ Hear Fail* Lippincott Williams and Wilkins; 2018;**11**.
89. Wang J, Xu R, Lin F, Zhang S, Zhang G, Hu S, Zheng Z. MicroRNA: novel regulators involved in the remodeling and reverse remodeling of the heart. *Cardiology* 2009;**113**:81–88.
90. Akat KM, Moore-McGriff D, Morozov P, Brown M, Gogakos T, Correa Da Rosa J, Mihailovic A, Sauer M, Ji R, Ramarathnam A, Totary-Jain H, Williams Z, Tuschl T, Schulze PC. Comparative RNA-sequencing analysis of myocardial and circulating small RNAs in human heart failure and their utility as biomarkers. *Proc Natl Acad Sci U S A* National Academy of Sciences; 2014;**111**:11151–11156.
91. Bolkier Y, Nevo-Caspi Y, Salem Y, Vardi A, Mishali D, Paret G. Micro-RNA-208a, -208b, and -499 as Biomarkers for Myocardial Damage After Cardiac Surgery in Children. *Pediatr Crit Care Med* Lippincott Williams and Wilkins; 2016;**17**:e193-7.
92. Ucar A, Gupta SK, Fiedler J, Erikci E, Kardasinski M, Batkai S, Dangwal S, Kumarswamy R, Bang C, Holzmann A, Remke J, Caprio M, Jentzsch C, Engelhardt S, Geisendorf S, Glas C, Hofmann TG, Nessling M, Richter K, Schiffer M, Carrier L, Napp LC, Bauersachs J, Chowdhury K, Thum T. The miRNA-212/132 family regulates both cardiac hypertrophy and cardiomyocyte autophagy. *Nat Commun* 2012;**3**:1078.
93. Foinquinos A, Batkai S, Genschel C, Viereck J, Rump S, Gyöngyösi M, Traxler D, Riesenhuber M, Spannbauer A, Lukovic D, Weber N, Zlabinger K, Hašimbegović E, Winkler J, Fiedler J, Dangwal S, Fischer M, Roche J de la, Wojciechowski D, Kraft T, Garamvölgyi R, Neitzel S, Chatterjee S, Yin X, Bär C, Mayr M, Xiao K, Thum T. Preclinical development of a miR-132 inhibitor for heart failure treatment. *Nat Commun* Nature Publishing Group; 2020;**11**:633.
94. Marfella R, Filippo C Di, Potenza N, Sardu C, Rizzo MR, Siniscalchi M, Musacchio E, Barbieri M, Mauro C, Mosca N, Solimene F, Mottola MT, Russo A, Rossi F, Paolisso G, D'Amico M. Circulating microRNA changes in heart failure patients treated with cardiac resynchronization therapy: responders vs. non-responders. *Eur J Heart Fail* 2013;**15**:1277–1288.
95. Melman YF, Shah R, Danielson K, Xiao J, Simonson B, Barth A, Chakir K, Lewis GD,

- Lavender Z, Truong QA, Kleber A, Das R, Rosenzweig A, Wang Y, Kass DA, Singh JP, Das S. Circulating microRNA-30d is associated with response to cardiac resynchronization therapy in heart failure and regulates cardiomyocyte apoptosis: A translational pilot study. *Circulation*. Lippincott Williams and Wilkins; 2015. p. 2202–2216.
96. Villar A V, Merino D, Wenner M, Llano M, Cobo M, Montalvo C, García R, Martín-Durán R, Hurlé JM, Hurlé MA, Nistal JF. Myocardial gene expression of microRNA-133a and myosin heavy and light chains, in conjunction with clinical parameters, predict regression of left ventricular hypertrophy after valve replacement in patients with aortic stenosis. *Heart* 2011;**97**:1132–1137.
 97. Sucharov CC, Kao DP, Port JD, Karimpour-Fard A, Quaife RA, Minobe W, Nunley K, Lowes BD, Gilbert EM, Bristow MR. Myocardial microRNAs associated with reverse remodeling in human heart failure. *JCI Insight* American Society for Clinical Investigation; 2017;**2**:e89169.
 98. Matkovich SJ, Booven DJ Van, Youker KA, Torre-Amione G, Diwan A, Eschenbacher WH, Dorn LE, Watson MA, Margulies KB, Dorn GW. Reciprocal regulation of myocardial microRNAs and messenger RNA in human cardiomyopathy and reversal of the microRNA signature by biomechanical support. *Circulation* 2009;**119**:1263–1271.
 99. Yang KC, Yamada KA, Patel AY, Topkara VK, George I, Cheema FH, Ewald GA, Mann DL, Nerbonne JM. Deep RNA sequencing reveals dynamic regulation of myocardial noncoding RNAs in failing human heart and remodeling with mechanical circulatory support. *Circulation* Lippincott Williams and Wilkins; 2014;**129**:1009–1021.
 100. Wang LL, Liu Y, Chung JJ, Wang T, Gaffey AC, Lu M, Cavanaugh CA, Zhou S, Kanade R, Atluri P, Morrissey EE, Burdick JA. Local and sustained miRNA delivery from an injectable hydrogel promotes cardiomyocyte proliferation and functional regeneration after ischemic injury. *Nat Biomed Eng* Nature Publishing Group; 2017;**1**:983–992.
 101. Barsanti C, Trivella MG, D'Aurizio R, Baroudi M El, Baumgart M, Groth M, Caruso R, Verde A, Botta L, Cozzi L, Pitto L. Differential regulation of microRNAs in end-stage failing hearts is associated with left ventricular assist device unloading. *Biomed Res Int* 2015;**2015**:592512.
 102. Lok SI, Mil A van, Bovenschen N, Weide P van der, Kuik J van, Wichen D van, Peeters T, Siera E, Winkens B, Sluijter JPG, Doevendans PA, Costa Martins PA da, Jonge N de, Weger RA de. Post-transcriptional regulation of α -1-antichymotrypsin by microRNA-137 in chronic heart failure and mechanical support. *Circ Heart Fail* 2013;**6**:853–861.
 103. Wang T, O'Brien EC, Rogers JG, Jacoby DL, Chen ME, Testani JM, Bowles DE, Milano CA, Felker GM, Patel CB, Bonde PN, Ahmad T. Plasma Levels of MicroRNA-155 Are Upregulated with Long-Term Left Ventricular Assist Device Support. *ASAIO J* Lippincott Williams and Wilkins; 2017;**63**:536–541.
 104. Morley-Smith AC, Mills A, Jacobs S, Meyns B, Rega F, Simon AR, Pepper JR, Lyon AR, Thum T. Circulating microRNAs for predicting and monitoring response to mechanical circulatory support from a left ventricular assist device. *Eur J Heart Fail*

John Wiley and Sons Ltd; 2014;**16**:871–879.

105. Røsjø H, Dahl MB, Bye A, Andreassen J, Jørgensen M, Wisløff U, Christensen G, Edvardsen T, Omland T. Prognostic value of circulating microRNA-210 levels in patients with moderate to severe aortic stenosis. *PLoS One* Public Library of Science; 2014;**9**.
106. Ramani R, Vela D, Segura A, McNamara D, Lemster B, Samarendra V, Kormos R, Toyoda Y, Bermudez C, Frazier OH, Moravec CS, Gorcsan J, Taegtmeier H, McTiernan CF. A micro-ribonucleic acid signature associated with recovery from assist device support in 2 groups of patients with severe heart failure. *J Am Coll Cardiol* 2011;**58**:2270–2278.
107. Villar A V., García R, Merino D, Llano M, Cobo M, Montalvo C, Martín-Durán R, Hurlé MA, Nistal JF. Myocardial and circulating levels of microRNA-21 reflect left ventricular fibrosis in aortic stenosis patients. *Int J Cardiol Int J Cardiol*; 2013;**167**:2875–2881.
108. Fabiani I, Scatena C, Mazzanti CM, Conte L, Pugliese NR, Franceschi S, Lessi F, Menicagli M, Martino A De, Pratali S, Bortolotti U, Naccarato AG, Carrubba S La, Bello V Di. Micro-RNA-21 (biomarker) and global longitudinal strain (functional marker) in detection of myocardial fibrotic burden in severe aortic valve stenosis: A pilot study. *J Transl Med* BioMed Central Ltd.; 2016;**14**.
109. Varrone F, Gargano B, Carullo P, Silvestre D Di, Palma A De, Grasso L, Somma C Di, Mauri P, Benazzi L, Franzone A, Jotti GS, Bang ML, Esposito G, Colao A, Condorelli G, Catalucci D. The circulating level of FABP3 is an indirect biomarker of microRNA-1. *J Am Coll Cardiol J Am Coll Cardiol*; 2013;**61**:88–95.
110. Fabiani I, Pugliese NR, Calogero E, Conte L, Mazzanti MC, Scatena C, Scopelliti C, Tantillo E, Passiatore M, Angelillis M, Naccarato GA, Stefano R Di, Petronio AS, Bello V Di. MicroRNAs distribution in different phenotypes of Aortic Stenosis. *Sci Rep* Nature Publishing Group; 2018;**8**.
111. Kleeberger JA, Neuser J, Gonzalo-Calvo D de, Kempf T, Bauersachs J, Thum T, Widder JD. microRNA-206 correlates with left ventricular function after transcatheter aortic valve implantation. *Am J Physiol Heart Circ Physiol* NLM (Medline); 2017;**313**:H1261–H1266.
112. García R, Villar A V., Cobo M, Llano M, Martín-Durán R, Hurlé MA, Francisco Nistal J. Circulating levels of miR-133a predict the regression potential of left ventricular hypertrophy after valve replacement surgery in patients with aortic stenosis. *J Am Heart Assoc* 2013;**2**.
113. Chen Z, Li C, Xu Y, Li Y, Yang H, Rao L. Circulating level of miR-378 predicts left ventricular hypertrophy in patients with aortic stenosis. *PLoS One* PLoS One; 2014;**9**.
114. Gonzalo-Calvo D de, Cediel G, Bär C, Núñez J, Revuelta-Lopez E, Gavara J, Ríos-Navarro C, Llorente-Cortes V, Bodí V, Thum T, Bayes-Genis A. Circulating miR-1254 predicts ventricular remodeling in patients with ST-Segment-Elevation Myocardial Infarction: A cardiovascular magnetic resonance study. *Sci Rep* 2018;**8**:15115.
115. Lv P, Zhou M, He J, Meng W, Ma X, Dong S, Meng X, Zhao X, Wang X, He F. Circulating miR-208b and miR-34a are associated with left ventricular remodeling

- after acute myocardial infarction. *Int J Mol Sci* MDPI AG; 2014;**15**:5774–5788.
116. Grabmaier U, Clauss S, Gross L, Klier I, Franz WM, Steinbeck G, Wakili R, Theiss HD, Brenner C. Diagnostic and prognostic value of miR-1 and miR-29b on adverse ventricular remodeling after acute myocardial infarction – The SITAGRAMI-miR analysis. *Int J Cardiol* Elsevier Ireland Ltd; 2017;**244**:30–36.
 117. Maciejak A, Kostarska-Srokosz E, Gierlak W, Dluzniewski M, Kuch M, Marchel M, Opolski G, Kiliszek M, Matlak K, Dobrzycki S, Lukasik A, Segiet A, Sygitowicz G, Sitkiewicz D, Gora M, Burzynska B. Circulating miR-30a-5p as a prognostic biomarker of left ventricular dysfunction after acute myocardial infarction. *Sci Rep* Nature Publishing Group; 2018;**8**:9883.
 118. Friedberg MK, Redington AN. Right versus left ventricular failure: Differences, similarities, and interactions. *Circulation* Lippincott Williams and Wilkins; 2014;**129**:1033–1044.
 119. Batkai S, Bär C, Thum T. MicroRNAs in right ventricular remodelling. *Cardiovasc Res* 2017;**113**:1433–1440.
 120. Shi L, Kojonazarov B, Elgheznawy A, Popp R, Dahal BK, Böhm M, Pullamsetti SS, Ghofrani H-A, Gödecke A, Jungmann A, Katus HA, Müller OJ, Schermuly RT, Fisslthaler B, Seeger W, Fleming I. miR-223-IGF-IR signalling in hypoxia- and load-induced right-ventricular failure: a novel therapeutic approach. *Cardiovasc Res* 2016;**111**:184–193.
 121. Potus F, Ruffenach G, Dahou A, Thebault C, Breuils-Bonnet S, Tremblay È, Nadeau V, Paradis R, Graydon C, Wong R, Johnson I, Paulin R, Lajoie AC, Perron J, Charbonneau E, Joubert P, Pibarot P, Michelakis ED, Provencher S, Bonnet S. Downregulation of MicroRNA-126 Contributes to the Failing Right Ventricle in Pulmonary Arterial Hypertension. *Circulation* Lippincott Williams and Wilkins; 2015;**132**:932–943.
 122. Liang D, Xu X, Deng F, Feng J, Zhang H, Liu Y, Zhang Y, Pan L, Liu Y, Zhang D, Li J, Liang X, Sun Y, Xiao J, Chen Y-H. miRNA-940 reduction contributes to human Tetralogy of Fallot development. *J Cell Mol Med* Blackwell Publishing Inc.; 2014;**18**:1830–1839.
 123. Zhang H, Liu S, Dong T, Yang J, Xie Y, Wu Y, Kang K, Hu S, Gou D, Wei Y. Profiling of differentially expressed microRNAs in arrhythmogenic right ventricular cardiomyopathy. *Sci Rep* Nature Publishing Group; 2016;**6**:28101.
 124. Wang R, Zhou S, Wu P, Li M, Ding X, Sun L, Xu X, Zhou X, Zhou L, Cao C, Fei G. Identifying Involvement of H19-miR-675-3p-IGF1R and H19-miR-200a-PDCD4 in Treating Pulmonary Hypertension with Melatonin. *Mol Ther Nucleic Acids* Cell Press; 2018;**13**:44–54.
 125. Salvo TG Di, Guo Y, Su YR, Clark T, Brittain E, Absi T, Maltais S, Hemnes A. Right ventricular long noncoding RNA expression in human heart failure. *Pulm Circ* University of Chicago Press; 2015;**5**:135–161.
 126. Chatterjee S, Gupta SK, Bär C, Thum T. Noncoding RNAs: potential regulators in cardioncology. *Am J Physiol Circ Physiol* 2019;**316**:H160–H168.

127. Tony H, Yu K, Qiutang Z. MicroRNA-208a Silencing Attenuates Doxorubicin Induced Myocyte Apoptosis and Cardiac Dysfunction. *Oxid Med Cell Longev* 2015;**2015**:597032.
128. Wang JX, Zhang XJ, Feng C, Sun T, Wang K, Wang Y, Zhou LY, Li PF. MicroRNA-532-3p regulates mitochondrial fission through targeting apoptosis repressor with caspase recruitment domain in doxorubicin cardiotoxicity. *Cell Death Dis* Nature Publishing Group; 2015;**6**.
129. Fiedler J, Gupta SK, Thum T. Identification of cardiovascular microRNA targetomes. *J Mol Cell Cardiol* 2011;**51**:674–681.
130. Yin Z, Zhao Y, Li H, Yan M, Zhou L, Chen C, Wang DW. miR-320a mediates doxorubicin-induced cardiotoxicity by targeting VEGF signal pathway. *Aging (Albany NY)* Impact Journals LLC; 2016;**8**:192–207.
131. Gupta SK, Garg A, Avramopoulos P, Engelhardt S, Streckfuss-Bömeke K, Batkai S, Thum T. miR-212/132 Cluster Modulation Prevents Doxorubicin-Mediated Atrophy and Cardiotoxicity. *Mol Ther* Cell Press; 2019;**27**:17–28.
132. Gupta SK, Bang C, Thum T. Circulating MicroRNAs as Biomarkers and Potential Paracrine Mediators of Cardiovascular Disease. *Circ Cardiovasc Genet* 2010;**3**:484–488.
133. Rigaud VOC, Ferreira LRP, Ayub-Ferreira SM, ávila MS, Brandão SMG, Cruz FD, Santos MHH, Cruz CBBV, Alves MSL, Issa VS, Guimarães G V., Cunha-Neto E, Bocchi EA. Circulating miR-1 as a potential biomarker of doxorubicin-induced cardiotoxicity in breast cancer patients. *Oncotarget* Impact Journals LLC; 2017;**8**:6994–7002.
134. Leger KJ, Leonard D, Nielson D, Lemos JA de, Mammen PPA, Winick NJ. Circulating microRNAs: Potential Markers of Cardiotoxicity in Children and Young Adults Treated With Anthracycline Chemotherapy. *J Am Heart Assoc* 2017;**6**.
135. Oatmen KE, Toro-Salazar OH, Hauser K, Zellars KN, Mason KC, Hor K, Gillan E, Zeiss CJ, Gatti DM, Spinale FG. Identification of a novel microRNA profile in pediatric patients with cancer treated with anthracycline chemotherapy. *Am J Physiol Heart Circ Physiol* NLM (Medline); 2018;**315**:H1443–H1452.
136. Li H-Q, Wu Y-B, Yin C-S, Chen L, Zhang Q, Hu L-Q. Obestatin attenuated doxorubicin-induced cardiomyopathy via enhancing long noncoding Mhrt RNA expression. *Biomed Pharmacother* Elsevier Masson SAS; 2016;**81**:474–481.
137. Chen L, Yan K-P, Liu X-C, Wang W, Li C, Li M, Qiu C-G. Valsartan regulates TGF- β /Smads and TGF- β /p38 pathways through lncRNA CHRF to improve doxorubicin-induced heart failure. *Arch Pharm Res* Pharmaceutical Society of Korea; 2018;**41**:101–109.
138. Wang K, Gan TY, Li N, Liu CY, Zhou LY, Gao JN, Chen C, Yan KW, Ponnusamy M, Zhang YH, Li PF. Circular RNA mediates cardiomyocyte death via miRNA-dependent upregulation of MTP18 expression. *Cell Death Differ* Nature Publishing Group; 2017;**24**:1111–1120.
139. Gupta SK, Garg A, Bär C, Chatterjee S, Foinquinos A, Milting H, Streckfuß-Bömeke

- K, Fiedler J, Thum T. Quaking Inhibits Doxorubicin-Mediated Cardiotoxicity Through Regulation of Cardiac Circular RNA Expression Novelty and Significance. *Circ Res* 2018;**122**:246–254.
140. Chen JF, Murchison EP, Tang R, Callis TE, Tatsuguchi M, Deng Z, Rojas M, Hammond SM, Schneider MD, Selzman CH, Meissner G, Patterson C, Hannon GJ, Wang DZ. Targeted deletion of Dicer in the heart leads to dilated cardiomyopathy and heart failure. *Proc Natl Acad Sci U S A* 2008;**105**:2111–2116.
 141. Zhao Y, Ransom JF, Li A, Vedantham V, Drehele M von, Muth AN, Tsuchihashi T, McManus MT, Schwartz RJ, Srivastava D. Dysregulation of cardiogenesis, cardiac conduction, and cell cycle in mice lacking miRNA-1-2. *Cell* 2007;**129**:303–317.
 142. Torrini C, Cubero RJ, Dirx E, Braga L, Ali H, Prosdocimo G, Gutierrez MI, Collesi C, Licastro D, Zentilin L, Mano M, Zacchigna S, Vendruscolo M, Marsili M, Samal A, Giacca M. Common Regulatory Pathways Mediate Activity of MicroRNAs Inducing Cardiomyocyte Proliferation. *Cell Rep Elsevier B.V.*; 2019;**27**:2759-2771.e5.
 143. Eulalio A, Mano M, Ferro MD, Zentilin L, Sinagra G, Zacchigna S, Giacca M. Functional screening identifies miRNAs inducing cardiac regeneration. *Nature* 2012;**492**:376–381.
 144. Porrello ER, Johnson BA, Aurora AB, Simpson E, Nam Y-J, Matkovich SJ, Dorn GW, Rooij E van, Olson EN. MiR-15 family regulates postnatal mitotic arrest of cardiomyocytes. *Circ Res* 2011;**109**:670–679.
 145. Hoelscher SC, Doppler SA, Dreßen M, Lahm H, Lange R, Krane M. MicroRNAs: pleiotropic players in congenital heart disease and regeneration. *J Thorac Dis AME Publishing Company*; 2017;**9**:S64–S81.
 146. Ohtani K, Dimmeler S. Control of cardiovascular differentiation by microRNAs. *Basic Res Cardiol* 2011;**106**:5–11.
 147. Ieda M, Fu J-D, Delgado-Olguin P, Vedantham V, Hayashi Y, Bruneau BG, Srivastava D. Direct reprogramming of fibroblasts into functional cardiomyocytes by defined factors. *Cell Elsevier*; 2010;**142**:375–386.
 148. Jayawardena TM, Egemnazarov B, Finch EA, Zhang L, Alan Payne J, Pandya K, Zhang Z, Rosenberg P, Mirotsoy M, Dzau VJ. MicroRNA-mediated in vitro and in vivo direct reprogramming of cardiac fibroblasts to cardiomyocytes. *Circ Res* 2012;**110**:1465–1473.
 149. Muraoka N, Yamakawa H, Miyamoto K, Sadahiro T, Umei T, Isomi M, Nakashima H, Akiyama M, Wada R, Inagawa K, Nishiyama T, Kaneda R, Fukuda T, Takeda S, Tohyama S, Hashimoto H, Kawamura Y, Goshima N, Aeiba R, Yamagishi H, Fukuda K, Ieda M. MiR-133 promotes cardiac reprogramming by directly repressing Snail and silencing fibroblast signatures. *EMBO J EMBO*; 2014;**33**:1565–1581.
 150. Miyamoto K, Akiyama M, Tamura F, Isomi M, Yamakawa H, Sadahiro T, Muraoka N, Kojima H, Haginiwa S, Kurotsu S, Tani H, Wang L, Qian L, Inoue M, Ide Y, Kurokawa J, Yamamoto T, Seki T, Aeiba R, Yamagishi H, Fukuda K, Ieda M. Direct In Vivo Reprogramming with Sendai Virus Vectors Improves Cardiac Function after Myocardial Infarction. *Cell Stem Cell Cell Press*; 2018;**22**:91-103.e5.

151. Rotini A, Martínez-Sarrà E, Pozzo E, Sampaolesi M. Interactions between microRNAs and long non-coding RNAs in cardiac development and repair. *Pharmacol Res Academic Press*; 2018;**127**:58–66.
152. Sweta S, Dudnakova T, Sudheer S, Baker AH, Bhushan R. Importance of Long Non-coding RNAs in the Development and Disease of Skeletal Muscle and Cardiovascular Lineages. *Front cell Dev Biol Frontiers Media SA*; 2019;**7**:228.
153. Ounzain S, Pezzuto I, Micheletti R, Burdet F, Sheta R, Nemir M, Gonzales C, Sarre A, Alexanian M, Blow MJ, May D, Johnson R, Dauvillier J, Pennacchio LA, Pedrazzini T. Functional importance of cardiac enhancer-associated noncoding RNAs in heart development and disease. *J Mol Cell Cardiol* 2014;**76**:55–70.
154. Ounzain S, Micheletti R, Beckmann T, Schroen B, Alexanian M, Pezzuto I, Crippa S, Nemir M, Sarre A, Johnson R, Dauvillier J, Burdet F, Ibberson M, Guigó R, Xenarios I, Heymans S, Pedrazzini T. Genome-wide profiling of the cardiac transcriptome after myocardial infarction identifies novel heart-specific long non-coding RNAs. *Eur Heart J* 2015;**36**:353–68a.
155. Klattenhoff CA, Scheuermann JC, Surface LE, Bradley RK, Fields PA, Steinhäuser ML, Ding H, Butty VL, Torrey L, Haas S, Abo R, Tabebordbar M, Lee RT, Burge CB, Boyer LA. Braveheart, a long noncoding RNA required for cardiovascular lineage commitment. *Cell* 2013;**152**:570–583.
156. Kurian L, Aguirre A, Sancho-Martinez I, Benner C, Hishida T, Nguyen TB, Reddy P, Nivet E, Krause MN, Nelles DA, Rodriguez Esteban C, Campistol JM, Yeo GW, Izpisua Belmonte JC. Identification of novel long noncoding RNAs underlying vertebrate cardiovascular development. *Circulation* 2015;**131**:1278–1290.
157. Chen G, Li H, Li X, Li B, Zhong L, Huang S, Zheng H, Li M, Jin G, Liao W, Liao Y, Chen Y, Bin J. Loss of long non-coding RNA CRRL promotes cardiomyocyte regeneration and improves cardiac repair by functioning as a competing endogenous RNA. *J Mol Cell Cardiol Academic Press*; 2018;**122**:152–164.
158. Werfel S, Nothjunge S, Schwarzmayer T, Strom T-M, Meitinger T, Engelhardt S. Characterization of circular RNAs in human, mouse and rat hearts. *J Mol Cell Cardiol* 2016;**98**:103–107.
159. Szabo L, Morey R, Palpant NJ, Wang PL, Afari N, Jiang C, Parast MM, Murry CE, Laurent LC, Salzman J. Statistically based splicing detection reveals neural enrichment and tissue-specific induction of circular RNA during human fetal development. *Genome Biol BioMed Central Ltd.*; 2015;**16**:126.
160. Khan MAF, Reckman YJ, Aufiero S, Hoogenhof MMG van den, Made I van der, Beqqali A, Koolbergen DR, Rasmussen TB, Velden J van der, Creemers EE, Pinto YM. RBM20 Regulates Circular RNA Production From the Titin Gene. *Circ Res Lippincott Williams and Wilkins*; 2016;**119**:996–1003.
161. Tian Y, Liu Y, Wang T, Zhou N, Kong J, Chen L, Snitow M, Morley M, Li D, Petrenko N, Zhou S, Lu M, Gao E, Koch WJ, Stewart KM, Morrissey EE. A microRNA-Hippo pathway that promotes cardiomyocyte proliferation and cardiac regeneration in mice. *Sci Transl Med* 2015;**7**:279ra38–279ra38.
162. Borden A, Kurian J, Nickoloff E, Yang Y, Troupes CD, Ibetti J, Lucchese AM, Gao E,

- Mohsin S, Koch WJ, Houser SR, Kishore R, Khan M. Transient Introduction of miR-294 in the Heart Promotes Cardiomyocyte Cell Cycle Reentry After Injury. *Circ Res* NLM (Medline); 2019;**125**:14–25.
163. Wang Y, Baskerville S, Shenoy A, Babiarz JE, Baehner L, Brelloch R. Embryonic stem cell-specific microRNAs regulate the G1-S transition and promote rapid proliferation. *Nat Genet* 2008;**40**:1478–1483.
 164. Marson A, Levine SS, Cole MF, Frampton GM, Brambrink T, Johnstone S, Guenther MG, Johnston WK, Wernig M, Newman J, Calabrese JM, Dennis LM, Volkert TL, Gupta S, Love J, Hannett N, Sharp PA, Bartel DP, Jaenisch R, Young RA. Connecting microRNA genes to the core transcriptional regulatory circuitry of embryonic stem cells. *Cell* 2008;**134**:521–533.
 165. Mogilyansky E, Rigoutsos I. The miR-17/92 cluster: a comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease. *Cell Death Differ* 2013;**20**:1603–1614.
 166. Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M, Prueitt RL, Yanaihara N, Lanza G, Scarpa A, Vecchione A, Negrini M, Harris CC, Croce CM. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci U S A* National Academy of Sciences; 2006;**103**:2257–2261.
 167. He L, Thomson JM, Hemann MT, Hernando-Monge E, Mu D, Goodson S, Powers S, Cordon-Cardo C, Lowe SW, Hannon GJ, Hammond SM. A microRNA polycistron as a potential human oncogene. *Nature* 2005;**435**:828–833.
 168. Chen J, Huang ZP, Seok HY, Ding J, Kataoka M, Zhang Z, Hu X, Wang G, Lin Z, Wang S, Pu WT, Liao R, Wang DZ. Mir-17-92 induce cardiomyocyte proliferation in postnatal and adult hearts, 2013. *Circ Res* American Heart Association, Inc.; 2013;**112**:1557–1566.
 169. Gao F, Kataoka M, Liu N, Liang T, Huang Z-P, Gu F, Ding J, Liu J, Zhang F, Ma Q, Wang Y, Zhang M, Hu X, Kyselovic J, Hu X, Pu WT, Wang J, Chen J, Wang D-Z. Therapeutic role of miR-19a/19b in cardiac regeneration and protection from myocardial infarction. *Nat Commun* Nature Publishing Group; 2019;**10**:1802.
 170. Cao X, Wang J, Wang Z, Du J, Yuan X, Huang W, Meng J, Gu H, Nie Y, Ji B, Hu S, Zheng Z. MicroRNA profiling during rat ventricular maturation: A role for miR-29a in regulating cardiomyocyte cell cycle re-entry. *FEBS Lett* 2013;**587**:1548–1555.
 171. Hullinger TG, Montgomery RL, Seto AG, Dickinson BA, Semus HM, Lynch JM, Dalby CM, Robinson K, Stack C, Latimer PA, Hare JM, Olson EN, Rooij E van. Inhibition of miR-15 protects against cardiac ischemic injury. *Circ Res* 2012;**110**:71–81.
 172. Saucedo LJ, Edgar BA. Filling out the Hippo pathway. *Nat. Rev. Mol. Cell Biol.* 2007. p. 613–621.
 173. Pan D. The hippo signaling pathway in development and cancer. *Dev. Cell.* 2010. p. 491–505.
 174. Diez-Cuñado M, Wei K, Bushway PJ, Maurya MR, Perera R, Subramaniam S, Ruiz-

- Lozano P, Mercola M. miRNAs that Induce Human Cardiomyocyte Proliferation Converge on the Hippo Pathway. *Cell Rep Elsevier B.V.*; 2018;**23**:2168–2174.
175. Ounzain S, Pedrazzini T. The promise of enhancer-associated long noncoding RNAs in cardiac regeneration. *Trends Cardiovasc Med Elsevier Inc.*; 2015;**25**:592–602.
 176. Cai B, Ma W, Ding F, Zhang L, Huang Q, Wang X, Hua B, Xu J, Li J, Bi C, Guo S, Yang F, Han Z, Li Y, Yan G, Yu Y, Bao Z, Yu M, Li F, Tian Y, Pan Z, Yang B. The Long Noncoding RNA CAREL Controls Cardiac Regeneration. *J Am Coll Cardiol Elsevier USA*; 2018;**72**:534–550.
 177. Li X, He X, Wang H, Li M, Huang S, Chen G, Jing Y, Wang S, Chen Y, Liao W, Liao Y, Bin J. Loss of AZIN2 splice variant facilitates endogenous cardiac regeneration. *Cardiovasc Res* 2018;**114**:1642–1655.
 178. Chen Y, Li X, Li B, Wang H, Li M, Huang S, Sun Y, Chen G, Si X, Huang C, Liao W, Liao Y, Bin J. Long Non-coding RNA ECRAR Triggers Post-natal Myocardial Regeneration by Activating ERK1/2 Signaling. *Mol Ther Cell Press*; 2019;**27**:29–45.
 179. Li B, Hu Y, Li X, Jin G, Chen X, Chen G, Chen Y, Huang S, Liao W, Liao Y, Teng Z, Bin J. Sirt1 Antisense Long Noncoding RNA Promotes Cardiomyocyte Proliferation by Enhancing the Stability of Sirt1. *J Am Heart Assoc American Heart Association Inc.*; 2018;**7**:e009700.
 180. Ponnusamy M, Liu F, Zhang Y-H, Li R-B, Zhai M, Liu F, Zhou L-Y, Liu C-Y, Yan K-W, Dong Y-H, Wang M, Qian L-L, Shan C, Xu S, Wang Q, Zhang Y-H, Li P-F, Zhang J, Wang K. Long Noncoding RNA CPR (Cardiomyocyte Proliferation Regulator) Regulates Cardiomyocyte Proliferation and Cardiac Repair. *Circulation Lippincott Williams and Wilkins*; 2019;**139**:2668–2684.
 181. Zhang X, Azhar G, Wei JY. The Expression of microRNA and microRNA Clusters in the Aging Heart. *PLoS One* 2012;**7**:e34688.
 182. Zhang X, Azhar G, Williams ED, Rogers SC, Wei JY. MicroRNA Clusters in the Adult Mouse Heart : Age-Associated Changes. *Biomed Res Int Hindawi Publishing Corporation*; 2015;**2015**:732397.
 183. Jazbutyte V, Fiedler J, Kneitz S, Galuppo P, Just A, Holzmann A, Bauersachs J, Thum T. MicroRNA-22 increases senescence and activates cardiac fibroblasts in the aging heart. *Age (Omaha)* 2013;**35**:747–762.
 184. Yamakuchi M, Lowenstein CJ. MiR-34, SIRT1, and p53: The feedback loop. *Cell Cycle* 2009;**8**:712–715.
 185. Potente M, Ghaeni L, Baldessari D, Mostoslavsky R, Rossig L, Dequiedt F, Haendeler J, Mione M, Dejana E, Alt FW, Zeiher AM, Dimmeler S. SIRT1 controls endothelial angiogenic functions during vascular growth. *Genes Dev* 2007;**21**:2644–2658.
 186. Ito T, Yagi S, Yamakuchi M. MicroRNA-34a regulation of endothelial senescence. *Biochem Biophys Res Commun* 2010;**398**:735–740.
 187. Boon RA, Iekushi K, Lechner S, Seeger T, Fischer A, Heydt S, Kaluza D, Tréguer K, Carmona G, Bonauer A, Horrevoets AJG, Didier N, Girmatsion Z, Biliczki P, Ehrlich JR, Katus HA, Müller OJ, Potente M, Zeiher AM, Hermeking H, Dimmeler S.

- MicroRNA-34a regulates cardiac ageing and function. *Nature* 2013;**495**:107–110.
188. Liu H, Wu H-Y, Wang W-Y, Zhao Z-L, Liu X-Y, Wang L-Y. Regulation of miR-92a on vascular endothelial aging via mediating Nrf2-KEAP1-ARE signal pathway. *Eur Rev Med Pharmacol Sci* 2017;**21**:2734–2742.
 189. Kuosmanen SM, Kansanen E, Sihvola V, Levonen A-L. MicroRNA Profiling Reveals Distinct Profiles for Tissue-Derived and Cultured Endothelial Cells. *Sci Rep* 2017;**7**:10943.
 190. Almen GC van, Verhesen W, Leeuwen REW Van, Vrie M van de, Eurlings C, Schellings MW, Swinnen M, Cleutjens JPM, Zandvoort MAMJ van, Heymans S, Schroen B. MicroRNA-18 and microRNA-19 regulate CTGF and TSP-1 expression in age-related heart failure. *Aging Cell* 2011;**10**:769–779.
 191. Du WW, Li X, Li T, Li H, Khorshidi A, Liu F, Yang BB. The microRNA miR-17-3p inhibits mouse cardiac fibroblast senescence by targeting Par4. *J Cell Sci* 2015;**128**:293–304.
 192. Boon RA, Hofmann P, Michalik KM, Lozano-Vidal N, Berghäuser D, Fischer A, Knau A, Jaé N, Schürmann C, Dimmeler S. Long Noncoding RNA Meg3 Controls Endothelial Cell Aging and Function. *J Am Coll Cardiol* 2016;**68**:2589–2591.
 193. Hofmann P, Sommer J, Theodorou K, Kirchhof L, Fischer A, Li Y, Perisic L, Hedin U, Maegdefessel L, Dimmeler S, Boon RA. Long non-coding RNA H19 regulates endothelial cell aging via inhibition of STAT3 signalling. *Cardiovasc Res* 2019;**115**:230–242.
 194. Li DY, Busch A, Jin H, Chernogubova E, Pelisek J, Karlsson J, Sennblad B, Liu S, Lao S, Hofmann P, Bäcklund A, Eken SM, Roy J, Eriksson P, Dacken B, Ramanujam D, Dueck A, Engelhardt S, Boon RA, Eckstein H-H, Spin JM, Tsao PS, Maegdefessel L. H19 Induces Abdominal Aortic Aneurysm Development and Progression. *Circulation* 2018;**138**:1551–1568.
 195. Abdelmohsen K, Panda A, Kang M-J, Xu J, Selimyan R, Yoon J-H, Martindale JL, De S, Wood WH, Becker KG, Gorospe M. Senescence-associated lncRNAs: senescence-associated long noncoding RNAs. *Aging Cell* John Wiley & Sons, Ltd (10.1111); 2013;**12**:890–900.
 196. Yan B, Yao J, Liu J-Y, Li X-M, Wang X-Q, Li Y-J, Tao Z-F, Song Y-C, Chen Q, Jiang Q. lncRNA-MIAT regulates microvascular dysfunction by functioning as a competing endogenous RNA. *Circ Res* 2015;**116**:1143–1156.
 197. Du WW, Yang W, Chen Y, Wu Z, Foster FS, Yang Z, Li X, Yang BB. Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses. *Eur Heart J* 2017;**38**:1402–1412.
 198. Elmén J, Lindow M, Schütz S, Lawrence M, Petri A, Obad S, Lindholm M, Hedtjärn M, Hansen HF, Berger U, Gullans S, Kearney P, Sarnow P, Straarup EM, Kauppinen S. LNA-mediated microRNA silencing in non-human primates. *Nature* Nature Publishing Group; 2008;**452**:896–899.
 199. Fluiter K, Mook ORF, Vreijling J, Langkjaer N, Højland T, Wengel J, Baas F. Filling the gap in LNA antisense oligo gapmers: the effects of unlocked nucleic acid (UNA)

- and 4'-C-hydroxymethyl-DNA modifications on RNase H recruitment and efficacy of an LNA gapmer. *Mol Biosyst* 2009;**5**:838–843.
200. Yang Y, Cheng H-W, Qiu Y, Dupee D, Noonan M, Lin Y-D, Fisch S, Unno K, Sereti K-I, Liao R. MicroRNA-34a Plays a Key Role in Cardiac Repair and Regeneration Following Myocardial Infarction. *Circ Res* Lippincott Williams and Wilkins; 2015;**117**:450–459.
 201. Aguirre A, Montserrat N, Zacchigna S, Nivet E, Hishida T, Krause MN, Kurian L, Ocampo A, Vazquez-Ferrer E, Rodriguez-Esteban C, Kumar S, Moresco JJ, Yates JR, Campistol JM, Sancho-Martinez I, Giacca M, Izpisua Belmonte JC. In vivo activation of a conserved microRNA program induces mammalian heart regeneration. *Cell Stem Cell* Cell Press; 2014;**15**:589–604.
 202. Hu Y, Jin G, Li B, Chen Y, Zhong L, Chen G, Chen X, Zhong J, Liao W, Liao Y, Wang Y, Bin J. Suppression of miRNA let-7i-5p promotes cardiomyocyte proliferation and repairs heart function post injury by targeting CCND2 and E2F2. *Clin Sci (Lond)* Portland Press Ltd; 2019;**133**:425–441.
 203. Gabisonia K, Prosdocimo G, Aquaro GD, Carlucci L, Zentilin L, Secco I, Ali H, Braga L, Gorgodze N, Bernini F, Burchielli S, Collesi C, Zandonà L, Sinagra G, Piacenti M, Zacchigna S, Bussani R, Recchia FA, Giacca M. MicroRNA therapy stimulates uncontrolled cardiac repair after myocardial infarction in pigs. *Nature* Nature Publishing Group; 2019;**569**:418–422.
 204. Lesizza P, Prosdocimo G, Martinelli V, Sinagra G, Zacchigna S, Giacca M. Single-Dose Intracardiac Injection of Pro-Regenerative MicroRNAs Improves Cardiac Function after Myocardial Infarction. *Circ Res* Lippincott Williams and Wilkins; 2017;**120**:1298–1304.
 205. Deng S, Zhao Q, Zhen L, Zhang C, Liu C, Wang G, Zhang L, Bao L, Lu Y, Meng L, Lü J, Yu P, Lin X, Zhang Y, Chen Y-H, Fan H, Cho WC, Liu Z, Yu Z. Neonatal Heart-Enriched miR-708 Promotes Proliferation and Stress Resistance of Cardiomyocytes in Rodents. *Theranostics* Ivyspring International Publisher; 2017;**7**:1953–1965.
 206. Grimm D, Lee JS, Wang L, Desai T, Akache B, Storm TA, Kay MA. In Vitro and In Vivo Gene Therapy Vector Evolution via Multispecies Interbreeding and Retargeting of Adeno-Associated Viruses. *J Virol* American Society for Microbiology (ASM); 2008;**82**:5887.

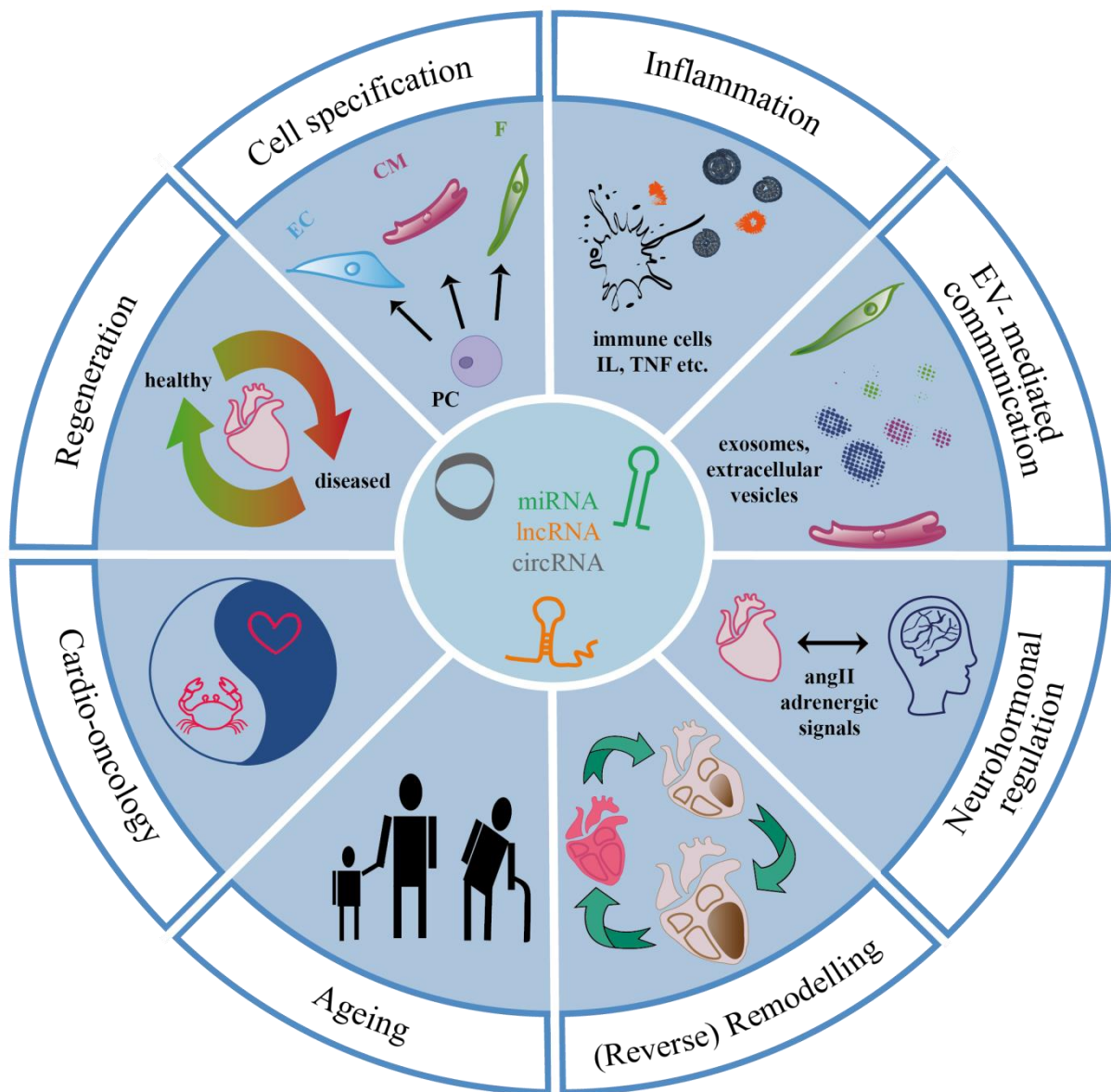


Figure 1: Overview of diverse cardiac processes which underlie ncRNA control. Recent research identified a number of ncRNAs including miRs, lncRNAs and circRNAs in the depicted physiological and pathological processes which are subject of ongoing research in the ESC Working Groups of Myocardial Function and Cellular Biology of the Heart and which will be highlighted in the subsequent chapters. PC-progenitor cells, EC- endothelial cells, CM-cardiomyocytes, F-Fibroblasts, IL-interleukins, TNF-Tumor necrosis factors, EV-extra-cellular vesicles, AngII-AngiotensinII

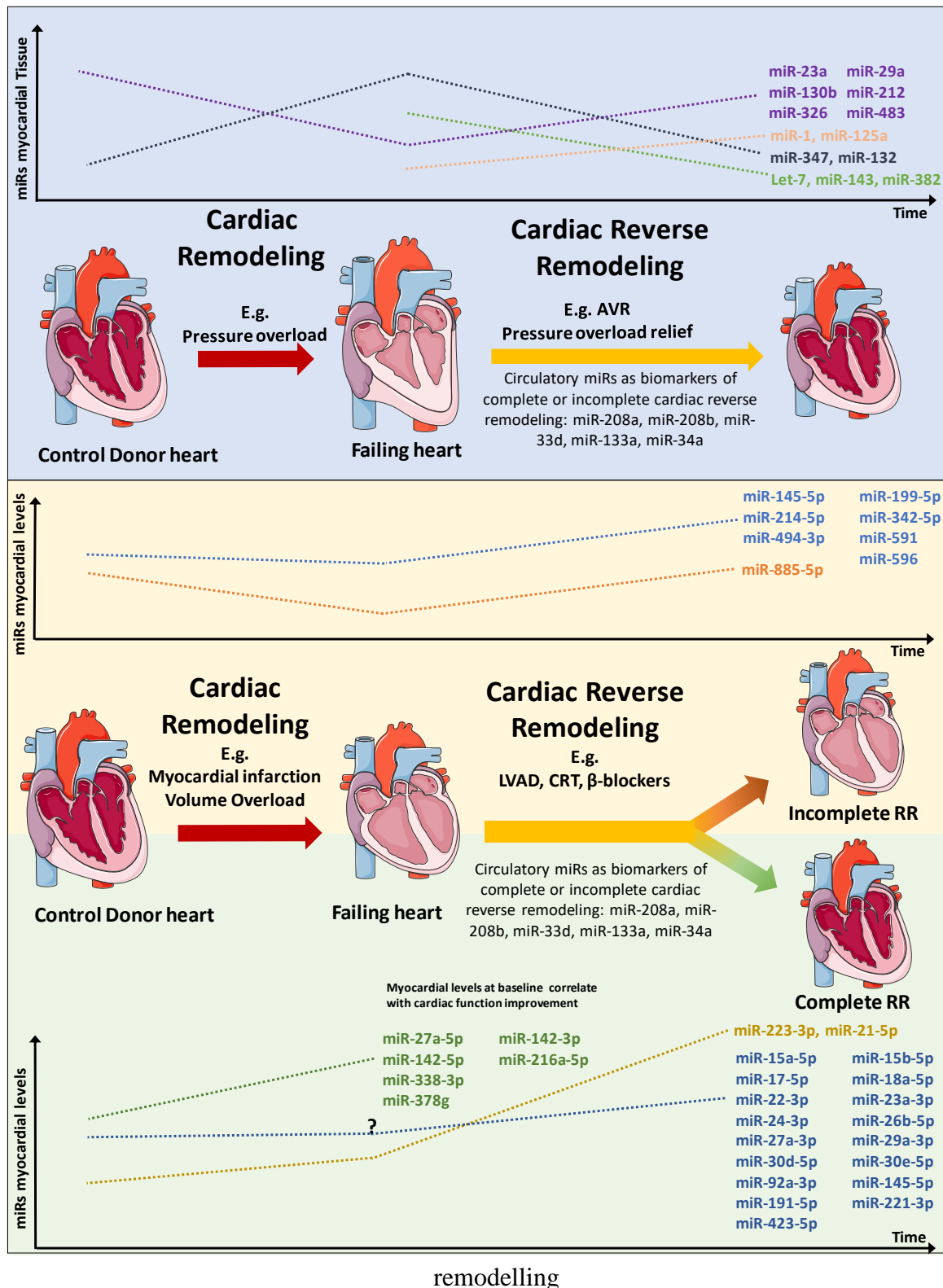


Figure 2: MiRs in cardiac (reverse) remodelling. The left side of the figure represents the changes in miRs levels during ventricular remodelling induced by several pathologies, such as aortic valve stenosis, myocardial infarction, volume overload, etc. The right side depicts changes in miRs levels during reverse remodelling triggered by interventions such as aortic valve replacement, left ventricle assist devices (LVAD) implantation, cardiac resynchronisation

therapy (CRT) or treatment with β -blocker. These changes depend on the extent of myocardial reverse remodelling. Myocardial recovery is considered when RR leads to a total normalization of cardiac function and structure.